

# Diterpenoid alkaloids†

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The lasting attention that researchers have devoted to diterpenoid alkaloids is due to their various bioactivities and toxicities, structural complexity, and intriguing chemistry. From 1998 to the end of 2008, more than 300 new diterpenoid alkaloids were isolated from Nature. This review focuses on their structural relationships, and investigations into their chemical reactions, synthesis, and biological activities. A table that lists the names, plant sources, and structural types is given along with 363 references.

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## 1 Introduction

The important pharmacological activities and structural complexity of the diterpenoid alkaloids have long stimulated scientists' strong interest in their phytochemistry, synthesis, and medicinal chemistry. Since diterpenoid alkaloids were last reviewed in 1999 in this journal by Rahman and Choudhary,<sup>1</sup> they have remained an active area of research. Accordingly, a number of reviews considering various aspects have been published during this period. Hale and Manaviyar have reviewed some important chemical reactions and syntheses of diterpenoid alkaloids, covering the literature from 1985 to early 1998.<sup>2</sup> Ameri summarized the biological activities and mechanisms of action of several diterpenoid alkaloids,<sup>3</sup> while Wang and Xie highlighted the analgesic and anti-inflammatory activities of diterpenoid alkaloids investigated by Chinese researchers.<sup>4</sup> In 1999, Pelletier and coworkers reviewed the past 15 years' work of their research group on the isolation, determination of structures, rearrangement reactions, and spectroscopic studies of diterpenoid alkaloids.<sup>5</sup> Two years later, they compiled an exceedingly useful collection of <sup>13</sup>C NMR and <sup>1</sup>H NMR data and physical constants of C<sub>20</sub>-diterpenoid alkaloids.<sup>6</sup> Bessonova and Saidkhodzhaeva listed the hetisine-type C<sub>20</sub>-diterpenoid alkaloids isolated up to 1998.<sup>7</sup> Wang and Liang have contributed a comprehensive review covering the classification, distribution and occurrence, biosynthesis, spectroscopy, chemical reaction and stereochemistry, and pharmacological

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† Dedicated to the memory of Professor Xiao-Tian Liang (1923–2009).

activity of the C<sub>20</sub>-diterpenoid alkaloids.<sup>8</sup> In this review, a new system of classification for C<sub>20</sub>-diterpenoid alkaloids was proposed, and it was also suggested that the diterpenoid alkaloids be classified into three categories: the C<sub>18</sub>-, C<sub>19</sub>-, and C<sub>20</sub>-diterpenoid alkaloids. In addition, Wang *et al.* have reviewed the advances in various aspects of this field, such as the structures of the C<sub>19</sub>-diterpenoid alkaloids between 1988 and 1998,<sup>9</sup> single-crystal X-ray analyses of the diterpenoid alkaloids up to 2002,<sup>10</sup> and the pharmacological activities of diterpenoid alkaloids between 1984 and 2002.<sup>11</sup> It is also worthwhile to note here that Brimble and coworkers have published an excellent review on the advances in the synthesis of methyllycaconitine and its analogues.<sup>12</sup>

During the review period, in addition to the great progress in the phytochemical investigations on the diterpenoid alkaloids, the following advances should also be pointed out:

1) From the view of pharmacological activities, 8-*O*-azeloyle-14-benzoylaconine, first isolated from *Aconitum karacolicum* by the French scientists Robert and coworkers, exhibits *in vitro* cytotoxicity (IC<sub>50</sub> = 10–20 μM).<sup>13</sup> It was reported by Chinese scientists that talatisamine is a potent and specific blocker of the delayed rectifier K<sup>+</sup> channel in rat hippocampal neurons,<sup>14</sup> and songorine, designated as a novel GABA<sub>A</sub> receptor antagonist in rat brain, can enhance the excitatory synaptic transmission in rat hippocampus.<sup>15</sup>

2) Guan-fu base A has been developed by Jing-Han Liu *et al.* for the therapeutic treatment of arrhythmia in China.<sup>16</sup>

3) Xiao and Wang *et al.* have already systematically summarized the phytotaxonomic characteristic of diterpenoid alkaloids from Chinese *Aconitum*, and their reliable taxonomic character of the genus.<sup>17</sup>

4) Soft-ionization mass spectrometry (ESI-MS, LC-MS-MS, ESI-MS<sup>n</sup>, LC-ESI-MS<sup>n</sup> *etc.*) has been extensively applied to the monitoring of diterpenoid alkaloids from mixed samples. Ohta, Wada, Liu, and Katz are among the scientists who are active in this field.

5) Wang and coworkers have converted a C<sub>19</sub>-diterpenoid alkaloid, deltaline, to the ABC core system of taxanes.<sup>18</sup>

6) The total synthesis of the C<sub>20</sub>-diterpenoid alkaloid nominine has been completed independently by Muratake and Natsume,<sup>19</sup> as well as by Gin and Peese.<sup>20</sup>

This review focuses on the phytochemical investigations, chemical reactions, synthetic studies, and biological activities of diterpenoid alkaloids, covering the literature from 1998 to 2008. In addition, the advances in soft-ionization MS studies and phytotaxonomic researches during the period are also discussed.

## 2 Phytochemical investigations

The system of classification of diterpenoid alkaloids we previously proposed is adopted throughout this review.<sup>8</sup> Accordingly, C<sub>18</sub>-, C<sub>19</sub>-, and C<sub>20</sub>-diterpenoid alkaloids will be discussed by category, and their types and code numbers referred to in the review are shown in Fig. 1. The chemical structures given in the review are predominantly those of new alkaloids. However, structures of known alkaloids are also given when necessary. The structural abbreviations used here



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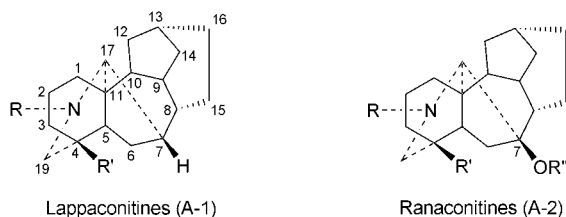
*USA. Appointed as a lecturer in 2001, she was promoted to the position of full professor in 2003 at Sichuan University. Her research interests are focused on natural product-based drug design and drug discovery.*



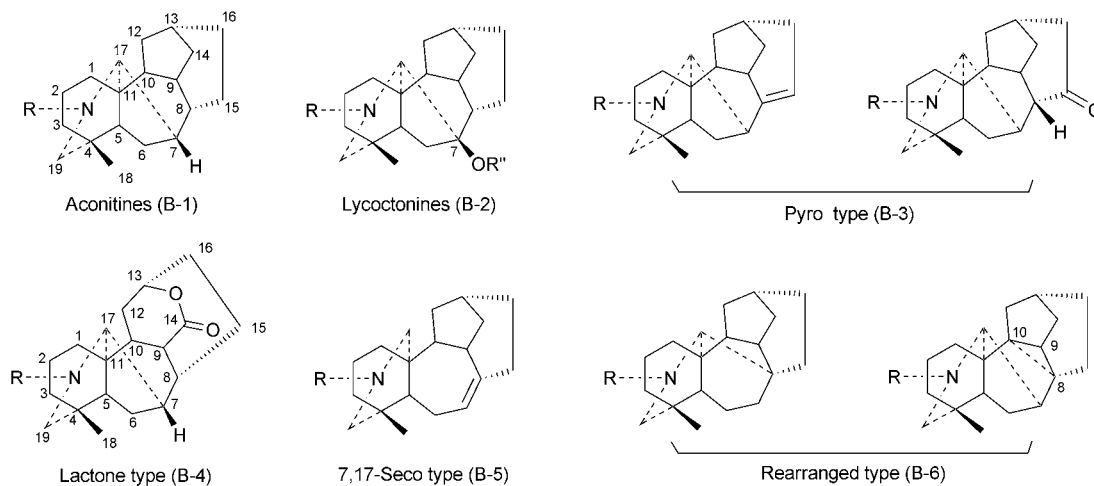
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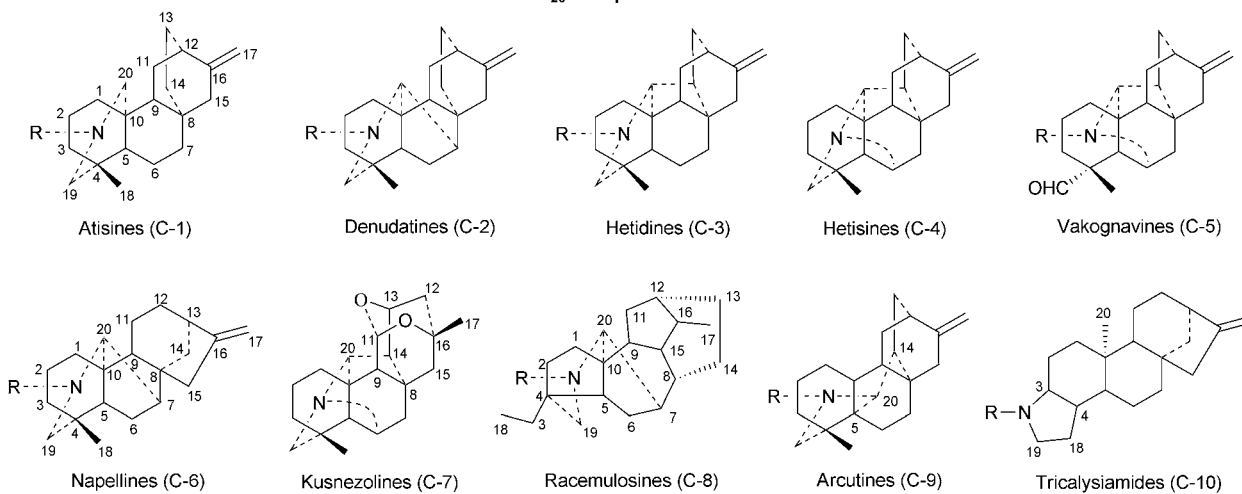
### C<sub>18</sub>-Diterpenoid alkaloids



### C<sub>19</sub>-Diterpenoid alkaloids



### C<sub>20</sub>-Diterpenoid alkaloids



**Fig. 1** Types of diterpenoid alkaloids covered in this review (R and R'' can be hydrogen, alkyl, or acyl groups; R' can be hydrogen, hydroxyl, alkoxy, or acyloxy groups).

are indicated in Fig. 2.‡ Listed in Table 1 are the names, plant sources, types, along with the references of all the new diterpenoid alkaloids reported from 1998 to 2008. Most of the diterpenoid alkaloids during this period were isolated from

the genera *Aconitum*, *Consolida* and *Delphinium* of the Ranunculaceae family, and the genus *Spiraea* of the Rosaceae family, with just a few from other genera.

## 2.1 C<sub>18</sub>-Diterpenoid alkaloids

**2.1.1 Lappaconitines.** The lappaconitine-type C<sub>18</sub>-diterpenoid alkaloids are structurally characterized by the presence of

‡ Throughout this review, the ● symbol has been used at ring junctions in chemical structures to indicate a hydrogen atom that is pointing 'up', i.e. out of the plane of the paper.

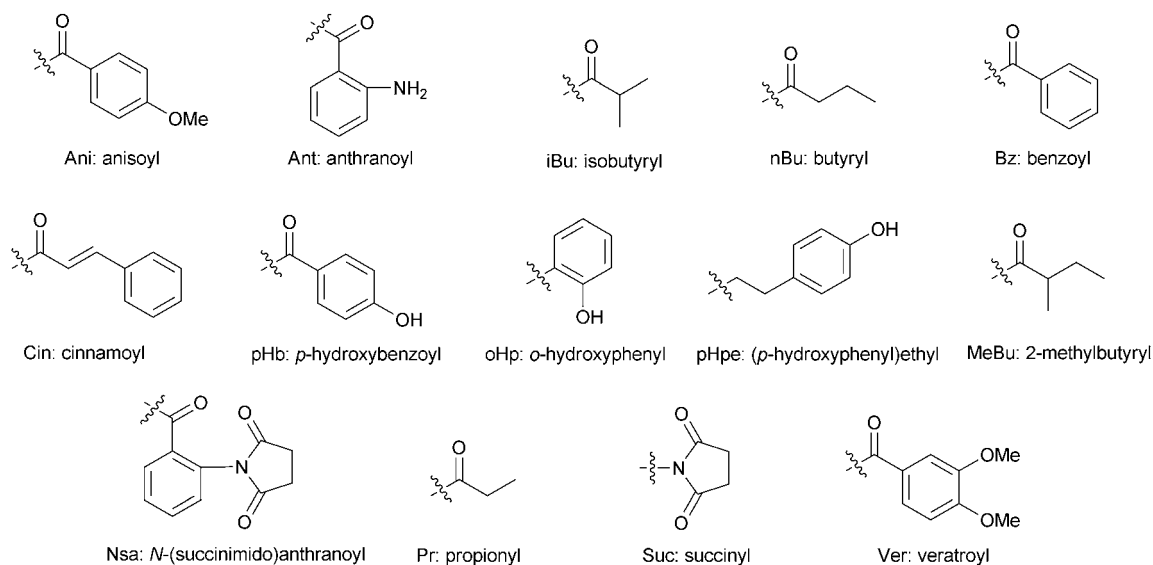


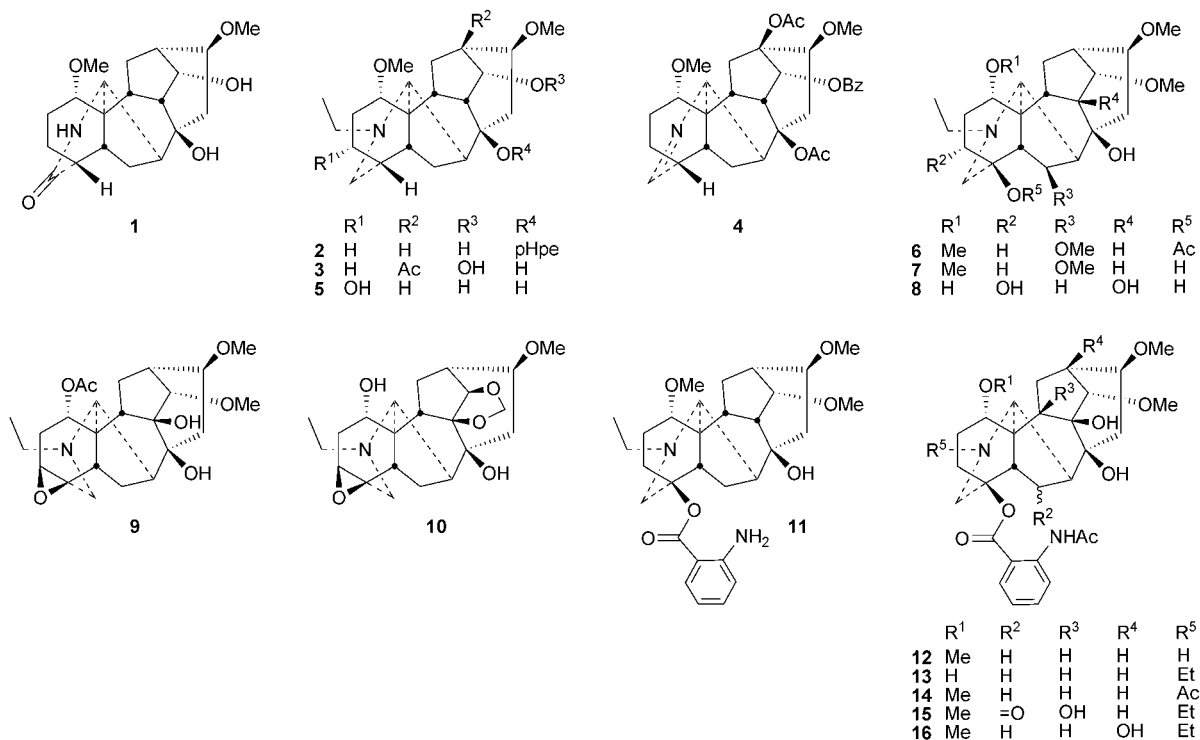
Fig. 2 Structural abbreviations used in this review.

a methine unit at C-7.<sup>21</sup> Sixteen lappaconitine-type alkaloids (**1–16**) were obtained.<sup>22–33</sup> Most of them come from *Aconitum* species, with the exception of alkaloid **11**, which is derived from a *Delphinium* species.

Structurally, piepunendine A (**1**) includes a lactam carbonyl at C-19, while piepunendine B (**2**) is the first example of a C<sub>18</sub>-diterpenoid alkaloid from Nature with a 2-(*p*-hydroxyphenyl)ethoxy group at C-8.<sup>22</sup> Delavaconitine G (**4**) possesses an azomethine group between *N* and C-19.<sup>23</sup> Akiradin (**9**) and

kiridine (**10**) possess a C-3,C-4 epoxide group.<sup>28,29</sup> In addition, kiridine (**10**), whose structure was established by X-ray crystallographic analysis, is also the first example of a diterpenoid alkaloid that contains a 9,14-methylenedioxy group. Lappaconitines **12–16** possess an *N*-acetylanthranoyloxy substituent at C-18.<sup>31–33</sup>

**2.1.2 Ranaconitines.** The ranaconitine-type C<sub>18</sub>-diterpenoid alkaloids, featuring an oxygen-containing functionality at C-7,



**Table 1** New diterpenoid alkaloids isolated from 1998 to 2008

Source	Diterpenoid alkaloid	Type <sup>a</sup>	Ref.	
<i>Aconitum arcuatum</i>	Arcutin(e) (346)	C-9	220	
	Arcutinine (347)	C-9	221	
<i>Aconitum balfourii</i>	9-Hydroxysenbushine A (112)	B-1	84	
<i>Aconitum barbatum</i> var. <i>hispidum</i>	11 $\alpha$ -Hydroxylepenine (294)	C-2	182	
<i>Aconitum brunneum</i>	3 $\alpha$ -Hydroxy-12- <i>epi</i> -napelline (338)	C-6	210	
<i>Aconitum bulleyanum</i>	Talatisamine 8-acetyl-14- <i>p</i> -methoxybenzoate (54)	B-1	57	
	Talatisamine 14- <i>p</i> -methoxybenzoate (55)	B-1	57	
<i>Aconitum carmichaeli</i>	14- <i>O</i> -Anisoyleone (90)	B-1	71	
	8- <i>O</i> -Cinnamoylneoline (91)	B-1	72	
	14- <i>O</i> -Cinnamoylneoline (89)	B-1	71	
	Lipo-14- <i>O</i> -anisoyleone (134)	B-1	71	
	Lipoforesaconitine (135)	B-1	102	
	14- <i>O</i> -Veratroylneoline (88)	B-1	71	
<i>Aconitum cochleare</i>	Acochlearine (288)	C-2	180	
	Acoleareine (116)	B-1	88	
	Aconitileareine (94)	B-1	74	
	Cochleareine (262)	C-1	88	
<i>Aconitum coreanum</i>	<i>N</i> -Deethylmethyllycaconitine (211)	B-2	74	
	13-Acetyl-14-hydroxy-2-propionylhetisine (309)	C-4	191	
	Guan-fu base K (343)	C-7	218	
	Guan-fu base P (310)	C-4	192	
	Guan-fu base Q (312)	C-4	194	
	Guan-fu base R (311)	C-4	193	
	Guan-fu base S (315)	C-4	196	
	Guan-fu base T (313)	C-4	195	
	Guan-fu base U (314)	C-4	195	
	<i>Aconitum delavayi</i>	Delavaconitine F (3)	A-1	23
Delavaconitine G (4)		A-1	23	
<i>Aconitum episcopale</i>	Liaconitine A (81)	B-1	68	
	Liaconitine B (82)	B-1	68	
	Liaconitine C (83)	B-1	68	
<i>Aconitum excelsum</i>	Secoyunaconitine (251)	B-5	157	
	6-Demethyldeleoline (150)	B-2	35	
	Exceconidine (17)	A-2	34	
	Excecoitine (150)	B-2	34	
	8-Methyl-10-hydroxyllycoctonine (168)	B-2	35	
	8-Methyllycoctonine (169)	B-2	35	
	6-Methylumbrofine (17)	A-2	35	
<i>Aconitum falconeri</i>	Faleoconitine (104)	B-1	76	
	3'-Methoxyacoforestinine (98)	B-1	76	
<i>Aconitum geniculatum</i>	Geniculatine A (87)	B-1	52	
	Geniculatine B (88)	B-1	52	
	Geniculatine C (52)	B-1	52	
	Geniculatine D (47)	B-1	52	
	Geniculine (99)	B-1	77	
	Genicunine A (40)	B-1	49	
	Genicunine B (41)	B-1	49	
	Genicunine C (50)	B-1	49	
	Habaenine A (146)	B-1	81	
<i>Aconitum habaense</i>	Habaenine B (108)	B-1	81	
	Habaenine C (147)	B-1	106	
	Hemsleyatine (130)	B-1	99	
<i>Aconitum hemsleyanum</i>	Atropurpursine (103)	B-1	79	
	3-Hydroxyfranchetine (246)	B-5	79	
<i>Aconitum hemsleyanum</i> var. <i>atropurpureum</i>	Circinadine A (63)	B-1	62	
<i>Aconitum hemsleyanum</i> var. <i>circinatum</i>	Circinadine B (64)	B-1	62	
	Circinasine A (68)	B-1	61	
	Circinasine B (69)	B-1	61	
	Circinasine C (65)	B-1	61	
	Circinasine D (61)	B-1	61	
	Circinasine E (62)	B-1	61	
	Circinasine F (70)	B-1	61	
	Circinasine G (140)	B-1	61	
	Hemsleyanine A (66)	B-1	63	
	Hemsleyanine B (67)	B-1	63	
	Hemsleyanine C (59)	B-1	60	
	Hemsleyanine D (60)	B-1	60	
	<i>Aconitum hemsleyanum</i> var. <i>leueanthus</i>	Leueandine (245)	B-5	142
		Leueantine A (100)	B-1	56
Leueantine B (79)		B-1	56	
	Leueantine C (53)	B-1	56	

Table 1 (Contd.)

Source	Diterpenoid alkaloid	Type <sup>a</sup>	Ref.
<i>Aconitum hemisleyanum</i> var. <i>pengzhouense</i>	Leueantine D (110)	B-1	56
	8-Deacetylungpaconitine (97)	B-1	75
	13-Deoxyludaconitine (96)	B-1	75
	6- <i>epi</i> -Forsticine (109)	B-1	82
	Pengshenine A (136)	B-1	103
<i>Aconitum heterophyllum</i>	Pengshenine B (138)	B-1	103
	6-Dehydroacetylsepaconitine (15)	A-1	33
<i>Aconitum jaluense</i>	13-Hydroxylappaconitine (16)	A-1	33
<i>Aconitum karacolicum</i>	Jaluenine (316)	C-4	197
<i>Aconitum karakolicum</i>	8- <i>O</i> -Azelo-14-benzoylaconine (133)	B-1	13
	Secokaraconitine (250)	B-5	155,156
<i>Aconitum kirinense</i>	Acofamine A (119)	B-1	90
	Acofamine B (120)	B-1	90
<i>Aconitum kongboense</i>	Akiradin (9)	A-1	28
	Akiramidine (7)	A-1	26
	Akiramine (6)	A-1	25
	Kiridine (10)	A-1	29
	Kirinine B (295)	C-2	183
	Kirinine C (296)	C-2	183
	Kongboendine (244)	B-5	152
<i>Aconitum kusnezoffii</i>	Kongboentine A (132)	B-1	83
	Kongboentine B (111)	B-1	83
<i>Aconitum laeve</i>	3-Acetylaconifine (124)	B-1	94
	3-Acetylmesaconitine (125)	B-1	94
	Acsonine (247)	B-5	154
	Beiwudine (247)	B-5	153
	Beiwucine (122)	B-1	92
	Beiwusine A (257)	C-1	164
	Beiwusine B (258)	C-1	164
<i>Aconitum liljestrandii</i>	6- <i>epi</i> -Forsticine (109)	B-1	82
	Swatinine (161)	B-2	113
<i>Aconitum leave</i>	<i>N</i> -Deethyltalatisamine (56)	B-1	58
	Liljestrandinine (58)	B-1	58
	Liljestrandisine (57)	B-1	59
<i>Aconitum lycoctonum</i>	<i>N</i> -Deethyllycaconitine- <i>N</i> -aldehyde (203)	B-2	135
	14-Demethyllycaconitine (204)	B-2	135
	8-Methyllycaconitine (222)	B-2	135
	6- <i>O</i> -Acetyldemethylenedelcorine (152)	B-2	108
<i>Aconitum macrorhynchum</i>	6- <i>O</i> -Acetyl-14- <i>O</i> -methyldephinifoline (151)	B-2	108
	Macrorhynine A (142)	B-1	104
<i>Aconitum manshuricum</i>	Macrorhynine B (143)	B-1	104
<i>Aconitum nagarum</i>	Manshuritine (127)	B-1	97
<i>Aconitum nagarum</i> var. <i>lasiandrum</i>	10-Dehydroxyflavaconitine (126)	B-1	96
	13-Hydroxyfranchetine (249)	B-5	96
	14-Benzoylsachaconitine (33)	B-1	43
	<i>N</i> -Deethyl- <i>N</i> -methyl-12- <i>epi</i> -napelline (339)	C-6	179
	16,17-Dihydro-12 $\beta$ ,16 $\beta$ -epoxynapelline (340)	C-6	179
	11- <i>epi</i> -16 $\alpha$ ,17-Dihydroxylepenine (287)	C-2	179
	Francheline (248)	B-5	100
	Lasianine (131)	B-1	100
	Lasiansine (113)	B-1	85
	Nagadine (138)	B-1	43
<i>Aconitum napellus</i>	Merckonine (144)	B-1	105
<i>Aconitum nasutum</i>	Trabzonine (304)	C-3	187
<i>Aconitum naviculare</i>	Naviculine A (307)	C-3	189
	Naviculine B (306)	C-3	189
<i>Aconitum nemorum</i>	Navirine (308)	C-3	190
<i>Aconitum orochryseum</i>	1- <i>epi</i> -Deacetylaconitine (118)	B-1	89
	2- <i>O</i> -Acetyl-7 $\alpha$ -hydroxyorochrine (319)	C-4	198
	2- <i>O</i> -Acetylorochrine (318)	C-4	198
	Orochrine (317)	C-4	198
<i>Aconitum ouvrardianum</i>	Ouvrardiandine A (269)	C-1	78
	Ouvrardiandine B (270)	C-1	78
	Ouvrardiantine (102)	B-1	78
<i>Aconitum piepunense</i>	18-Acetylcammaconine (49)	B-1	54
	Piepunendine A (1)	A-1	22
	Piepunendine B (2)	A-1	22
	Piepunensine A (145)	B-1	54
<i>Aconitum pseudo-laeve</i> var. <i>erectum</i>	18- <i>O</i> -2-(2-Methyl-4-oxo-4 <i>H</i> -quinazoline-3-yl)benzoyllycoctonine (221)	B-2	65

Table 1 (Contd.)

Source	Diterpenoid alkaloid	Type <sup>a</sup>	Ref.
	14- <i>O</i> -Acetyl-8- <i>O</i> -methyl-18- <i>O</i> -2-(2-methyl-4-oxo-4 <i>H</i> -quinazoline-3-yl)benzoylcammaconine (77)	B-1	65
<i>Aconitum racemulosum</i>	Racemulodine (301)	C-3	185
var. <i>pengzhouense</i>	Racemuloline A (149)	B-2	50
	Racemuloline B (42)	B-1	50
	Racemulosine (345)	C-8	219
<i>Aconitum septentrionale</i>	Racemulotone (341)	C-6	211
	Acoseptine (252)	B-6	158
	Anhydrolycaconitine (253)	B-6	159
	Septonine (254)	B-6	162
	Septontrionine (255)	B-6	162
<i>Aconitum sinomontanum</i>	Sinaconitine A (31)	A-2	32
	Sinaconitine B (14)	A-1	32
	Sinomontanine A (12)	A-1	31
	Sinomontanine B (13)	A-1	31
	Sinomontanine C (75)	B-1	31
	Sinomontanine D (23)	A-2	27
	Sinomontanine E (8)	A-1	27
	Sinomontanine F (29)	A-2	41
	Sinomontanine G (28)	A-2	41
	Sinomontanine H (30)	A-2	41
	Sinomontanine I (205)	B-2	41
	Sinomontanitine A (45)	B-1	31
	Sinomontanitine B (46)	B-1	31
<i>Aconitum soongoricum</i>	12-Acetyl-12- <i>epi</i> -napelline (337)	C-6	209
<i>Aconitum spicatum</i>	Spicatine A (123)	B-1	93
	Spicatine B (126)	B-1	93
<i>Aconitum sungpanense</i> var. <i>leucanthum</i>	Leucanthumsine A (84)	B-1	69
	Leucanthumsine B (85)	B-1	69
	Leucanthumsine C (95)	B-1	69
	Leucanthumsine D (141)	B-1	69
	Leucanthumsine E (107)	B-1	69
<i>Aconitum taipaicum</i>	Isodelelatine (179)	B-2	122
<i>Aconitum tanguticum</i>	Tangutimine (305)	C-3	188
	Tangutisine A (332)	C-5	205
	Tangutisine B (333)	C-5	206
<i>Aconitum toxicum</i>	Acotoxicine (5)	A-1	24
	Acotoxinine (92)	B-1	73
<i>Aconitum transsectum</i>	<i>N</i> -Deethylchasmanine (80)	B-1	67
	8- <i>O</i> -Ethylunaconitine (101)	B-1	67
	Transconitine D (137)	B-1	67
	Transconitine E (105)	B-1	67
<i>Aconitum tuberosum</i>	Tuberaconitine (128)	B-1	98
	Tuberanine (125)	B-1	95
	Tubermesaconitine (129)	B-1	98
<i>Aconitum variegatum</i>	14-Acetylgenicinunine B (34)	B-1	44
	<i>N</i> -Deethyl- <i>N</i> -19-didehydrosachaconitine (139)	B-1	44
	8-Ethoxysachaconitine (37)	B-1	44
	16 $\beta$ -Hydroxycardiopetaline (44)	B-1	44
	<i>N</i> -Ethyl-1 $\alpha$ -hydroxy-17-veratrolydictizine (291)	C-2	44
	Variegatine (300)	C-3	44
	15-Veratroyl-17-acetyldictizine (289)	C-2	44
	15-Veratroyl-17-acetyl-19-oxodictizine (292)	C-2	44
	15-Veratrolydictizine (290)	C-2	44
<i>Aconitum vilmorinianum</i>	Vilmoraconitine (256)	B-6	163
<i>Aconitum vulparia</i>	Acovulparine (233)	B-2	145
	Vulparine (219)	B-2	140
<i>Aconitum</i> sp. (cultivated in Korea)	8- <i>O</i> -Methylhypoconine (121)	B-1	91
Ascidian <i>Lissoclinum</i> spp.	Haterumaimide A (354)	M	225
	Haterumaimide B (355)	M	225
	Haterumaimide C (356)	M	225
	Haterumaimide D (357)	M	225
	Haterumaimide E (358)	M	225
	Haterumaimide F (359)	M	226
	Haterumaimide G (360)	M	226
	Haterumaimide H (361)	M	226
	Haterumaimide I (362)	M	226
	Haterumaimide J (363)	M	227
	Haterumaimide K (364)	M	227

Table 1 (Contd.)

Source	Diterpenoid alkaloid	Type <sup>a</sup>	Ref.
<i>Chamaecyparis obtusa</i> cv. <i>tetragon</i>	Chamobtusin A (353)	M	224
<i>Clitocybe concava</i>	Concavine (352)	M	223
<i>Consolida ambigua</i>	13-Acetylvakhmatine (324)	C-4	199
<i>Consolida armeniaca</i>	Consolarine (148)	B-2	107
<i>Consolida glandulosa</i>	9- <i>O</i> -Acetylglanduline (325)	C-4	200
	9-Deoxyglanduline (328)	C-4	200
	11,13- <i>O</i> -Diacetylglanduline (326)	C-4	200
	Glandulosine (327)	C-4	200
<i>Consolida hohenackeri</i>	Consolinine (86)	B-1	70
	Hoheconsoline (93)	B-1	70
<i>Consolida oliveriana</i>	7 $\alpha$ -Hydroxycossonidine (329)	C-4	53
	8- <i>O</i> -Methylcolumbianine (48)	B-1	53
	Olividine (237)	B-2	53
	Olivimine (232)	B-2	53
<i>Consolida orientalis</i>	14- <i>O</i> -Acetyltakaosamine (162)	B-2	114
	18- <i>O</i> -Benzoyl-14- <i>O</i> -deacetyl-18- <i>O</i> -demethylpubescenine (174)	B-2	39
	14- <i>O</i> -Benzoyltakaosamine (163)	B-2	39
	Consorientaline (260)	C-1	167
	14-Deacetyl-18-demethylpubescenine (175)	B-2	118
	14- <i>O</i> -Deacetylpubescenine (172)	B-2	39
	Dehydrodeltatsine (226)	B-2	114
	14- <i>O</i> -Demethyldeboxine (26)	A-2	39
	18-Demethoxypubescenine (171)	B-2	114
	18-Demethylpubescenine (170)	B-2	117
	1- <i>O</i> -Demethyltricornine (164)	B-2	39
	1- <i>O</i> ,19-Didehydrotakaosamine (227)	B-2	39
	8- <i>O</i> -Methylconsolarine (173)	B-2	39
<i>Consolida scleroclada</i>	Willipelletierine (293)	C-2	181
<i>Delphinium alpinum</i>	Alpinine (225)	B-2	143
<i>Delphinium anthriscifolium</i> var. <i>savatieri</i>	Anthriscifolcine A (18)	A-2	36
	Anthriscifolcine B (19)	A-2	36
	Anthriscifolcine C (20)	A-2	36
	Anthriscifolcine D (21)	A-2	36
	Anthriscifolcine E (22)	A-2	36
<i>Delphinium bonvalotii</i>	Bonvalotidine A (196)	B-2	132
	Bonvalotidine B (197)	B-2	132
	Bonvalotidine C (198)	B-2	132
<i>Delphinium brunonianum</i>	Delbruninol (181)	B-2	124
<i>Delphinium buschianum</i>	Budelphine (240)	B-2	147
<i>Delphinium campylocentrum</i>	Campylocine (230)	B-2	128
	Campylotine (185)	B-2	128
<i>Delphinium carduchorum</i>	Carduchoron (298)	C-3	184
	Delcarduchol (297)	C-3	184
<i>Delphinium corymbosum</i>	Delcorinine (180)	B-2	123
<i>Delphinium crispulum</i>	Crispulidine (36)	B-1	30
	Delphicrispuline (11)	A-1	30
<i>Delphinium chrysotrichum</i>	Delphatisine A (267)	C-1	172
	Delphatisine B (268)	C-1	172
<i>Delphinium cuneatum</i>	16-Demethoxymethyllycaconitine (210)	B-2	137
	16-Demethoxydelavaine (218)	B-2	139
<i>Delphinium cyphoplectrum</i>	Cyphoplectine (74)	B-1	64
<i>Delphinium davidii</i>	Davidisine A (165)	B-2	115
	Davidisine B (166)	B-2	115
<i>Delphinium dissectum</i>	10-Hydroxymethyllycaconitine (212)	B-2	112
<i>Delphinium excelsum</i>	10-Hydroxymethyllycaconitine (212)	B-2	112
	10-Hydroxynudicaulidine (160)	B-2	112
	18- <i>O</i> -Methyldeleterine (159)	B-2	112
<i>Delphinium fangshanense</i>	16-Demethyldelesoline (153)	B-2	109
<i>Delphinium giraldii</i>	Giraldine A (154)	B-2	110
	Giraldine B (155)	B-2	110
	Giraldine C (156)	B-2	110
	Giraldine D (176)	B-2	111
	Giraldine E (157)	B-2	111
	Giraldine F (158)	B-2	111
	Giraldine G (216)	B-2	51
	Giraldine H (217)	B-2	51
	Giraldine I (43)	B-1	51
<i>Delphinium gracile</i>	Delphigraciline (321)	C-4	47



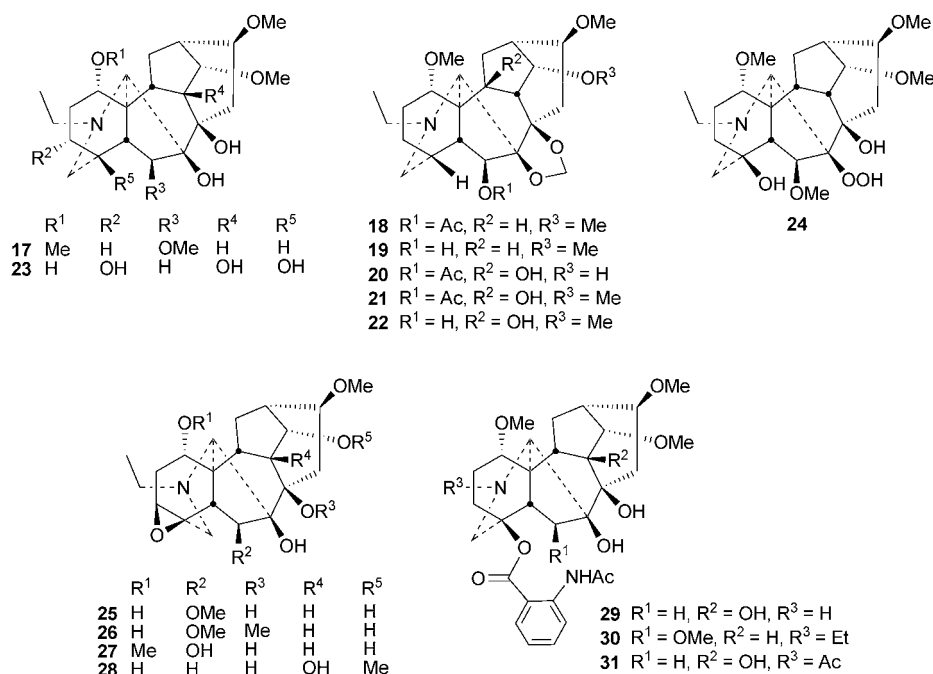
Table 1 (Contd.)

Source	Diterpenoid alkaloid	Type <sup>a</sup>	Ref.
	14-Hydroxyhetisinone <i>N</i> -oxide (322)	C-4	47
	8-Methylkarakoline (38)	B-1	47
<i>Delphinium laxicymosum</i> var. <i>pilostachyum</i>	Laxicymine (228)	B-2	125
	Laxicyminine (229)	B-2	125
	Laxicymisine (182)	B-2	125
<i>Delphinium linearilobum</i>	Linearilin (24)	A-2	37
	Linearilobin (76)	B-1	37
<i>Delphinium nordhagenii</i>	Nordhagenine A (193)	B-2	131
	Nordhagenine B (194)	B-2	131
	Nordhagenine C (195)	B-2	131
<i>Delphinium nuttallianum</i>	14-Acetylbearline (207)	B-2	136
	Bearline (209)	B-2	136
	16-Deacetylgeyerline (208)	B-2	136
<i>Delphinium orthocentrum</i>	Deacetylswinanine A (184)	B-2	127
	Orthocentrine (236)	B-2	127
<i>Delphinium pentagynum</i>	2-Dehydrodeacetylheterophylloidine (299)	C-3	146
	14-Demethyl-14-acetylanhweidelphinine (235)	B-2	146
	14-Demethyl-14-isobutyrylanhweidelphinine (234)	B-2	146
<i>Delphinium poltoratskii</i>	Delpoline (32)	B-1	42
<i>Delphinium potaninii</i>	Potansine F (223)	B-2	142
	Potansine G (224)	B-2	142
<i>Delphinium potaninii</i> var. <i>jiufengshanense</i>	Jiufengdine (200)	B-2	133
	Jiufengsine (177)	B-2	119
	Jiufengtine (199)	B-2	133
<i>Delphinium pyramidale</i>	8-Acetylcondelphine (51)	B-1	55
<i>Delphinium roylei</i>	Royleinine (106)	B-1	80
<i>Delphinium scabriflorum</i>	13-(2-Methylbutyryl)azatine (284)	C-1	178
<i>Delphinium shawurense</i>	Shawurensine (220)	B-2	141
<i>Delphinium siwanense</i> var. <i>leptogen</i>	Siwanine A (186)	B-2	129
	Siwanine B (187)	B-2	129
	Siwanine C (188)	B-2	129
	Siwanine D (189)	B-2	129
<i>Delphinium souliei</i>	Soulidine (191)	B-2	130
	Souline A (114)	B-1	86
	Souline B (243)	B-4	86
	Souline C (190)	B-2	48
	Souline D (39)	B-1	48
	Souline E (35)	B-1	45,46
	Souline F (320)	C-4	45,46
<i>Delphinium stapeliosum</i>	14-Deacetyl-14-isobutyrylajadine (202)	B-2	38
	14-Deacetyl-14-isobutyrylnudicauline (206)	B-2	38
	14-Demethyltuguaconitine (25)	A-2	38
<i>Delphinium staphisagria</i>	22- <i>O</i> -Acetyl-19-oxodihydroatisine (286)	C-1	177
	Isoazatine (283)	C-1	177
	19-Oxodihydroatisine (285)	C-1	177
<i>Delphinium tatsienense</i> var. <i>chinghaiense</i>	Tatsienine V (178)	B-2	121
<i>Delphinium tiantaishanense</i>	Tiantaishandine (323)	C-4	40
	Tiantaishanmine (238)	B-2	40
	Tiantaishannine (192)	B-2	40
	Tiantaishansine (27)	A-2	40
<i>Delphinium tongolense</i>	Tongolenine C (231)	B-2	144
	Tongolenine D (239)	B-2	144
<i>Delphinium trifoliolatum</i>	Trifoliolasine A (201)	B-2	134
	Trifoliolasine B (214)	B-2	134
	Trifoliolasine C (215)	B-2	134
	Trifoliolasine D (334)	C-5	207
	Trifoliolasine E (335)	C-5	207
	Trifoliolasine F (336)	C-5	207
<i>Delphinium uncinatum</i>	14-Acetylchasmantine (78)	B-1	66
	Uncinatine (261)	C-1	168
<i>Delphinium uralense</i>	Uraline (213)	B-2	138
	Uraphine (183)	B-2	126
<i>Delphinium virgatum</i>	<i>N</i> -Deethylperegrine alcohol (115)	B-1	87
<i>Spiraea formosana</i>	Spiraeaine A (272)	C-1	173
<i>Spiraea fritschiana</i> var. <i>parvifolia</i>	Spirafine II (302)	C-3	186
	Spirafine III (303)	C-3	186
<i>Spiraea japonica</i> var. <i>acuta</i>	Spiramide (265)	C-1	166,171
	Spiramine P (275)	C-1	175
	Spiramine Q (276)	C-1	175

**Table 1** (Contd.)

Source	Diterpenoid alkaloid	Type <sup>a</sup>	Ref.	
<i>Spiraea japonica</i> var. <i>fortunei</i>	Spiramine T (278)	C-1	175	
	Spiramine U (277)	C-1	175	
	Spiramine W (279)	C-1	176	
	Spiramine X (273)	C-1	174	
	Spiramine Y (274)	C-1	174	
	Spiramine Z (281)	C-1	174	
	Spiratine A (259)	C-1	165,166	
	Spiratine B (280)	C-1	165,166	
	6-Hydroxylspiraquine (331)	C-4	201	
	Spiraquine (330)	C-4	201	
	<i>Spiraea japonica</i> var. <i>ovalifolia</i>	Deacetylspiramine F (271)	C-1	169,170
		15-Deacetylspiramine S (266)	C-1	169,170
		19- <i>O</i> -Deethylspiramine N (282)	C-1	169,170
		Spiramidine A/spiramine Z-2 (263)	C-1	169,170
Spiramidine B/spiramine Z-3 (264)		C-1	169,170	
<i>Tricalysia dubia</i>	Tricalysiamide A (348)	C-10	222	
	Tricalysiamide B (349)	C-10	222	
	Tricalysiamide C (350)	C-10	222	
	Tricalysiamide D (351)	C-10	222	

<sup>a</sup> M: Miscellaneous.



comprise 15 new members (17–31).<sup>27,32,34–41</sup> All of these alkaloids come from the genera *Aconitum*, *Consolida* and *Delphinium*.

The alkaloid linearilin (24) has a rare hydroperoxyl group at C-7, which was confirmed by an iodine test.<sup>37</sup> There are four ranaconitine-type alkaloids that possess a C-3,C-4 epoxide unit, including 14-demethyltugaconitine (25),<sup>38</sup> 14-*O*-demethyldeboxine (26),<sup>39</sup> tiantaishansine (27),<sup>40</sup> and sinomontanine G (28).<sup>41</sup> The structure of sinaconitine A (31) was confirmed by X-ray crystallographic analysis.<sup>32</sup>

## 2.2 C<sub>19</sub>-Diterpenoid alkaloids

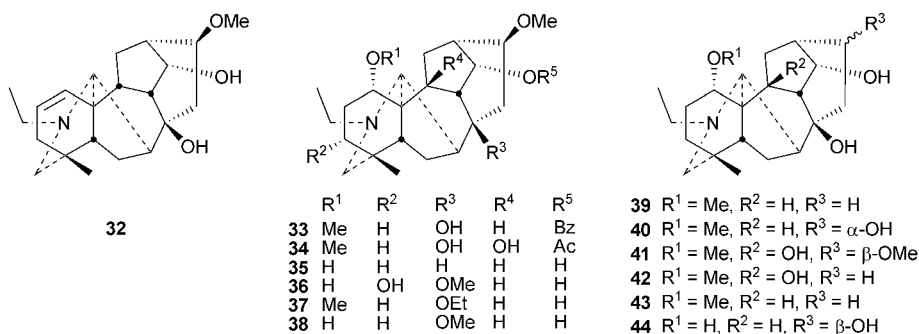
### 2.2.1 Aconitines.

Aconitine-type C<sub>19</sub>-diterpenoid alkaloids have no oxygen-containing functionality at C-7. Based on the

nitrogen patterns, they may be subdivided into the following four subtypes: the amine, the *N,O*-mixed acetal, the imine, and the amide subtypes. The majority of these alkaloids are of the amine subtype. Here we will discuss the amine subtype in various groups based on whether oxygenated functionalities are attached at C-6 or C-15.

Alkaloids 32–77 lack oxygenated groups at both C-6 and C-15.<sup>30,31,37,42–54,57,58,60–65</sup> Delpoline (32) is the first aconitine-type alkaloid that has a  $\Delta^{1,2}$  double bond.<sup>42</sup> Alkaloids 33–44 have a methyl group at C-4, and 36–38 contain an alkoxy group at C-8. Alkaloids 39, 42 and 43 have no oxygen-containing group at C-16; unusually, alkaloids 40 and 44 have a hydroxyl group at C-16.<sup>30,43–51</sup>

Sinomontanitines A (45) and B (46) possess an *N*-(succinimido)anthranoyl moiety at C-8, as well as a hydroxyl group at

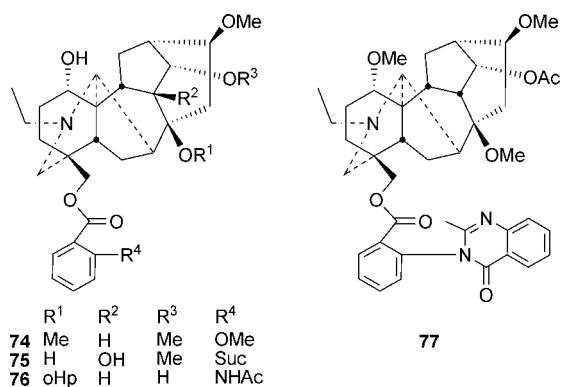


C-18.<sup>31</sup> Geniculatine D (**47**) and 8-*O*-methylcolumbianine (**48**) have a hydroxyl group at C-18; in contrast, the functionality at C-18 of 18-acetylcammaconine (**49**) is an acetate.<sup>52–54</sup>

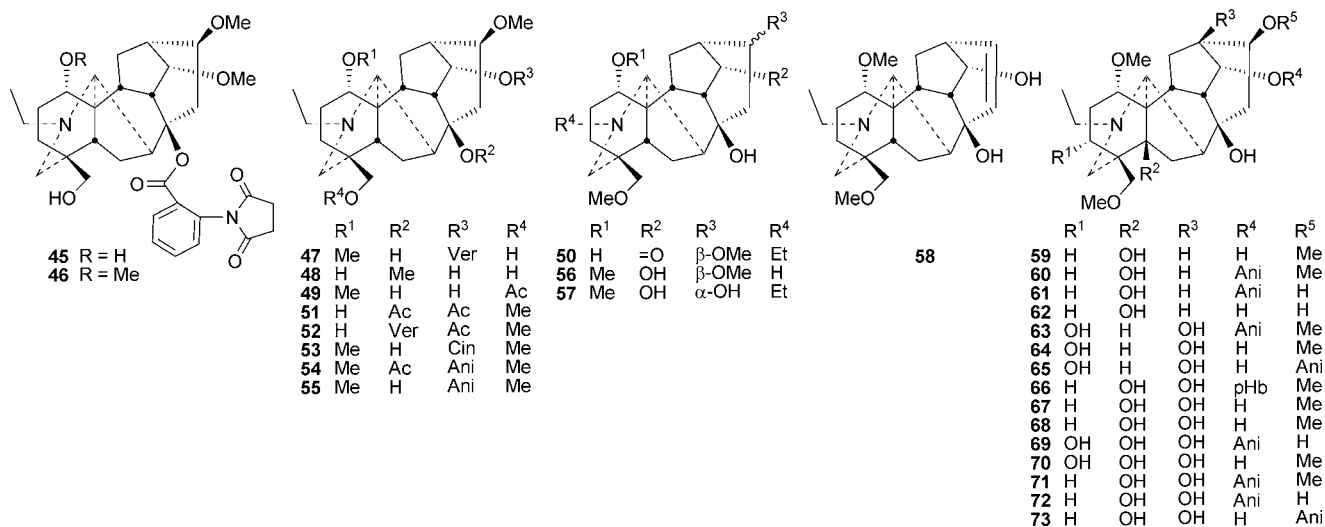
Alkaloids **50–73** all have a methoxyl group at C-18. All of them were isolated from the genus *Aconitum* with an exception of 8-acetylcondelphine (**51**), which was obtained from *Delphinium pyramidale*. Among these alkaloids, genicunine C (**50**) has a carbonyl group at C-14,<sup>49</sup> while liljestrandinine (**58**) possesses a  $\Delta^{15,16}$  double bond.<sup>58</sup> Investigation on the roots of *Aconitum hemsleyanum* var. *circinatum* by Wang *et al.* led to the isolation of twelve new aconitine-type alkaloids (**59–70**), as well as the revision of the structures of hemsleyadine, hemsleyanisine and iso-hemsleyanisine to the structures shown (**71–73**).<sup>60–63</sup> The structures of **68**, **70** and **71** were confirmed by single-crystal X-ray analysis. Of special interest are the alkaloids **59–62** and **66–70**, which possess an uncommon hydroxyl group at C-5.

Four new aconitine-type alkaloids (**74–77**) have benzoate or anthranilate derivatives at C-18. Sinomontanine C (**75**) contains a C-18 *N*-(succinimido)anthranoyl moiety,<sup>31</sup> and linearilobin (**76**) has an *N*-acetyl anthranoyl group at C-18 as well as a rare catechol unit at C-8.<sup>37</sup> Further investigation on *Aconitum pseudo-laeve* var. *erectum* resulted in the isolation of two unique C<sub>19</sub>-diterpenoid alkaloids, 14-*O*-acetyl-8-*O*-methyl-18-*O*-2-(2-methyl-4-oxo-4*H*-quinazoline-3-yl)benzoylcammaconine (**77**) and 18-*O*-2-(2-methyl-4-oxo-4*H*-quinazoline-3-yl)benzoyllycoctonine (**221**). Interestingly, it was found for the first time that the 2-(2-methyl-4-

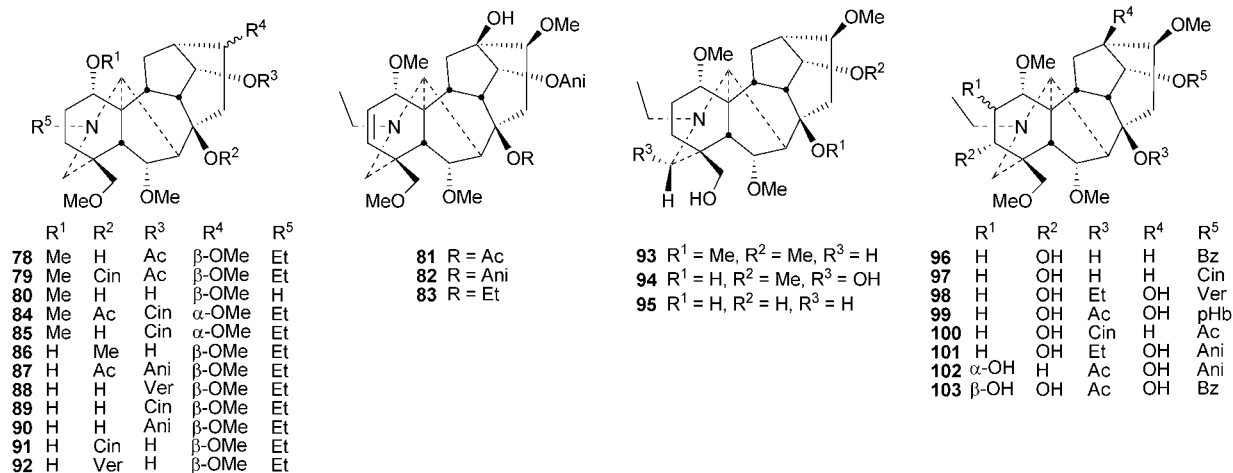
oxo-4*H*-quinazoline-3-yl)benzoate moiety could exist as a substituent of diterpenoid alkaloids.<sup>65</sup>



The alkaloids **78–116** possess an oxygenated group at C-6, most of which are configured in the  $\alpha$ -orientation.<sup>52,56,66–68,71,72,74,76,77,79,82</sup> Leueantine B (**79**) has a cinnamoyl group at C-8,<sup>56,66</sup> while compound **80** is an *N*-deethyl analogue of chasmanine.<sup>67</sup> Liaconitines A–C (**81–83**) are characterized by the presence of a  $\Delta^{2,3}$  double bond.<sup>68</sup> It should be mentioned that 14-*O*-veratrolylneoline and geniculatine B have the same structure, **88**.<sup>52,71</sup> Aconitilearine (**94**) contains a C-19 hydroxyl group.<sup>74</sup> Geniculine (**99**) represents the first example of



the occurrence of a *p*-hydroxybenzoyloxy group in the diterpenoid alkaloids.<sup>77</sup> Atropurpursine (**103**) is a rare aconitine-type alkaloid with 2β,3α-dihydroxyl groups.<sup>79</sup> Faleoconitine (**104**) contains a rare *N*-formyl functionality, which was confirmed by X-ray diffraction analysis.<sup>76</sup> Transconitine E (**105**) has a ketone group at C-1.<sup>67</sup>



Most of the aconitine-type alkaloids (such as **109–113**) possess an  $\alpha$ -oriented hydroxyl group at C-6. In contrast, alkaloids **114–116** are in a minority, with a  $\beta$ -oriented substituent at C-6. The structure of 6-*epi*-forsticine (**109**) was established by its 2D NMR data and X-ray diffraction analysis. Accordingly, the structure of forsticine was then revised from **109** to **117**.<sup>82</sup>

Compounds **118–129** contain oxygenated functionalities at both C-6 and C-15,<sup>89,91–96,98</sup> which might be regarded as a more evolved group of aconitine-type C<sub>19</sub>-diterpenoid alkaloids.<sup>17</sup> 1-*epi*-Deacetylaconitine (**118**) is the only aconitine-type alkaloid with a  $\beta$ -oriented methoxyl group at C-1, and an intramolecular hydrogen bond in **118** was observed between the nitrogen atom and the hydrogen of the 3 $\alpha$ -hydroxyl group.<sup>89</sup> 8-*O*-Methylhypaconine (**121**) was obtained from the underground parts of an unknown species of *Aconitum* cultivated in Korea, which originated from the Sichuan province of China.<sup>91</sup> Beiwucine (**122**) and 3-acetylaconifine (**124**) have a C-10 hydroxyl group.<sup>92,94</sup> The structure of 3-acetylmesaconitine (**125**) obtained from *Aconitum kusnezoffii* is as the same as that of tuberanine from *Aconitum tuberosum*.<sup>94,95</sup> Similarly, spicatin B (**126**) from *Aconitum spicatum* is structurally identical to 10-dehydroxyflavaconitine from *Aconitum nagarum*.<sup>93,96</sup> Tuberacnaitine (**128**) and tuberacnaitine (**129**) feature an acetonide unit at C-8 and C-15, as proved by their spectral data and chemical correlations with aconitine and mesaconitine.<sup>98</sup>

Three alkaloids (**130–132**) that possess an amino group at C-8 were isolated.<sup>83,99,100</sup> Hemsleyatine (**130**) represents the first example of this group,<sup>99</sup> while the possibility of lasianine (**131**) being an artifact was excluded through chemical transformations.<sup>100</sup>

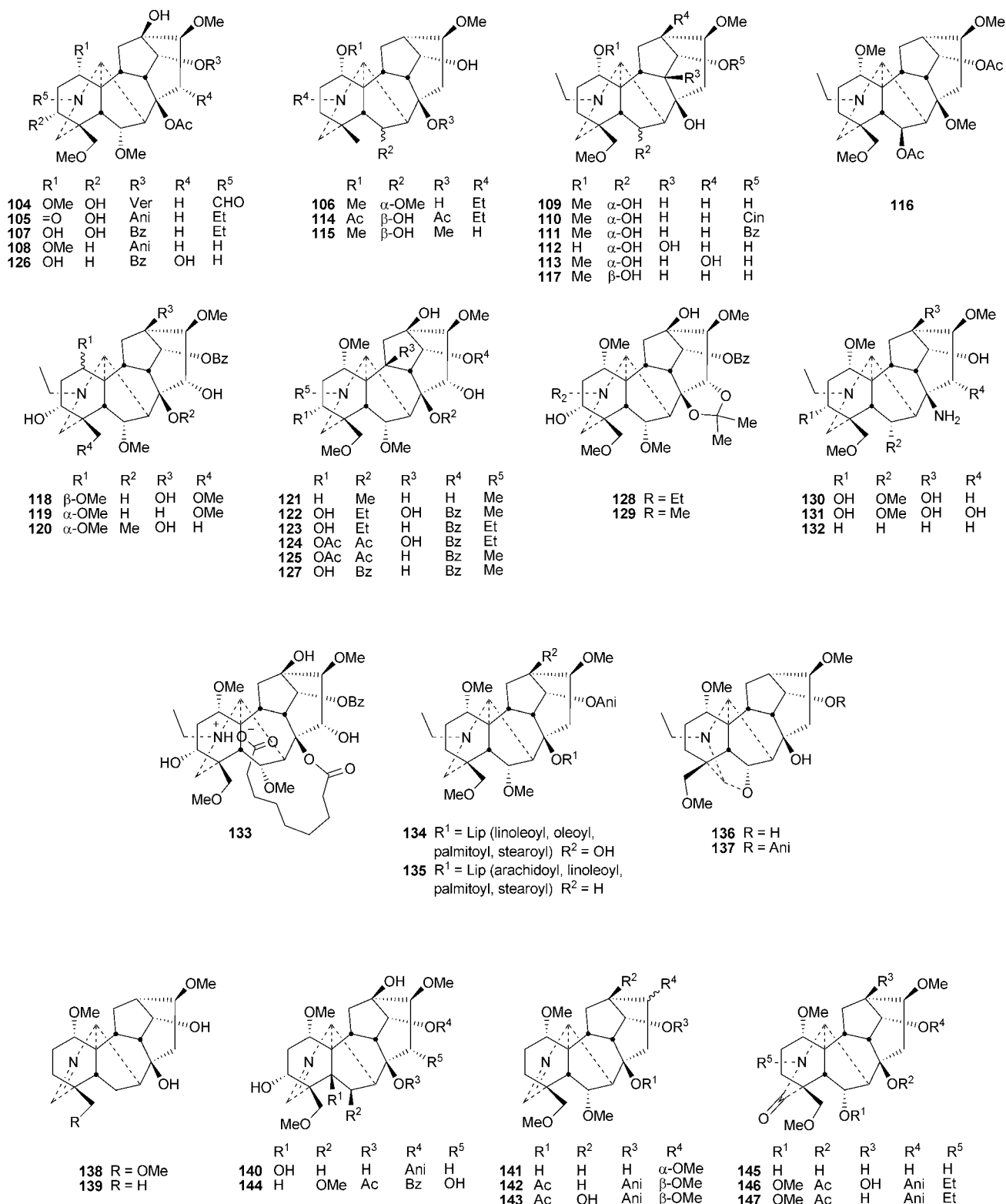
The term “lipo-alkaloid” was originally given by Kitagawa *et al.*,<sup>101</sup> and three new members of this group have been obtained

from *Aconitum* species since 1998. 8-*O*-Azeloil-14-benzoylaconine (**133**) features a zwitterionic structure between the nitrogen atom and the azelaic acid chain at C-8.<sup>13</sup> It was proposed, with support from MS data, that lipo-14-*O*-anisoylbikhaconine (**134**) and lipoforesaconitine (**135**) have long-chain fatty acid esters at C-8.<sup>71,102</sup>

Pengshenine A (**136**) and transconitine D (**137**) are the first two examples of naturally occurring aconitine-type C<sub>19</sub>-diterpenoid alkaloids with an *N*-C-19-*O*-C-6 mixed acetal moiety.<sup>67,103</sup> Several aconitine-type C<sub>19</sub>-diterpenoid alkaloids (**138–144**) with a C-19=*N* unit from various plant sources were identified. Among these, nagadine (**138**)<sup>43</sup> from *Aconitum nagarum* var. *lasiantrum* is identical to pengshenine B from *Aconitum hemsleyanum* var. *pengzhouense*.<sup>103</sup> Merckonine (**144**), the first member of the imine group, was determined on the basis of its physical and spectroscopic data and chemical correlation with aconitine.<sup>105</sup> Alkaloid piepunensine A (**145**) has an *N*-deethyl lactam group.<sup>54</sup> Habacnines A (**146**) and C (**147**) are two amide-subtype alkaloids.<sup>81,106</sup>

**2.2.2 Lycoctonines.** The lycoctonine-type C<sub>19</sub>-diterpenoid alkaloids are structurally characterized by the presence of an oxygenated group at C-7. They may also be subdivided into the following subtypes according to their nitrogen patterns: amines, *N,O*-mixed acetals, imines, and amides. The amine subtype constitutes the majority of this type of alkaloids, and could be further subdivided into various groups according to what kind of C-7/C-8 oxygen-containing groups they possess.

Alkaloids **148–167** are a group of lycoctonine-type alkaloids, isolated from various plants, which each contain a C-7,C-8 diol. Consolarine (**148**) features an  $\alpha$ -oriented hydroxyl at C-6,<sup>107</sup> and racemuloline A (**149**) lacks an oxygenated group at C-6.<sup>50</sup> **150–152** were obtained from two *Aconitum* species, among which **150** is identical to excecoitine from the same plant (*Aconitum excelsum*).<sup>34,35</sup> 16-Demethyldelsoline (**153**) is uncommon in having a hydroxyl group at C-16, whose structure was established based on its 1D and 2D NMR data and the correlations with delsoline.<sup>109</sup> Giraldines A–F (**154–156**, **176**, **157–158**) possess a  $\Delta^{2,3}$  double bond.<sup>110,111</sup> In addition, **158** has a carbonyl



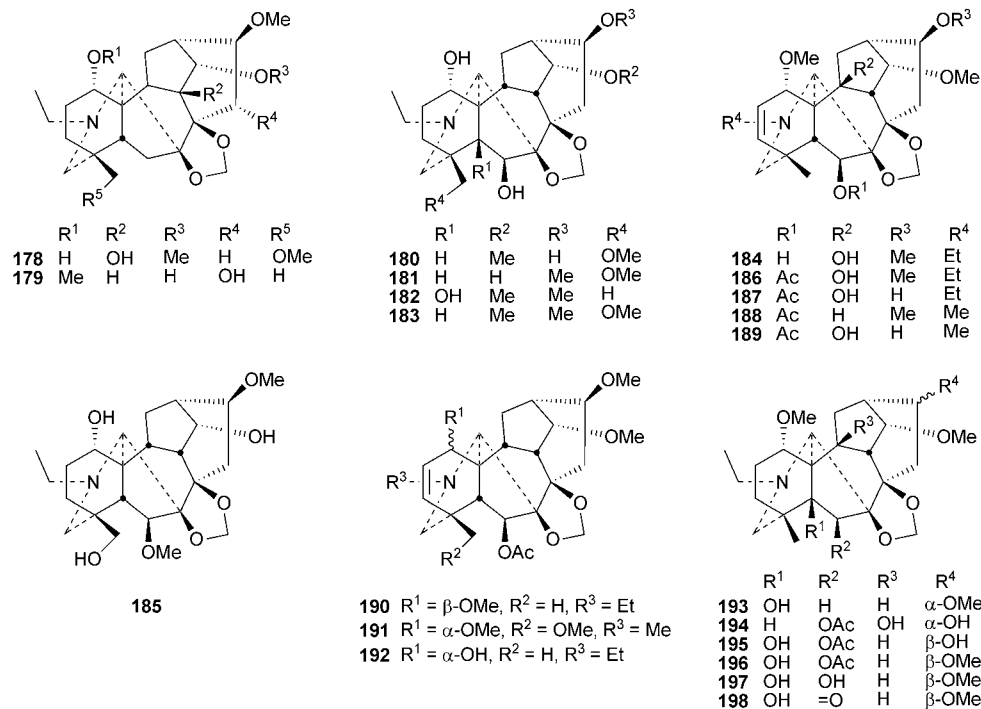
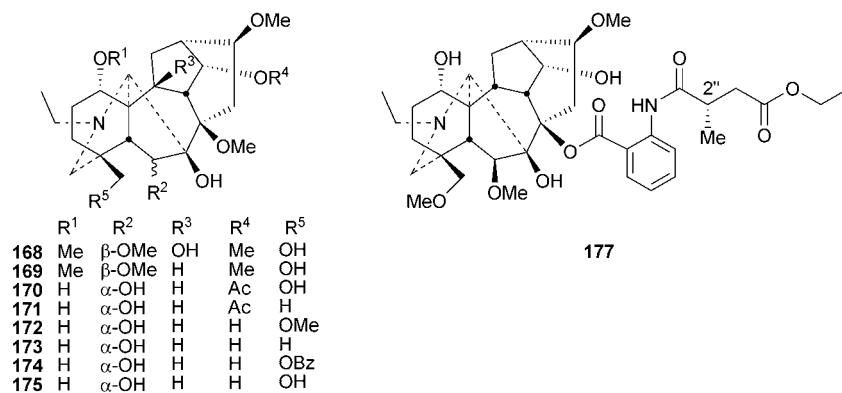
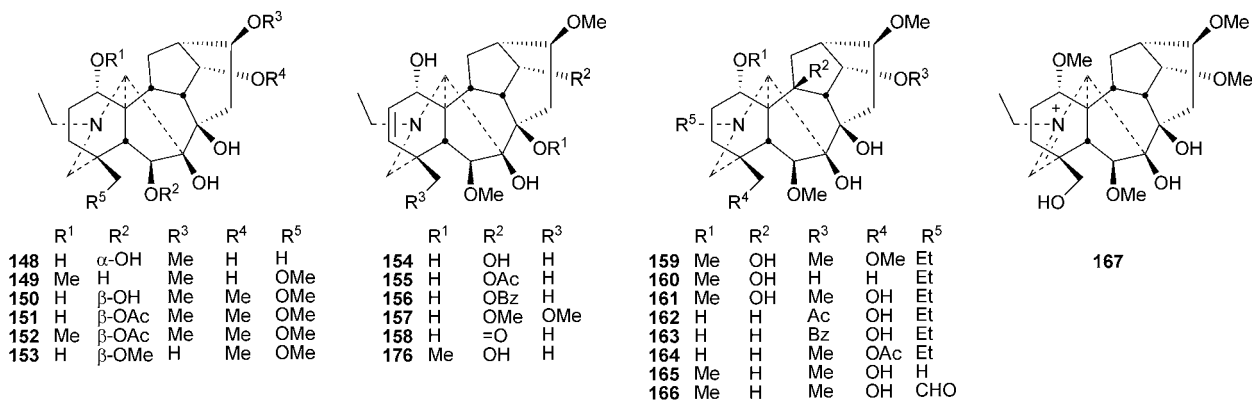
group at C-14, and **176** possesses a methoxy group at C-8.<sup>111</sup> Davidisines A (**165**) and B (**166**) were identified as *N*-deethyllycoctonine and *N*-aldehyde-lycoctonine, respectively,<sup>115</sup> by spectral methods, and the structure of the previously reported potanisine A was revised from **166** to **167**.<sup>116</sup>

Nine members (**168–176**) were added to the group of licoctonine-type alkaloids with a 7-OH/8-OMe unit. **170–175** are 6-*epi*-lycoctonine-type alkaloids.<sup>39,114,117,118</sup> Jiufengsine (**177**) is the first example of a naturally occurring licoctonine-type alkaloid with a C-8 anthranoyl group.<sup>119</sup> The absolute

configuration of C-2'' was suggested to be *S* because such a side chain was supposedly derived from the *N*-(methylsuccinimido)anthranoyl group.<sup>120</sup>

The alkaloids **178–198** fall into the group of lycotoxine-type alkaloids with a 7,8-methylenedioxy moiety. Almost all of these

compounds originate from the *Delphinium* species except for isodelatine (**179**), which comes from the roots of *Aconitum tai-paicum*.<sup>122</sup> It is really rare for such a lycotoxine-type alkaloid with a 7,8-methylenedioxy group to be found from the *Aconitum* species. Delcorinine (**180**) features a C-16  $\beta$ -hydroxyl group, and



its structure was established by its spectral data and the correlations with delcorine and delsoline.<sup>123</sup> Four new alkaloids, siwanines A–D (**186–189**), have a  $\Delta^{2,3}$  double bond.<sup>129</sup> The relative stereochemistry of **189** was confirmed by its X-ray crystallographic analysis. Nordhagenines A–C (**193–195**) have an unusual hydroxyl group at C-16,<sup>131</sup> the structures of **194** and **195** being confirmed by X-ray crystallographic analysis. Bonvalotindines A–C (**196–198**) possess a C-5 hydroxyl group.<sup>132</sup>

There is a group of lycoctonine-type C<sub>19</sub>-diterpenoid alkaloids characterized by the presence of an anthranoyl substituent at C-18. It consists of 27 new members (**199–225**), and most of them have a C-7,C-8 diol. Jiufengine (**199**) and jiufengdine (**200**) are two C-18 anthranoyl-containing alkaloids with an unusual  $\alpha$ -oriented methoxy group at C-6.<sup>133</sup> Alkaloid **202** is an analogue of ajadine.<sup>38</sup> The new alkaloids **203**, **204** and sinomontanine I (**205**) possess an *N*-(succinimido)anthranoyl unit at C-18.<sup>41,135</sup> Bearline (**207**), 14-acetylbearline (**208**), and 16-deacetylgeyerline (**209**) are three alkaloids that contain an *N*-(methylsuccinimido)anthranoyl unit.<sup>136</sup> The stereochemistry of C-3'' in both **214** and **215** was deduced as *S* based on comparison of the <sup>13</sup>C NMR data with those of the known delsemine B. 16-Demethoxydelavaine (**218a** and **218b**) is a mixture of two

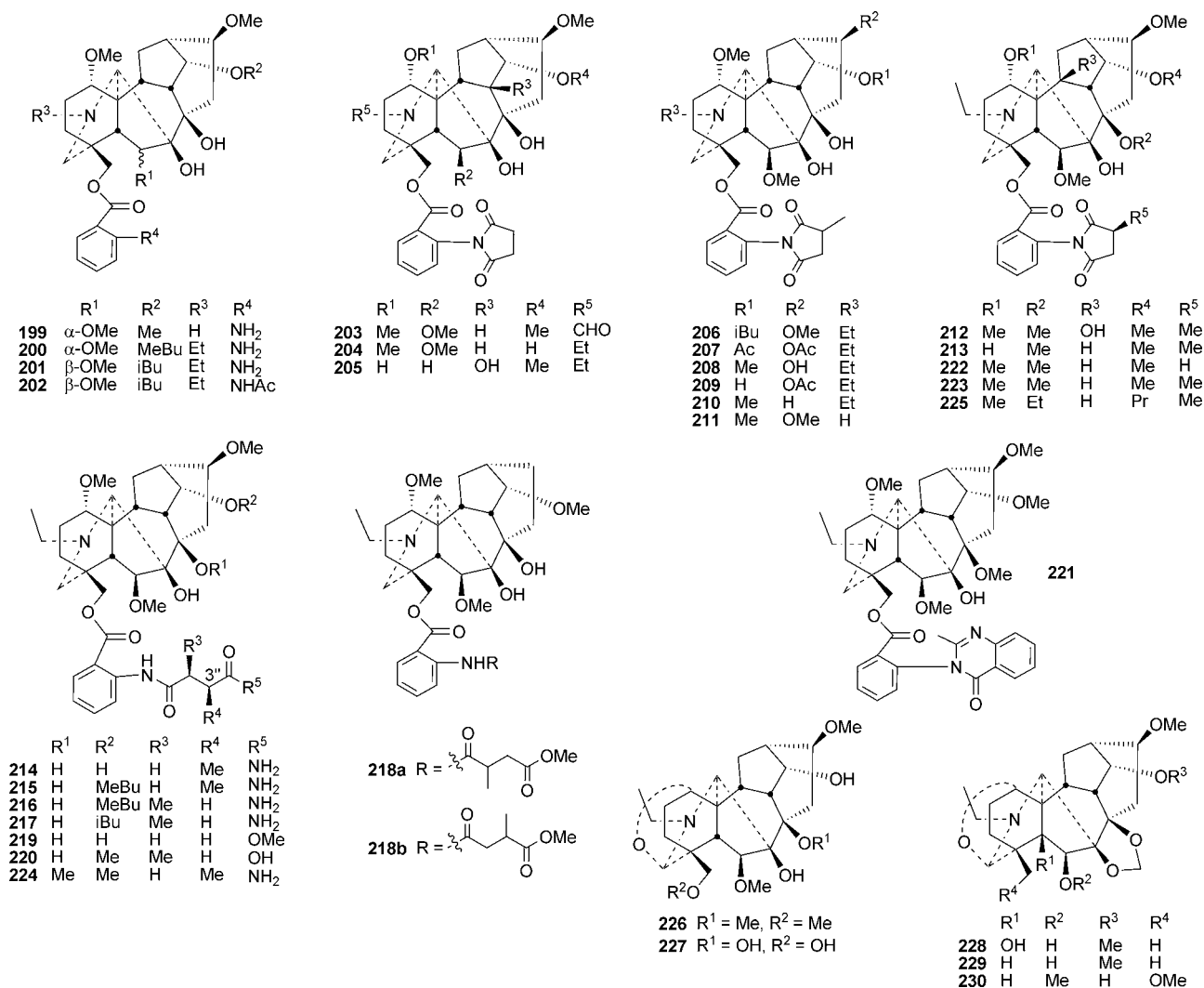
regioisomers of lycoctonine-type alkaloids.<sup>139</sup> Four alkaloids in this group (**222–225**) have alkoxy substituents at C-8.<sup>135,142,143</sup>

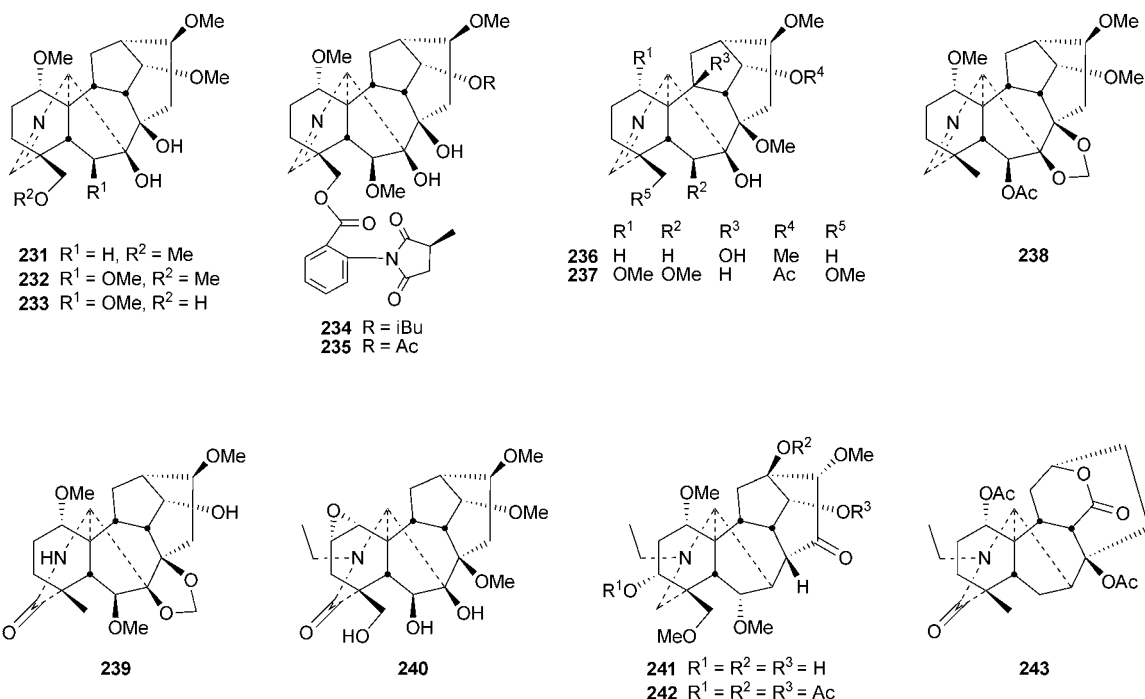
Five new lycoctonine-type alkaloids **226–230** with an *N*-C-19-*O*-C-1 mixed acetal unit were isolated from the genera *Consolida* and *Delphinium*.<sup>39,114,125,128</sup> Laxicymine (**228**) and laxicyminine (**229**) possess a methylenedioxy group at C-7 and C-8, and the former has an additional C-5 hydroxyl group.<sup>125</sup>

Alkaloids **231–238** have a C-19=*N* imine; they come from various plants. Several *N*-(methylsuccinimido)anthranoyl lycoctonine alkaloids, such as **234**, **235** etc., were isolated from *Delphinium pentagynum* by Herz *et al.*, suggesting that it could be a highly poisonous plant.<sup>146</sup>

There are three alkaloids (**166**, **239** and **240**) that belong to the lycoctonine-type C<sub>19</sub>-diterpenoid alkaloids with an amide group. Davidisine B (**166**) was established as *N*-formyllycoctonine;<sup>115</sup> tongolenine D (**239**) and budelphine (**240**) feature a lactam moiety.<sup>144,147</sup> In addition, budelphine (**240**) possesses an epoxy unit at C-1,C-2.<sup>147</sup>

**2.2.3 Pyro type.** Pyro-type C<sub>19</sub>-diterpenoid alkaloids that contain a  $\Delta^{8,15}$  double bond or an 8-H/15-ketone unit are considered to be derived from the elimination of the C-8



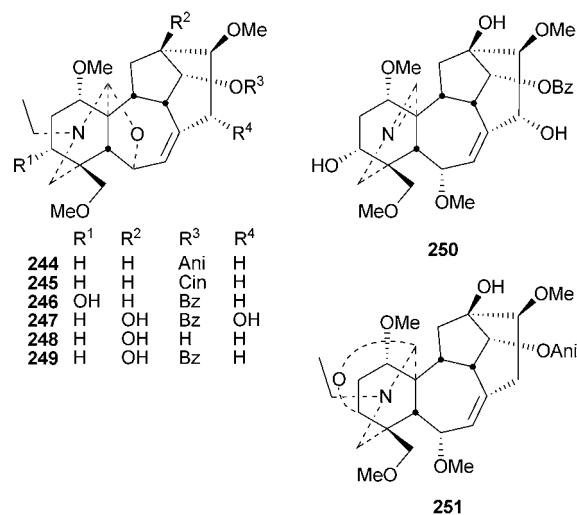


oxygenated group of aconitine-type alkaloids. 16-*epi*-Desbenzoylpyroaconitine (**241**) was initially prepared from aconitine by Katz and Rudin in 1984.<sup>148</sup> It was then isolated from processed aconite by Mori *et al.* in 1989, which was demonstrated to be induced by the heating of aconitine.<sup>149</sup> Further investigation on *Aconitum nagarum* var. *lasiandrum* by Wang *et al.* also led to the isolation of this 15-keto-type alkaloid. Its structure was established by extensive interpretation of its 1D and 2D NMR data, and confirmed by single-crystal X-ray analysis of its derivative **242**.<sup>85</sup>

**2.2.4 Lactone type.** So far, nine C<sub>19</sub>-diterpenoid alkaloids with a  $\delta$ -lactonized C ring have been found. Among these, only souline B (**243**), which possesses a lactam unit as well, was found during the review period.<sup>86</sup>

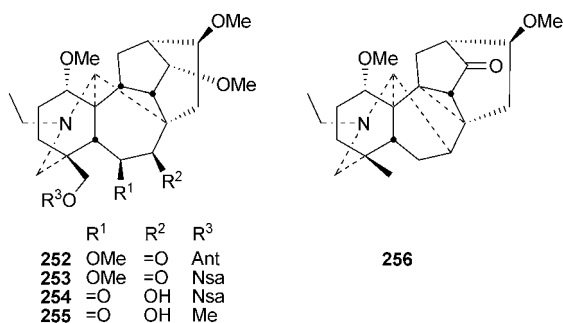
**2.2.5 7,17-Seco type.** The 7,17-seco-type C<sub>19</sub>-diterpenoid alkaloids, which also have a  $\Delta^{7,8}$  double bond, originated biosynthetically from aconitine-type alkaloids *via* Grob fragmentation. Franchetine, as one of the representatives of this type of alkaloids, was originally isolated from *Aconitum franchetii* by Chen and Sung.<sup>150</sup> Its initial structure was proposed based on its 1D NMR data. Ten years later, it was re-isolated by Wang *et al.* from *Aconitum hemsleyanum* var. *pengshiese*, and its structure was revised based on 2D NMR data and chemical correlations.<sup>151</sup> Six analogues (**244–249**) of franchetine were isolated from the genus *Aconitum*. Among them, the structure of aconine should be revised, and was found to be identical to that of beiwudine (**247**), based on extensive comparison of their NMR data.<sup>153,154</sup> Secokaraconitine (**250**), whose structure was established by X-ray crystallographic analysis, is a new 7,17-seco alkaloid with a C-17=*N* unit. Its rings seem more conformationally flexible than that of the usual lycotoconine skeleton.<sup>155</sup>

Secoyunaconitine (**251**) is the second example of C<sub>19</sub>-diterpenoid alkaloids with an epoxy ring between C-3 and C-17.<sup>157</sup>

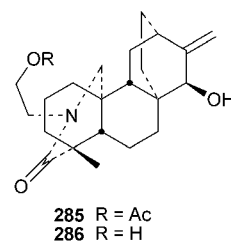
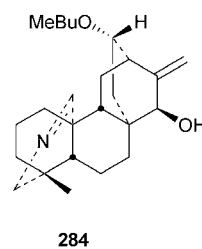
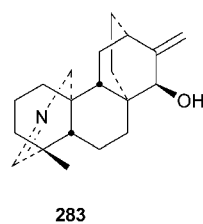
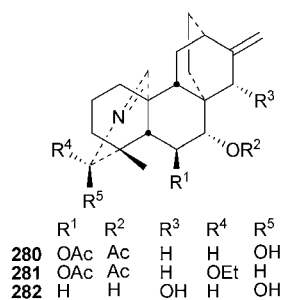
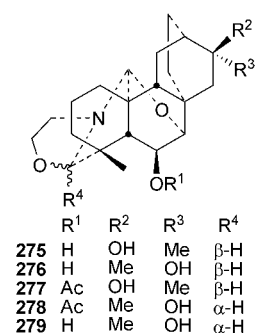
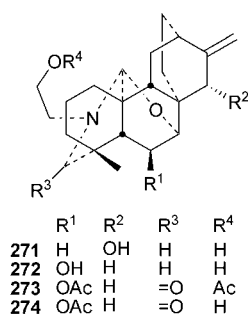
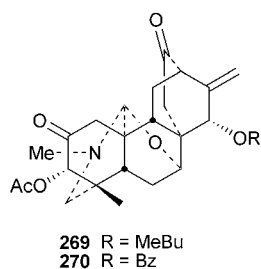
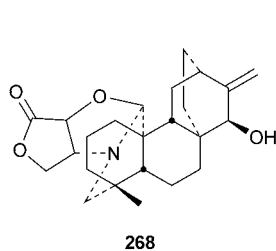
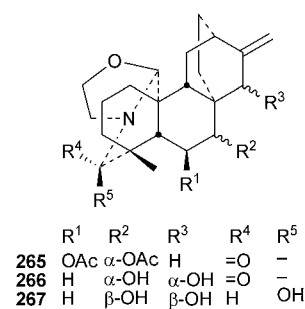
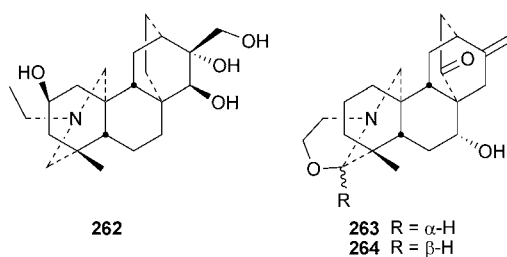
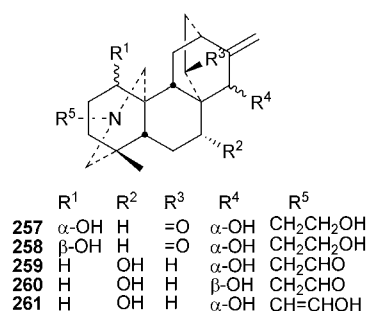


**2.2.6 Rearranged type.** There are five new C<sub>19</sub>-diterpenoid alkaloids with a rearranged skeleton. Acoseptine (**252**) and anhydrolycaconitine (**253**) contain a C-8–C-17 bridge (instead of a C-7–C-17 bridge) and a carbonyl group at C-7.<sup>158,159</sup> Rearranged skeletons of this type have been prepared from alkaloids with a 7,8-diol system *via* pinacol rearrangement.<sup>160,161</sup> The structure of **253** was confirmed by X-ray diffraction analysis of its hydrolysed derivative, and the possibility of rearrangement having occurred during the isolation procedure was excluded by experiment.<sup>159</sup> Septonine (**254**) and septontrionine (**255**) have a C-8–C-17 bridge, in addition to a ketone at C-6.<sup>162</sup>





Vilmoraconitine (**256**), as a novel rearranged C<sub>19</sub>-diterpenoid alkaloid, was isolated very recently by Tan and coworker from the roots of *Aconitum vilmorinianum*.<sup>163</sup> As the first representative of C<sub>19</sub>-diterpenoid alkaloid with a three-membered ring (C-8, C-9, C-10), its structure was determined by 2D NMR data and single-crystal X-ray analysis. It was proposed that **256** might be derived from vilmorrianine D, which can also be obtained from *A. vilmorinianum*.



## 2.3 C<sub>20</sub>-Diterpenoid alkaloids

**2.3.1 Atisines.** Atisine-type alkaloids have a pentacyclic core, and are considered to be the simplest group of C<sub>20</sub>-diterpenoid alkaloids.<sup>8</sup> There are 30 new members (**257–286**) isolated from various species of the genera *Aconitum*, *Consolida*, *Delphinium* and *Spiraea*.

Beiwusines A (**257**) and B (**258**) represent the first examples of atisine-type alkaloids with a hydroxyl group at C-1.<sup>164</sup> Spiratine A (**259**) and consorientaline (**260**), with an uncommon N-CH<sub>2</sub>CHO group, are epimers at C-15.<sup>165–167</sup> Uncinatine (**261**) possesses a Δ<sup>21,22</sup> double bond, whose structure was deduced by 1D and 2D NMR data, and its chemical correlations with dihydrojaconine.<sup>168</sup> Cochleareine (**262**) features a C-16,C-17 diol.<sup>88</sup> Compounds **263–267** all have an oxazolidine ring.<sup>166,169–171</sup> Delphatisine B (**268**) possesses a unique γ-lactone-fused oxazolidine ring.<sup>172</sup> Compounds **269–279**, isolated from the genus *Spiraea* in most cases, have a N-C-20-O-C-7 unit. Ouvrardiandines A (**269**) and B (**270**) have carbonyl groups at C-2 and

C-13.<sup>78</sup> Eleven atisine-type alkaloids (**259**, **265** and **273–281**) were isolated from *Spiraea japonica* var. *acuta* collected in Lijiang, Yunnan Province, China. Spiramine Z (**281**) with an ethoxyl group at C-19 could be an artifact formed during the extraction process.<sup>174</sup> The structures of spiramines P, Q, U and T (**275–278**) were revised based on extensive interpretation of its spectroscopic data and chemical transformations.<sup>175</sup> 13-(2-Methylbutyryl)azidine (**284**) features a C-17=N imine.<sup>178</sup>

**2.3.2 Denudatines.** The denudatine-type alkaloids are a class of hexacyclic C<sub>20</sub>-diterpenoid alkaloids based on atisines with an additional bond between C-20 and C-7. Ten new alkaloids (**287–296**) have been added to this group since 1998. Among them, only alkaloid **293** was isolated from the genus *Consolida*; the others were all from the genus *Aconitum*.

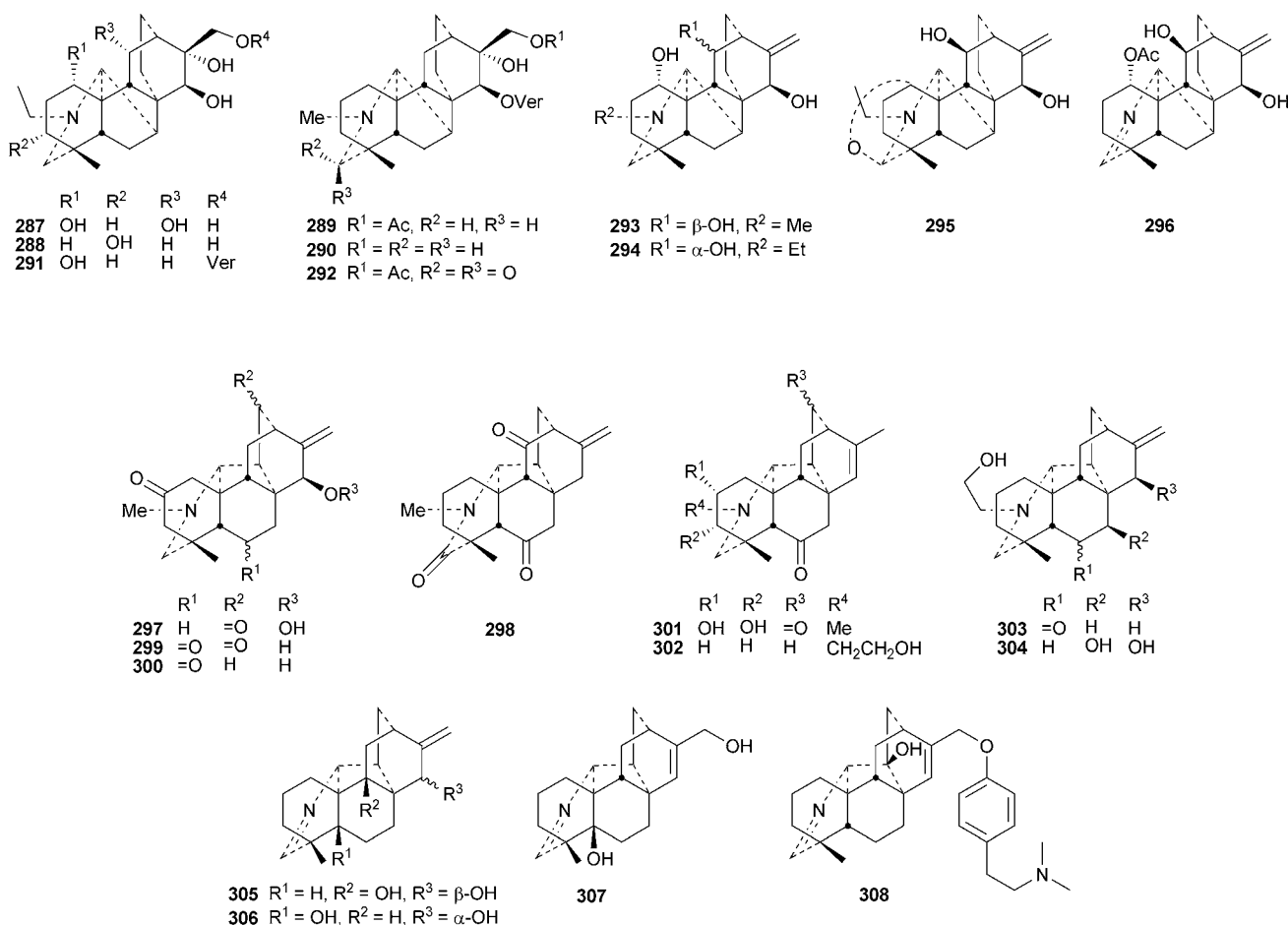
Alkaloids **287–292** possess oxygen-containing groups at both C-16 and C-17, while alkaloids **293–296** have a typical exocyclic double bond. In addition, **292** has a lactam unit.<sup>44</sup> Willipelletierine (**293**) is named after Professor S. W. Pelletier.<sup>181</sup> Kirinines B (**295**) and C (**296**) contain an *N,O*-mixed acetal and an imine unit, respectively.<sup>183</sup>

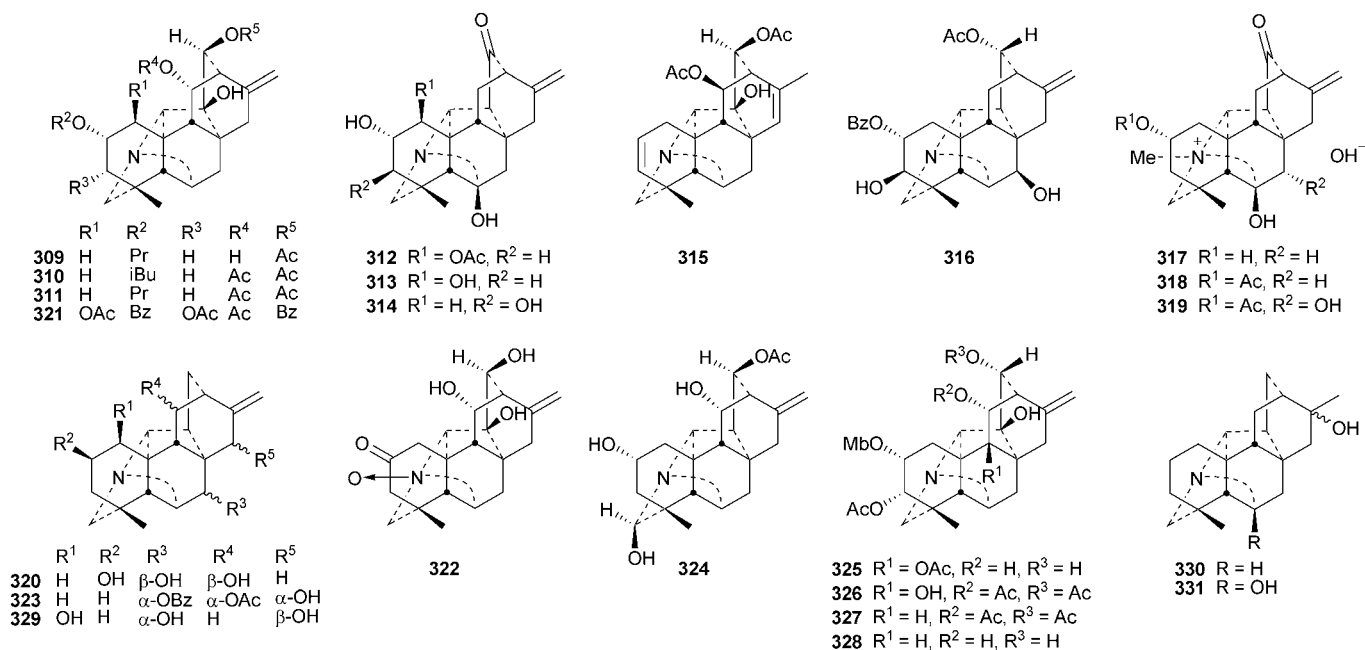
**2.3.3 Hetidines.** The hetidine-type alkaloids are a class of hexacyclic C<sub>20</sub>-diterpenoid alkaloids based on atisines with an additional C-20–C-14 bond. Alkaloids of this type include 13 new members: **297–299** from the genus *Delphinium*, **300**, **301** and **304–308** from the genus *Aconitum*, and **302–303** from the genus *Spiraea*.

Delcarduchol (**297**) has two carbonyl groups at C-2 and C-13, while carduchoron (**298**) contains three carbonyl groups at C-6, C-11 and C-19.<sup>184</sup> Racemulodine (**301**) has an endocyclic Δ<sup>15,16</sup> double bond, a 2,3-diol, and two carbonyl groups at C-6 and C-11.<sup>185</sup> Spirafine II (**302**) and spirafine III (**303**) possess an *N*-CH<sub>2</sub>CH<sub>2</sub>OH group and a carbonyl group at C-6.<sup>186</sup> Navirine (**308**) features a hordenine moiety at C-17 of the diterpenoid skeleton.<sup>190</sup> Naviculines A (**307**) and B (**306**) both possess an *N*=C-19 imine unit and a C-5 hydroxyl group.<sup>189</sup>

**2.3.4 Hetisines.** Hetisine-type C<sub>20</sub>-diterpenoid alkaloids have a heptacyclic system with an additional N–C-6 bond, compared to the hetidine-type alkaloids, and are one of the most complex groups derived from the atisine skeleton. Twenty-three new members (**309–331**), from various species of the genera *Aconitum*, *Consolida*, *Delphinium* and *Spiraea*, were added to this group during the review period.

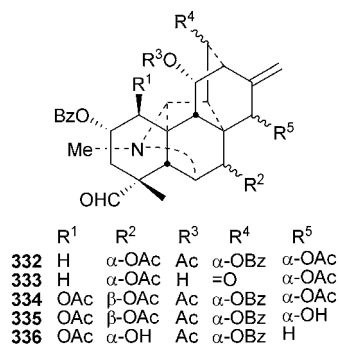
Alkaloids **309–311** have the same substitution patterns and differ in substitutions at C-2 and C-11, while **312–314** differ in substitutions at C-1 and C-3. In contrast, **315** possesses a Δ<sup>2,3</sup> double bond.<sup>191–193</sup> The regio-isomers guan-fu bases T (**313**) and U (**314**) were separated by preparative high-speed counter-current chromatography (HSCCC) coupled with evaporative light scattering detection (ELSD).<sup>195</sup> The structure of guan-fu base Q (**312**) was confirmed by single-crystal X-ray diffraction analysis.<sup>194</sup> Guan-fu base S (**315**) was obtained along with a new *ent*-kaurane diterpene, which may support the hypothesis of the





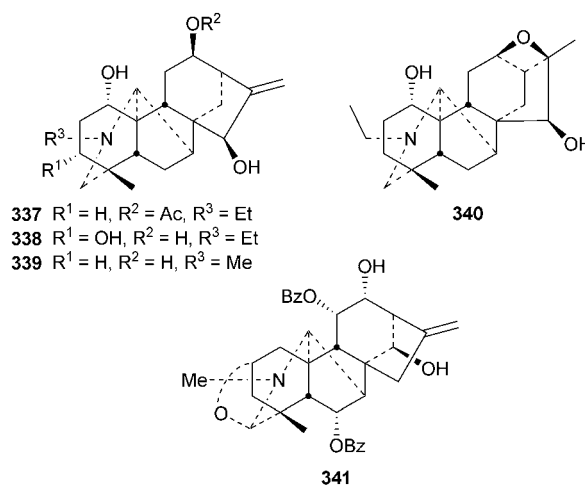
biosynthetic pathway of diterpenoid alkaloids involving a diterpene.<sup>196</sup> Three quaternary ammonium hydroxides, orochrine (317), 2-*O*-acetylorochrine (318), and 2-*O*-acetyl-7 $\alpha$ -hydroxyorochrine (319), were isolated from *Aconitum orochryseum*, a Bhutanese traditional medicine.<sup>198</sup> Four hetisine-type alkaloids (320–323) were isolated from the genus *Delphinium*, among which 322 from *Delphinium gracile* is a rare hetisine-type *N*-oxide.<sup>40,45–47</sup> 13-Acetylvakhmatine (324) contains a C-19 hydroxyl group. High-performance centrifugal partition chromatography (HPCPC) was used in the isolation of 324.<sup>199</sup> The four glanduline derivatives 325–328 differ from each other in the substituents at C-9, C-11 and C-13. Their structures were confirmed by single-crystal X-ray diffraction analysis of 326.<sup>200</sup> Spiraquine (330) and 6-hydroxylspiraquine (331) differ in the substituent at C-6.<sup>201</sup>

**2.3.5 Vakognavines.** Vakognavine-type C<sub>20</sub>-diterpenoid alkaloids have an *N*,19-*seco* hetisine skeleton in addition to a C-4 aldehyde group, and it was named after the first example of this type, vakognavine.<sup>202–204</sup> During the review period, five new members were added: 332–333 from *Aconitum tanguticum*, and 334–336 from *Delphinium trifoliolatum*. The structure of



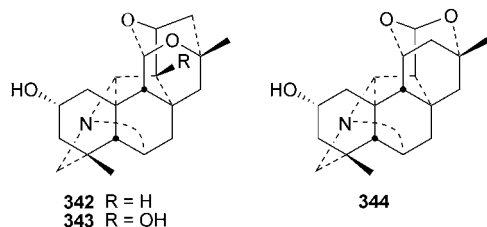
tangutisine B (333) was confirmed by an X-ray crystallographic analysis.<sup>205,206</sup> The structures of trifoliolasines D–E (334–336) were established on the basis of spectroscopic data, and by the X-ray diffraction analysis of 334.<sup>207</sup> The orientation of the axial C-13 benzyloxyl group is represented as  $\alpha$  according to the convention described by Pelletier.<sup>208</sup> In addition, we observed that a transannular effect occurs between the lone pairs of the nitrogen atom and the C-4 aldehyde group in the vakognavine-type alkaloids.<sup>207</sup>

**2.3.6 Napellines.** The napelline-type C<sub>20</sub>-diterpenoid alkaloids have a hexacyclic carbon framework with an additional C-20–C-7 bridge as compared with veatchine-type. Five new alkaloids (337–341) were added into this group since 1998. All five originated from various species of *Aconitum*. 12-Acetyl-12-*epi*-napelline (337) was structurally confirmed by X-ray crystallographic analysis.<sup>209</sup> The structure of 340 features an oxetane

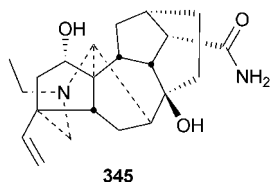


moiety consisting of C-12, C-13 and C-16, as confirmed by X-ray single-crystal analysis. Racemulotone (**341**) has an *N*-C-19-*O*-C-2 mixed acetal.<sup>211</sup>

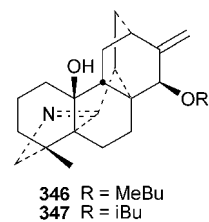
**2.3.7 Kusnezolines and omeielines.** Two novel products **342** and **344**, with an adamantane-type skeleton, were prepared by Pelletier *et al.* from hetisine *via* acid-catalyzed rearrangement.<sup>212,213</sup> They were not named at the time, and their structures were established by X-ray crystallographic analysis. Afterwards, we isolated **342** from *Aconitum kusnezoffii*, *Aconitum racemosum* var. *pengzhouense* and *Delphinium omeiense*,<sup>214,215</sup> and **344** from *Delphinium omeiense*.<sup>216</sup> Meanwhile, we designated them as kusnezoline (**342**) and omeieline (**344**), which belong to the kusnezoline-type and omeieline-type diterpenoids, respectively. Full assignments of <sup>1</sup>H and <sup>13</sup>C NMR data for these two alkaloids were achieved through extensive interpretation of the 2D NMR data.<sup>217</sup> Guan-fu base K (**343**), whose structure was proved by single-crystal X-ray diffraction analysis, represented the only new example of kusnezoline-type alkaloids from Nature during the period 1998–2008.<sup>218</sup>



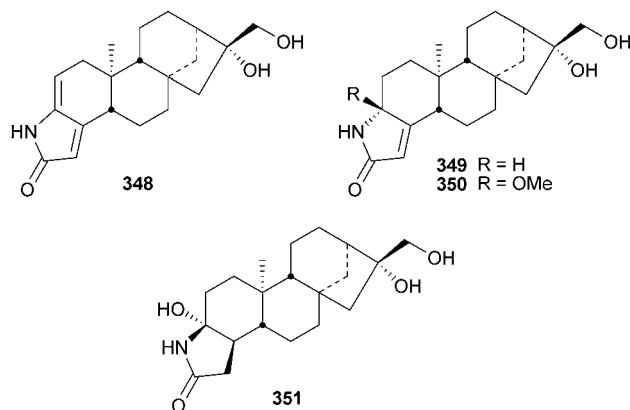
**2.3.8 Racemosines.** Further investigation of whole plants of *Aconitum racemosum* var. *pengzhouense* led to the discovery of racemosine, a novel C<sub>20</sub>-diterpenoid alkaloid with a unique skeleton (**345**).<sup>219</sup> The structure of **345** was established by 1D and 2D NMR, as well as X-ray crystallographic analysis. It was proposed that racemosine originates from the denudatine-type diterpenoid alkaloids through double Wagner–Meerwein rearrangements of both rings A and C followed by functionalization of the exocyclic methylene group. This unique compound was assigned to the racemosine-type, and it is the only example so far.



**2.3.9 Arcutinines.** Two new alkaloids, arcutin(e) (**346**) and arcutinine (**347**), were isolated from the aerial parts of *Aconitum arcuatum* by Saidkhodzhaeva *et al.*<sup>220,221</sup> They possess an unusual C-5–C-20 bond in lieu of the traditional C-10–C-20 bond typical of this class of alkaloids, and were designated as arcutine-type alkaloids accordingly.<sup>8</sup> The structure of arcutin(e) was determined by single-crystal X-ray diffraction analysis, while that of arcutinine was established by spectral data and comparison with arcutin(e).



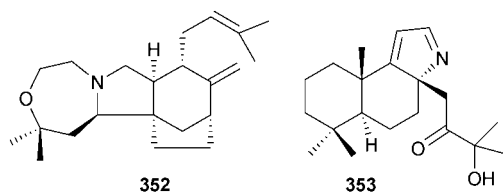
**2.3.10 Tricalysiamides.** From the wood of *Tricalysia dubia* collected in Okinawa, Japan, four rearranged veatchine-type C<sub>20</sub>-diterpenoid alkaloids with a cafestol-type diterpene framework, tricalysiamides A–D (**348–351**), were isolated recently.<sup>222</sup> These novel compounds are designated, after the first example, as tricalysiamide-type diterpenoids, which are characterized by the nitrogen atom being between C-3 and C-19. Their structures were determined based on their 2D NMR spectroscopic data and chemical correlations, as well as the single-crystal X-ray analysis of **348**.



## 2.4 Miscellaneous diterpenoid alkaloids

A series of nitrogen-containing diterpenes that do not belong to the typical diterpenoid alkaloids are discussed in this section.

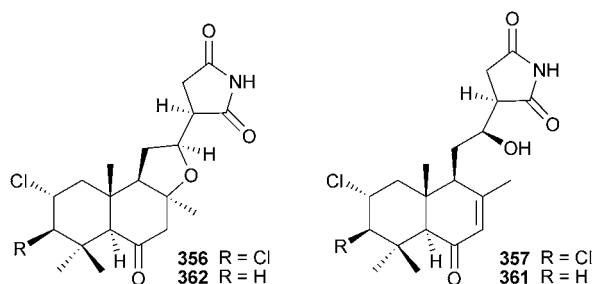
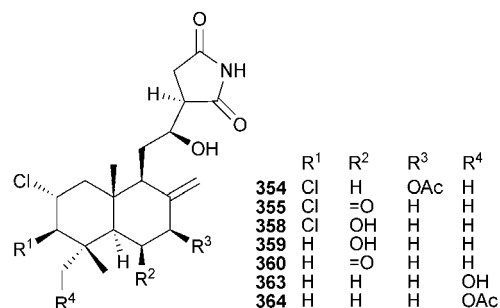
A novel alkaloid concavine (**352**) with an unprecedented ring system, consisting of dodecahydro-7-oxa-9a-aza-benzo[*a*]azulene, was isolated from cultures of *Clitocybe concava* (Basidiomycetae).<sup>223</sup> Biosynthetically, it could be regarded as a C<sub>20</sub>-diterpenoid alkaloid, with an additional hydroxyethylamine group as part of the aza-azulene ring. The four isoprenic moieties are evident in the formula, but their sequence is different from that common in most C<sub>20</sub>-diterpenoid alkaloids. This is the first report of a diterpenoid alkaloid from the Basidiomycetae.



Investigation of *Chamaecyparis obtusa* cv. *tetragon* by Tan and coworkers resulted in the isolation of chamobtusin A (**353**), a diterpenoid alkaloid with a novel skeleton featuring a

2*H*-pyrrole fused with a decalin unit.<sup>224</sup> The structure was mainly established by MS, 2D NMR and X-ray methods, and it is the first naturally occurring diterpenoid alkaloid from the Cupressaceae family.

Eleven new nitrogen-containing diterpenes, haterumaimides A–K (354–364), were isolated from ascidian *Lissoclinum* species by Ueda and coworkers.<sup>225–227</sup> They could be considered to have derived from the labdane-type diterpenes, and structurally they contain a succinimide moiety as well as chlorine atoms.



Recently, the Hao group isolated nine known diterpenoid alkaloids (one veatchine-type alkaloid and eight aconitines) from the fruits of *Daphniphyllum longeracemosum*.<sup>228</sup> It is worth noting that this is the first report of diterpenoid alkaloids from the genus *Daphniphyllum*.

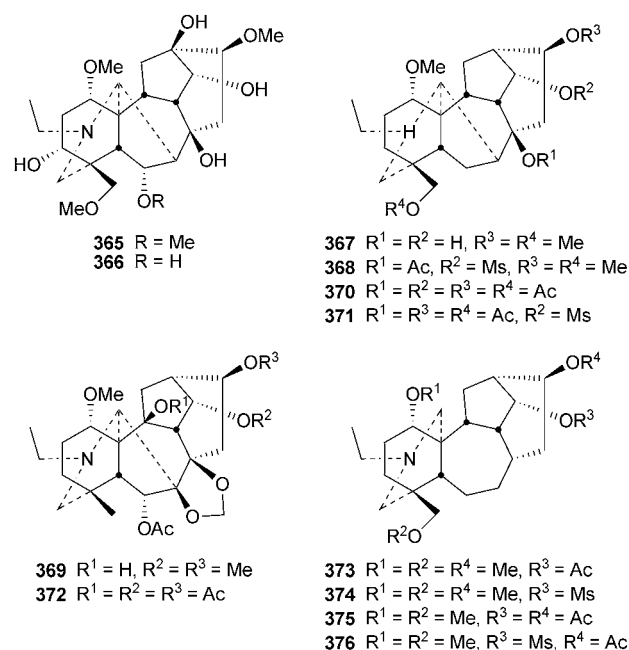
### 3 Chemical reactions

#### 3.1 *O*-Demethylation

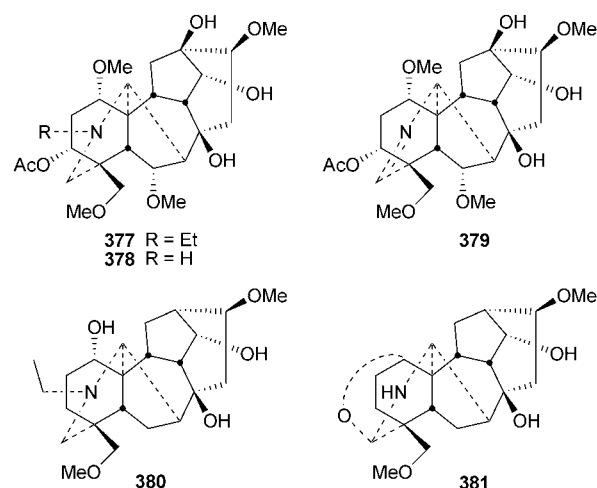
The 6-*O*-methyl group of pseudoaconine (365) could be selectively removed with 50% H<sub>2</sub>SO<sub>4</sub> to give the 6-*O*-demethyl product 366 in high yield (96%).<sup>229</sup> HBr–glacial acetic acid was reported to be an effective *O*-demethylation method for C<sub>19</sub>-diterpenoid alkaloids.<sup>230</sup> When this method [HBr–AcOH (20 equiv.), 50–80 °C, 7–20 h] was applied to the demethylation of the aconitine- and lycoctonine-type alkaloids (367–369), the corresponding *O*-demethylated products (370–372) were obtained in high yields (81–90%). However, when the 7,17-*seco* C<sub>19</sub>-diterpenoid alkaloids 373 and 374 were exposed to similar conditions, the *O*-demethylated products 375 and 376 were obtained only in low yields.

#### 3.2 Oxidation involving the nitrogen atom

**3.2.1 *N*-Deethylation.** During our investigation on the chemical reactions of diterpenoid alkaloids, a series of *N*-deethylation products were prepared using various oxidants such as KMnO<sub>4</sub> and NBS.<sup>231–233</sup> The resulting products depended greatly

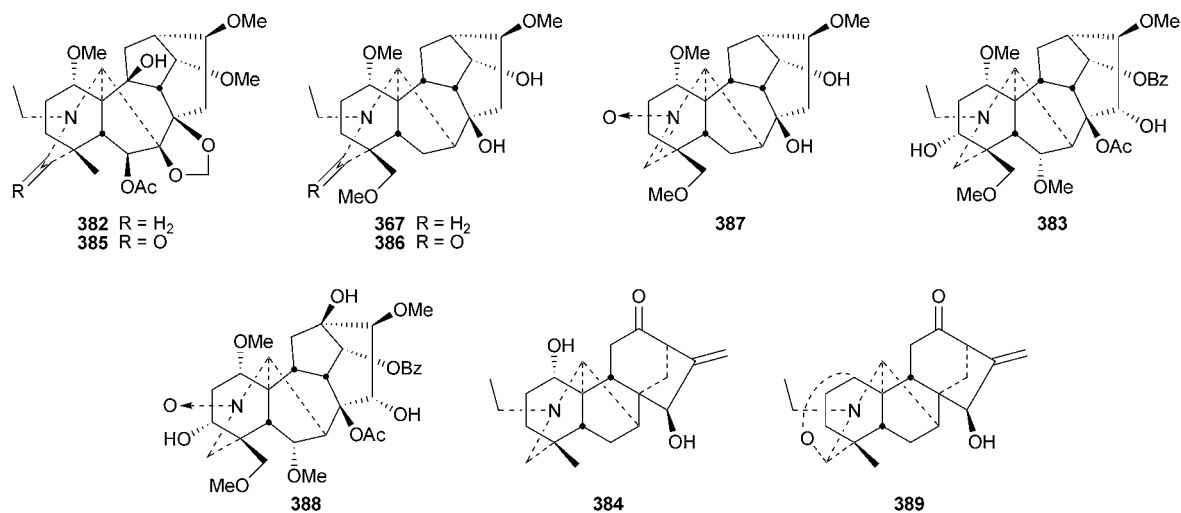


upon the reagents, reaction conditions and methods. It was found that treatment of 377 with NBS at room temperature produced the deethylated product 378, and the subsequent imine product 379.<sup>232</sup> In addition, an *N*-deethylation product 381, with a rare *N,O*-mixed acetal, could be prepared in high yield (98%) by prolonged treatment of isotalatizidine (380) with KMnO<sub>4</sub> at 40 °C.<sup>233</sup>



**3.2.2 Formation of amides, imines, nitrones and *N,O*-mixed acetals.** Oxidation of eldeline (deltaline, 382), talatisamine (367), aconitine (383) and songorine (384) with dimethyldioxirane yielded the compounds 385 (lactam, 19%), 386 (lactam, 26%) and 387 (nitrone, 45%), 388 (nitrone, 65%), and 389 (*N,O*-mixed acetal, 19%), respectively.<sup>234</sup> The authors have proposed a mechanism, which involves the oxidation of C-19 atom or the methylene moiety of *N*-ethyl group followed by elimination of acetaldehyde and reoxidation of dimethyldioxirane.<sup>235</sup> For

example, two nitrones **393** and **397** were prepared from lappaconitine (**390**) and elatine (**394**), respectively, using the method developed by Osadchii *et al.* The key reactions might include *N*-oxidation, Cope elimination (heat, *in vacuo*), and oxidation with  $K_3Fe(CN)_6/NaHCO_3$  (Scheme 1).<sup>235</sup> Interestingly, alkaline hydrolysis of **396** produces a mixture of *N*-deethyl-*N*-hydroxyelatidine **398** and nitron **399** in a ratio of 3 : 1. Oxidation of **398** with  $K_3Fe(CN)_6$  afforded **399** in quantitative yield, while reduction of **399** with  $NaBH_4$  gave **398** in 87% yield.<sup>235</sup>



Wang and coworkers showed that the corresponding imines can be readily prepared in 65–83% yield by heating (100–170 °C, 3–7 h) of certain diterpenoid alkaloids with DMSO.<sup>236</sup> Several imines, such as **402**–**404**, could be prepared as major products from talatisamine (**367**) and its derivatives **400** and **401** by this method. In the cases of **367** and **400**, trace amounts of iminium salts were also isolated. However, heating deltaline (**382**) and lappaconitine (**390**) with DMSO yields exclusively the *N*-deethyl derivatives instead of the imines.<sup>236</sup>

It was found that treatment of acetyllycoctonine (**405**) with *m*CPBA at room temperature yielded acetyllycoctonine *N*-oxide **406** as a major product, together with other interesting by-products (**407**–**413**). The key steps are proposed to be nitrogen oxidation, Cope elimination, and Polonovski-like fragmentation (Scheme 2).<sup>237</sup>

Six new products (**408**, **411**, **414**–**417**) were obtained from the reaction of acetyllycoctonine (**405**) with NBS (Scheme 3).<sup>238</sup> The products and yields of this reaction depend greatly upon the reaction conditions and the nature of the substrates.

In addition, it was also reported that the oxidation of elatidine (**418**) with  $CrO_3$  in acetic acid gives elatidal (**419**), from which two diamines, **420** and **421**, were prepared by the formation of the Schiff base followed by reduction with  $NaBH_4$ .<sup>239</sup>

### 3.3 Oxidation of alcohols

The intriguing aldol product **422**, as confirmed by crystal X-ray analysis, could be obtained in 41% yield from the reaction of

lappaconitine (**390**) with  $HIO_4$ . Subsequent reaction of this aldol with  $Br_2-HOAc$  could give diketone **423** (31%) and demethylated aldol product **424** (13%) (Scheme 4).<sup>240</sup> Very interestingly, a one-pot reaction of lappaconitine (**390**) with  $NaIO_4$  and  $Br_2-HOAc$  afforded the *N*-deethylated derivative **425** with a brominated aromatic ring.<sup>241</sup>

Wang *et al.* reported that the oxidation of pseudoaconine (**365**) with  $HIO_4$  generated various products (**426**–**431**) using different reaction media and work-up conditions (Scheme 5).<sup>242</sup>

Structurally, the product **431** is the first representative of a *C/D*-nor-rearranged  $C_{19}$ -diterpenoid alkaloid.

### 3.4 Ester-exchange reaction

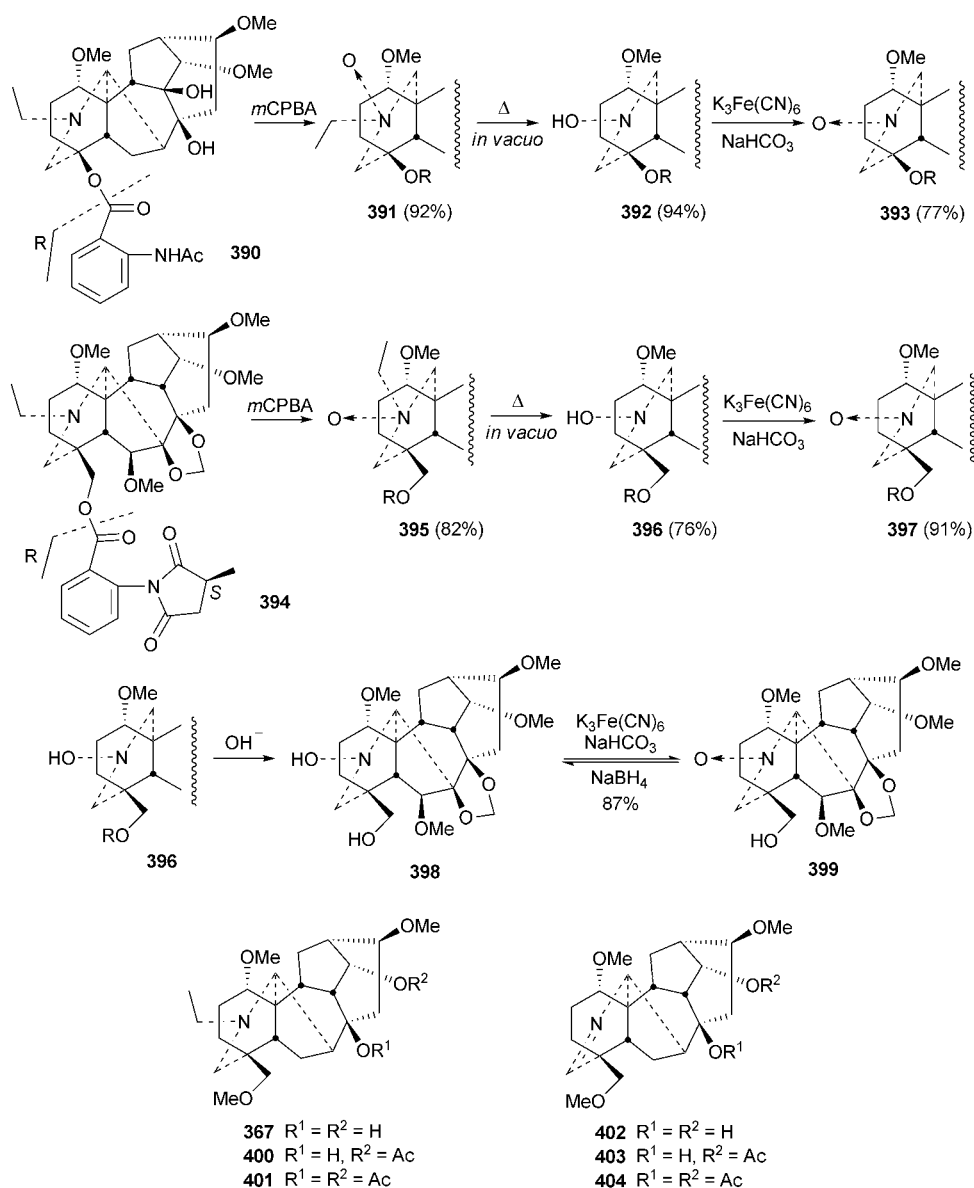
Liu *et al.* reported for the first time that certain diterpenoid alkaloids could be converted into the corresponding lipo-alkaloids during the process of decocting aconite root (involving the addition of palmitic acid to aconitine), or the mixing of mesaconitine and hypaconitine with liquorice roots.<sup>243</sup> A similar ester-exchange reaction was also observed in the biotransformation of aconitine in the presence of human intestinal bacteria.<sup>244</sup>

### 3.5 Biotransformation

It was demonstrated by ESI-MS/MS<sup>n</sup> studies that the incubation of aconitine with human intestinal bacteria *in vitro* produces very complex metabolites. More than 20 types of new compounds, such as mono- and di-ester aconitines, as well as lipo-alkaloids, were observed. The key transformations involve deacetylation, dehydroxylation, demethylation and esterification.<sup>244</sup>

### 3.6 Stability of aconitine, mesaconitine and hypaconitine

HPLC–ESI-MS/MS<sup>n</sup> showed that the decomposition products of aconitine, mesaconitine and hypaconitine mainly derived from hydrolysis and pyrolysis. The stability of these alkaloids depends on the pH value of buffer, the solvent, the storage time, and the substituents at C-3 and the nitrogen atom.<sup>245</sup>



Scheme 1 Formation of nitrones from lappaconitine (390) and elatine (394).

### 3.7 Cleavage of the $N=C$ -19 bond

The  $N=C$ -19 bond of **432** and **437** could be broken to yield  $N,19$ -seco  $C_{19}$ -diterpenoid alkaloids **436** and **438** with an oxaziridine ring, respectively. The sequence involves the formation of imine, quaternization of imine, formation of  $N,O$ -mixed acetal, and oxidation with *m*CPBA (Scheme 6).<sup>246</sup>

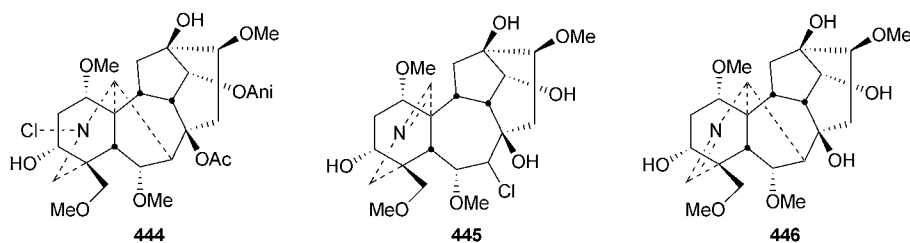
According to the method described in the literature,<sup>247</sup> the  $N,19$ -seco  $C_{19}$ -diterpenoid alkaloids **442** or **443** with a nitro or oxime group at C-17 were also prepared from **439** (Scheme 6).<sup>248</sup>

Some other  $N,19$ -seco  $C_{19}$ -diterpenoid alkaloids were also prepared employing the above-mentioned procedures.<sup>249</sup>

### 3.8 Cleavage of the C-7–C-17 bond

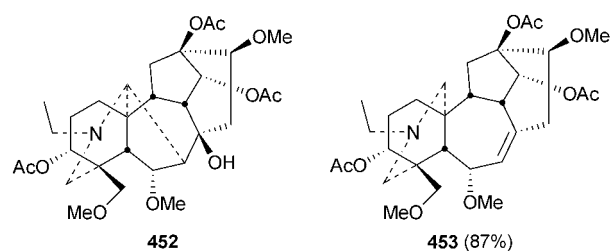
A mixture of 7,17-seco compound **445** and imine **446** was generated *via* the microwave irradiation of **444** in 1.2 M NaOMe for 30 s, but the yield was poor.<sup>250</sup>

Wang and co-workers developed a route to 7,17-seco  $C_{19}$ -diterpenoid alkaloids by a reaction sequence that included



selective hydrolysis, chlorination, Grob fragmentation, and NaBH<sub>4</sub> reduction (Scheme 7). By this approach, 7,17-seco C<sub>19</sub>-diterpenoid alkaloid **448** was generated from **447** in 60% yield; **450** and **451** were produced from isotalatizidine (**449**).<sup>251</sup> Reaction optimization suggested that the replacement of methanol with THF as solvent would significantly increase the yields.

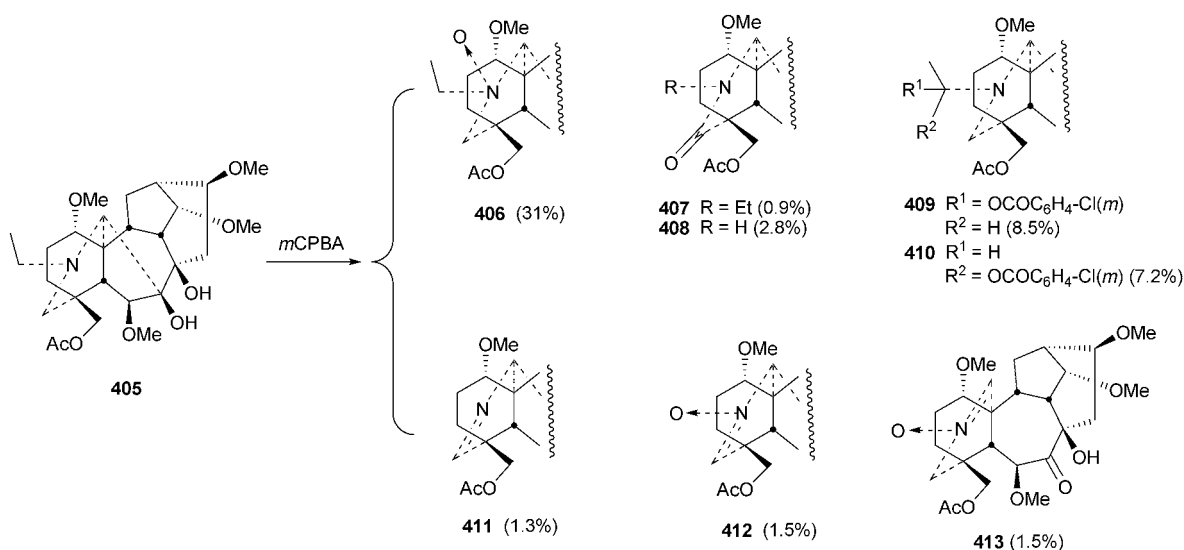
Compound **453** was generated by the rupture of the C-7–C-17 bond of alkaloid **452** in 87% yield.<sup>252</sup>



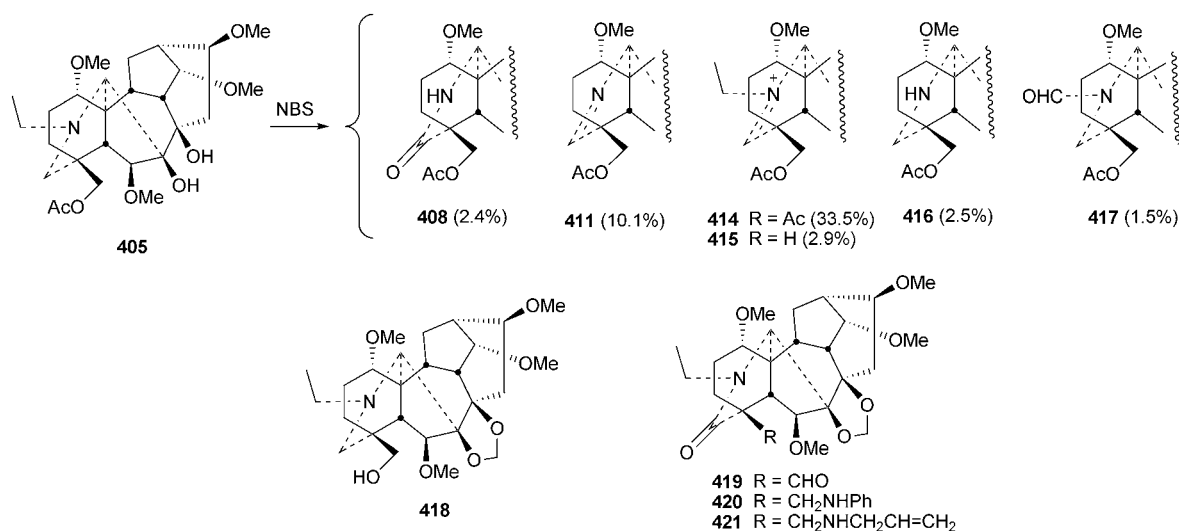
### 3.9 Rearrangement of ring A

An intriguing rearranged alkaloid **456**, whose structure was confirmed by single-crystal X-ray analysis of its acetylated derivative, was observed from a sequence of reactions starting from **454**. As shown in Scheme 8, the sequence involves the treatment of **454** with NBS followed by methyl iodide and

reaction of the resulting iminium salt **455** with 5% NaOH/MeOH.<sup>253</sup> A mechanism for this rearrangement was proposed, which involves double Grob fragmentations, formation of a new ring A by aldol reaction, coupled with an elimination of a molecule of methanol.

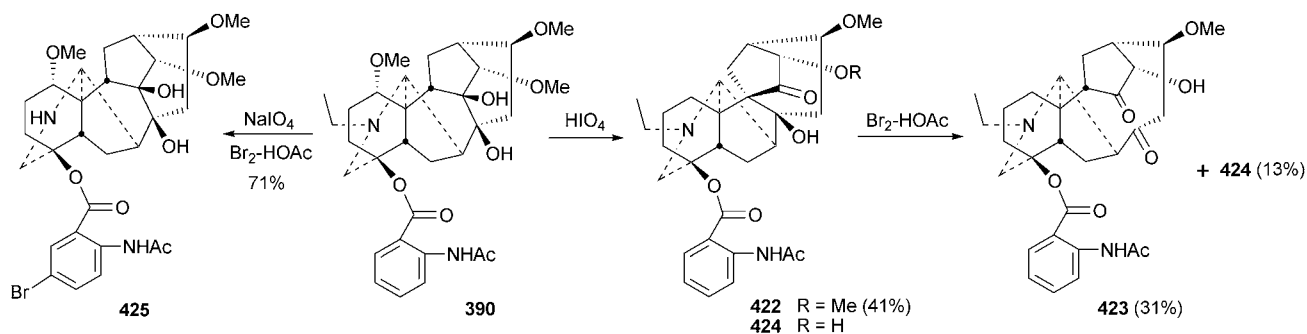


Scheme 2 Oxidation of acetylylcoctonine (**405**) with *m*CPBA.

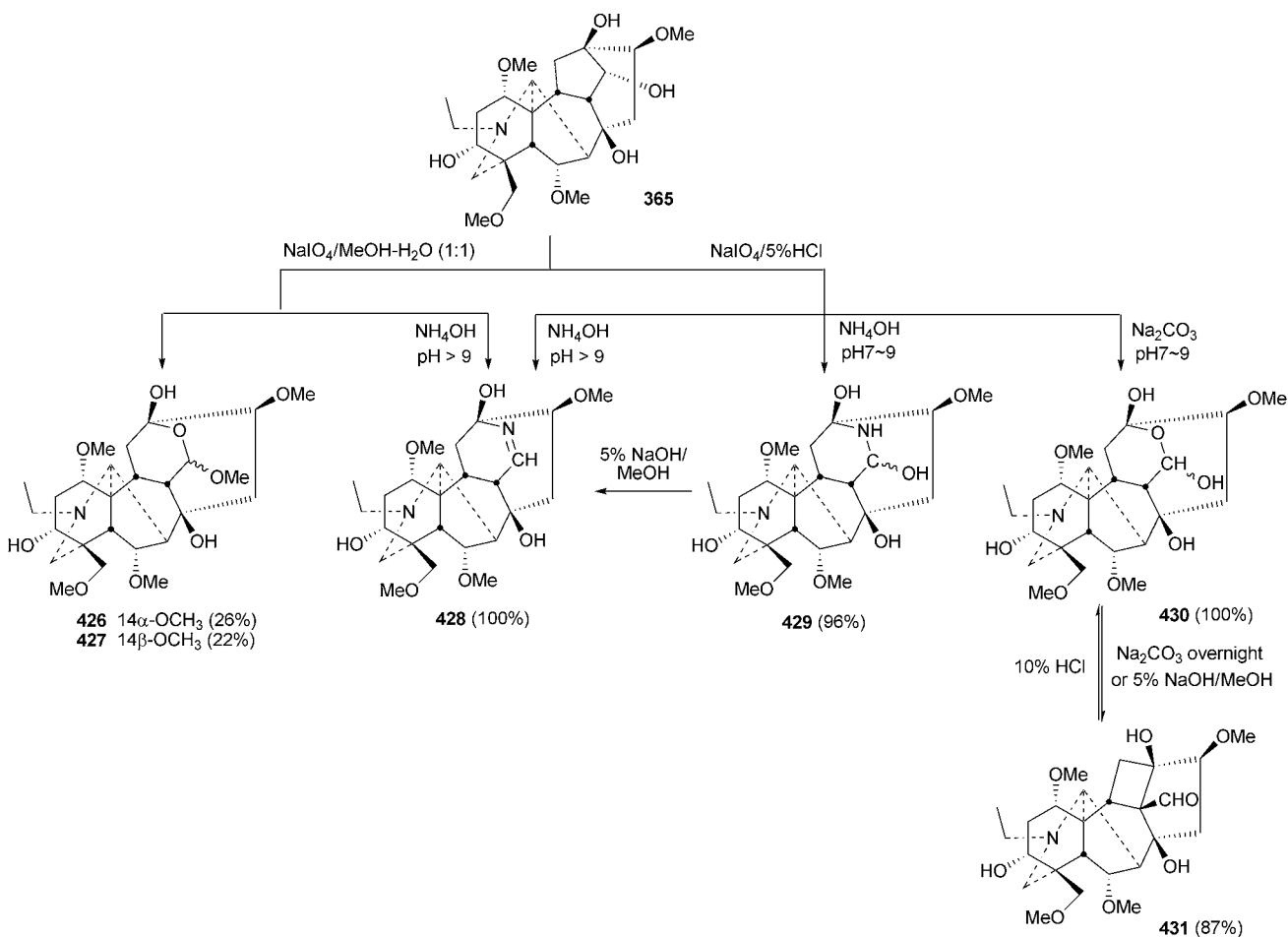


Scheme 3 Oxidation of acetylylcoctonine (**405**) with NBS.





Scheme 4  $\text{NaIO}_4$ -catalyzed bromination and  $\text{HIO}_4$  oxidation of lappaconitine (425).



Scheme 5 Oxidation of pseudoaconine (365) with  $\text{HIO}_4$ .

## 4 Synthetic studies

### 4.1 Synthesis of 12,13-seco $\text{C}_{19}$ -diterpenoid alkaloids

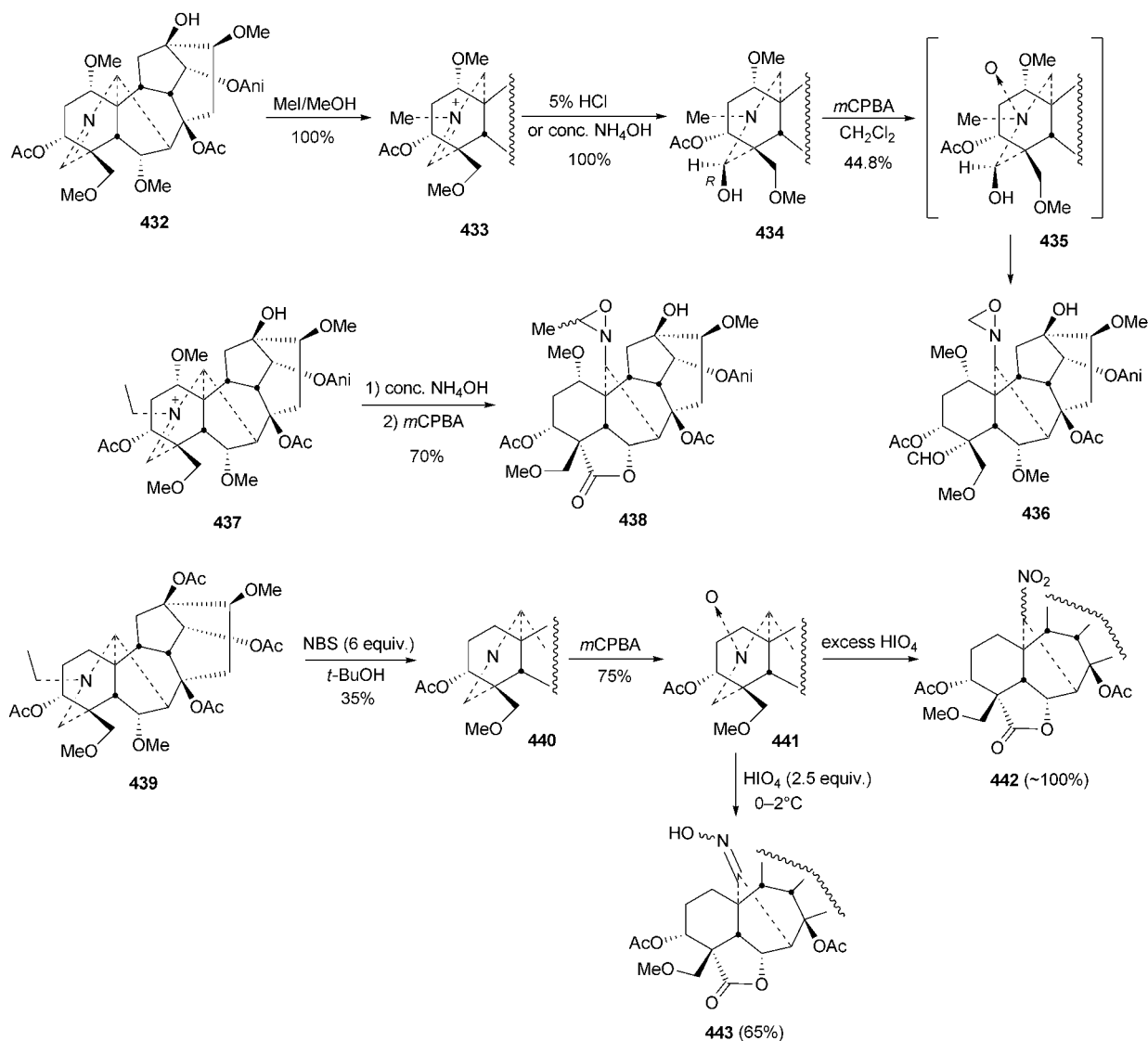
As shown in Scheme 9, Wang and coworkers developed a route to the 12,13-seco  $\text{C}_{19}$ -diterpenoid alkaloids 458 and 459, and the D ring aromatized 460–462, via semipinacol rearrangement of the pseudoaconine derivative 457 followed by reaction with  $\text{Br}_2\text{-HOAc}$ .<sup>254</sup>

The structures of 458 and 461 were confirmed by their 2D NMR data and single-crystal X-ray analysis.<sup>254</sup> Further studies

led to the preparation of a series of 12,13-seco  $\text{C}_{19}$ -diterpenoid alkaloids and their corresponding aromatic products in good to high yields.<sup>249</sup>

### 4.2 Conversion of $\text{C}_{19}$ -diterpenoid alkaloids into aconane-type diterpenes

Two novel aconane-type diterpenes (463 and 464) could be generated from *N*,19-seco nitro-compound 442 or nitrone 441



Scheme 6 Cleavage of the  $N=C$ -19 bond of compounds **432**, **437** and **439**.

through a Nef reaction or oxidation with  $\text{HIO}_4$ , respectively, in moderate yields (Scheme 10).<sup>255</sup>

### 4.3 Exploration on the approaches to taxoids from $\text{C}_{19}$ -diterpenoid alkaloids

After the exploration of four routes (ABC, ACB, BCA, and CAB) towards taxoids, Wang *et al.* reported that the vital intermediate **473** or **476** could be synthesized starting from yunaconitine (**465**) by the CAB approach, as shown in Scheme 11.<sup>256</sup> The key steps include the semipinacol rearrangement of **467** with  $\text{NaOH}/\text{DMF}$ , and the rupture of the  $N-C$ -19 bond (**472**  $\rightarrow$  **473**, **475**  $\rightarrow$  **476**).

### 4.4 Conversion of $\text{C}_{19}$ -diterpenoid alkaloid deltaline to the taxane ABC core system

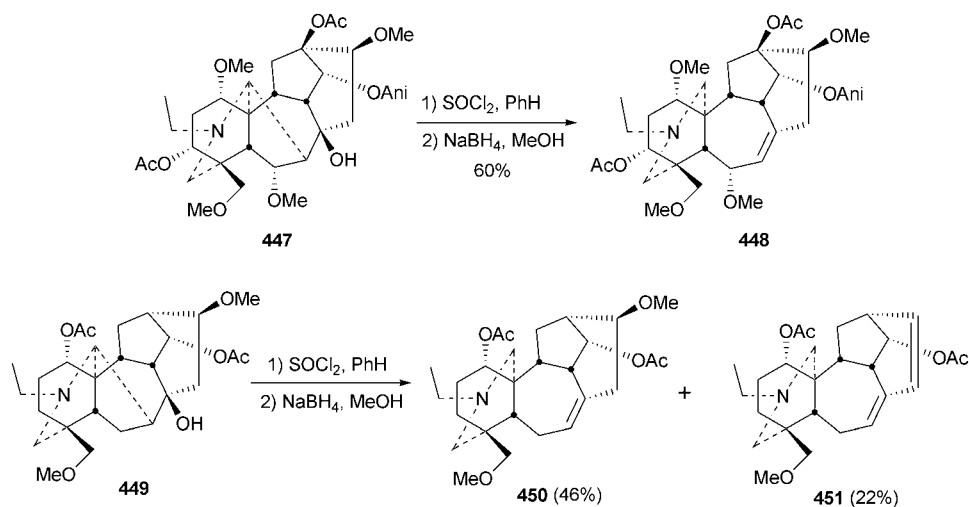
Following a long series of investigations into the chemistry of rings A,<sup>233,236,246,248,254,256</sup> B,<sup>249–251,256</sup> and  $\text{C}^{253,255,256}$  of the  $\text{C}_{19}$ -diterpenoid alkaloids and the preparation of key intermediates

towards the taxoid analogs,<sup>256</sup> Wang and coworkers recently reported a novel approach toward the taxane ABC core system employing deltaline (**382**) as the starting material in 18% overall yield.<sup>18</sup> As shown in Scheme 12, the key reactions included Grob fragmentation (**478**  $\rightarrow$  **479**), fission of the  $\Delta^{9,14}$  double bond followed by aldol condensation (**479**  $\rightarrow$  **480**), and Pelletier cleavage (**481**  $\rightarrow$  **482**). The structure of the taxoid analog **482** was confirmed by 2D NMR and single-crystal X-ray analysis.<sup>18</sup>

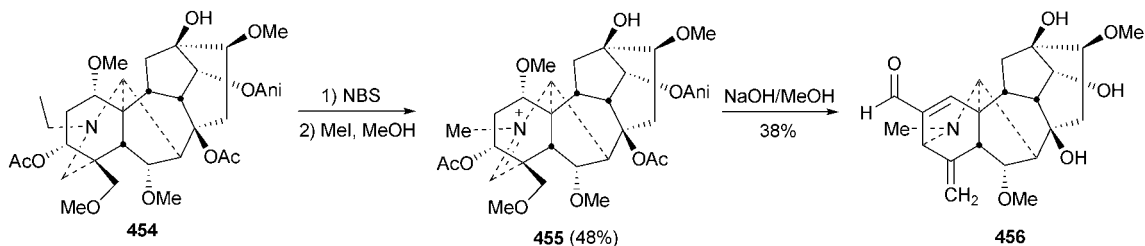
### 4.5 Total synthesis of $\text{C}_{20}$ -diterpenoid alkaloid nominine

#### 4.5.1 Muratake and Natsume's synthesis of nominine.

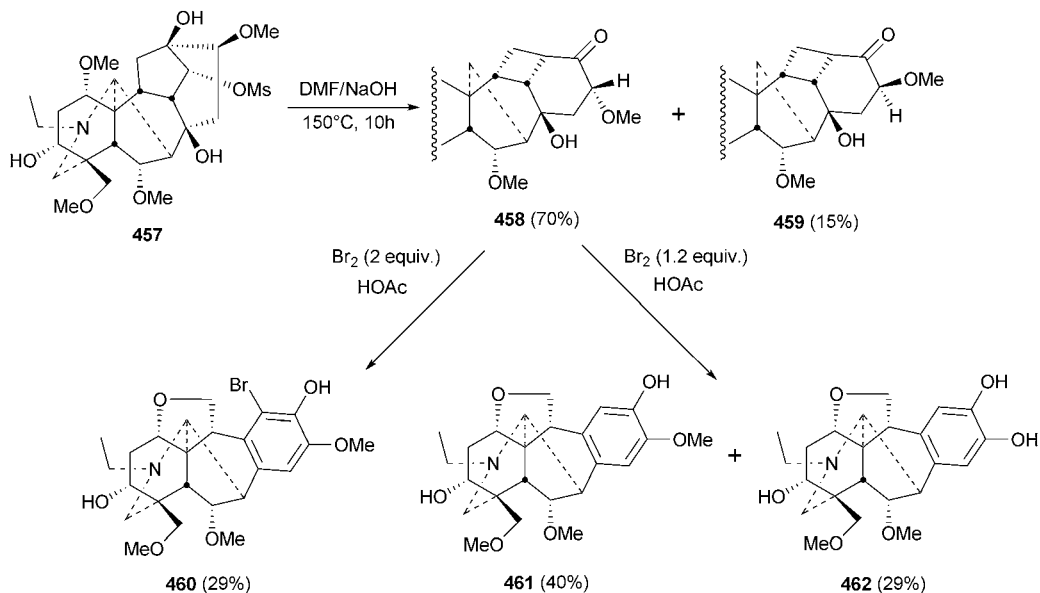
Nominine (**498**), a hetisine-type diterpenoid, was first isolated from *Aconitum sanyoense* by Ochiai *et al.* in 1956.<sup>257</sup> In 2004, the total synthesis of ( $\pm$ )-nominine was completed by Muratake and Natsume in 40 steps and 0.15% overall yield.<sup>19,258–260</sup> The key steps in the construction of the architecturally complex polycyclic structure of this diterpenoid alkaloid were a palladium-catalyzed intramolecular  $\alpha$ -arylation at the aldehyde group,



**Scheme 7** Cleavage of the C-7-C-17 bond of compounds **447** and **449**.



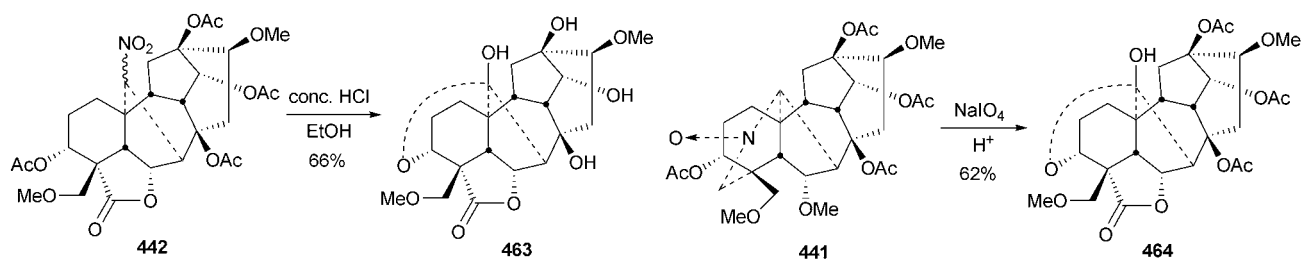
**Scheme 8** Rearrangement of ring A of compound **454**.



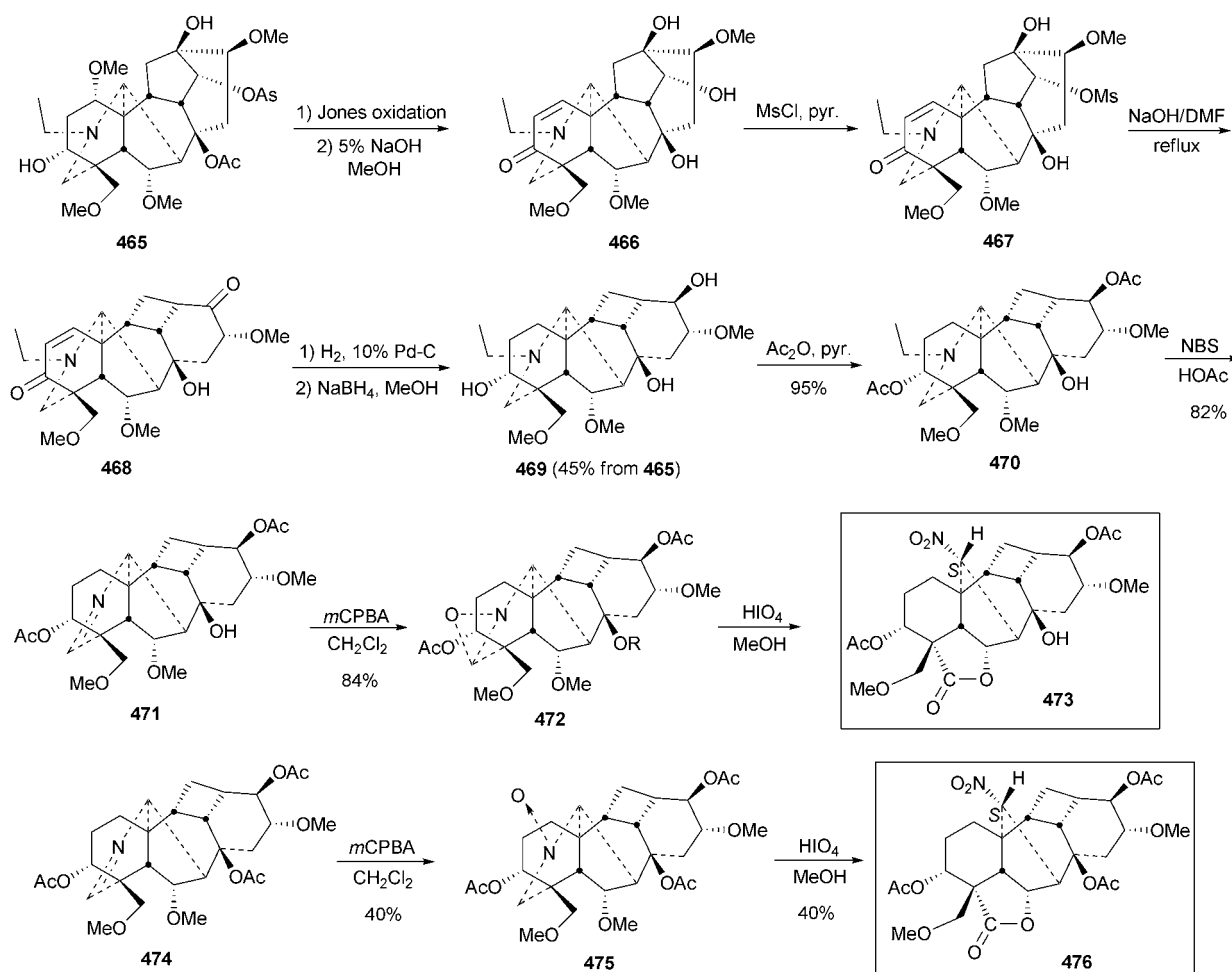
**Scheme 9** Synthesis of 12,13-seco diterpenoid alkaloids.

a Lewis acid-mediated acetal-ene reaction to form the C-14-C-20 bond, and a radical cyclization of an enyne. As depicted in Scheme 13, aldehyde **484**, prepared from 1-bromo-2-(2-iodoethyl)-4-methoxybenzene, could be readily converted to the desired isomer **485** via palladium-catalyzed intramolecular  $\alpha$ -arylation and repeated acetalization. Following the

transformation of **485** into **487**, the C-14-C-20 bond was constructed by an intramolecular acetal-ene reaction to form tetracycle **488**. After obtaining the enone **489** in an eight-step sequence from **488**, stereoselective hydrocyanation of **489** with  $\text{Et}_2\text{AlCN}$  afforded the *trans* isomer **490** with the desired C-4 stereochemistry.



Scheme 10 Conversion of diterpenoid alkaloids into aconane-type diterpenes.



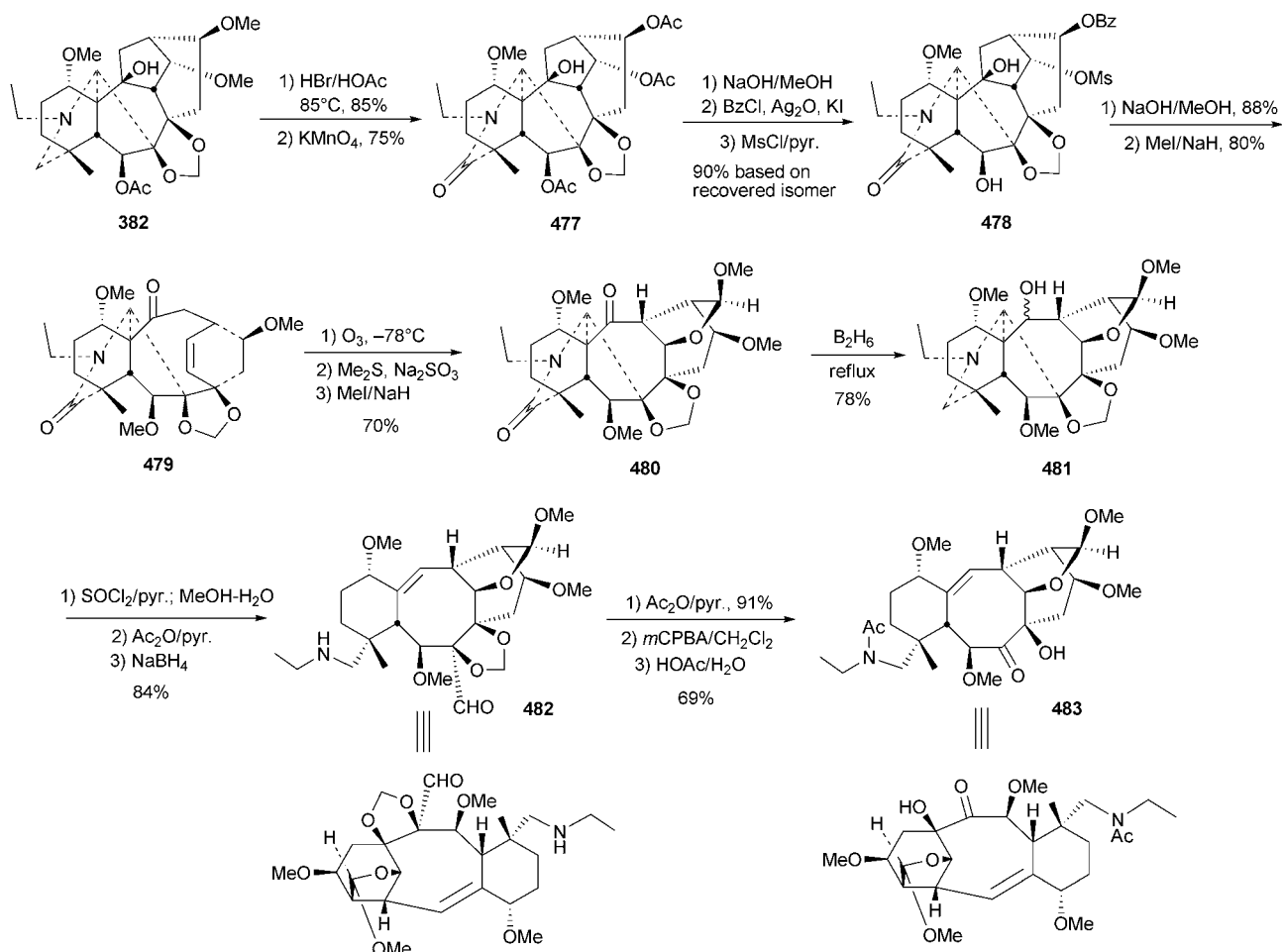
Scheme 11 Synthesis of 473 and 476.

Due to the failure of D ring construction at a later stage,<sup>261</sup> the authors turned to a new strategy to complete the end-game of the synthesis (Scheme 14).<sup>19</sup> The pyrrolidine ring in pentacyclic intermediate **491** was constructed in a three-step sequence from **490** in 63% yield. Then compound **491** was converted *via* Ohira alkylation to enyne precursor **493**, which was subjected to radical cyclization to yield the hexacyclic intermediate **494** in 57% yield. Eventually, the construction of ( $\pm$ )-nominine (**498**) was completed by selective introduction of a 15 $\beta$ -hydroxyl group and the azabicyclic ring.

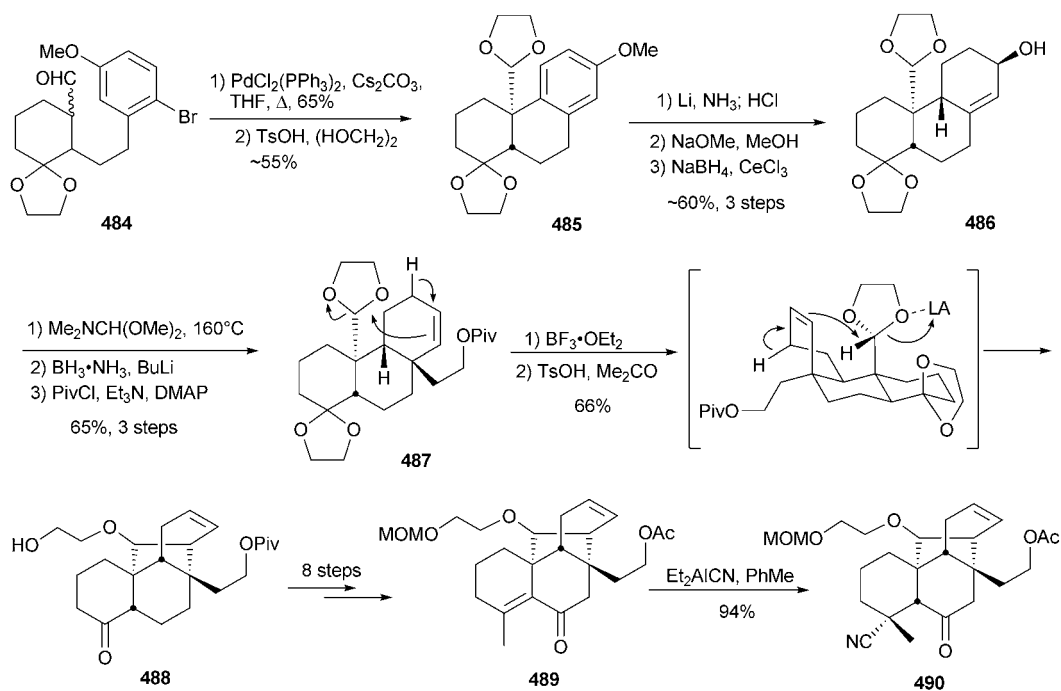
**4.5.2 Gin and Peese's synthesis of nominine.** Following the above-mentioned landmark achievement of Muratake and

Natsume, a short total synthesis of ( $\pm$ )-nominine in a 15-step sequence was accomplished by Gin and Peese two years later.<sup>20</sup> This elegant route features a reversible intramolecular 4-oxidoisoquinolinium betaine 1,3-dipolar cycloaddition (**506**  $\rightarrow$  **508**) as well as a pyrrolidine-induced dienamine isomerization–Diels–Alder cascade (**511**  $\rightarrow$  **513**).

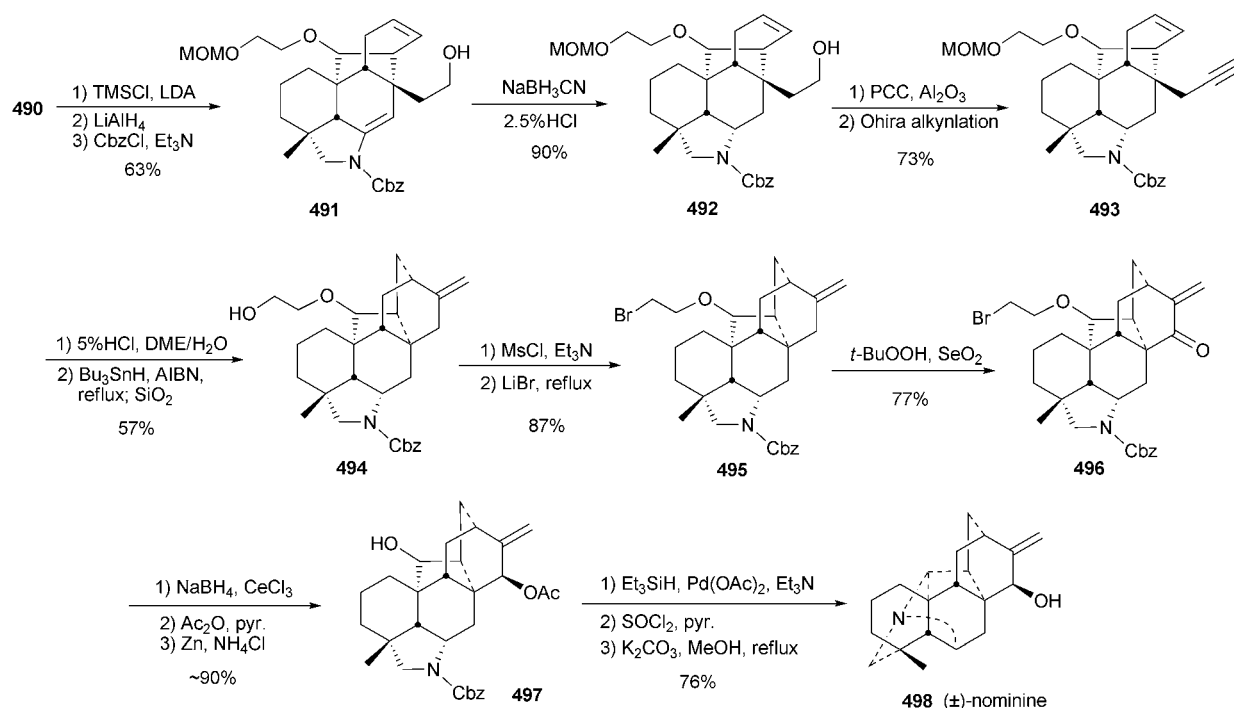
As shown in Scheme 15, the synthetic efforts commenced with the preparation of a substrate incorporating the requisite dipole-dipolarophile complement in conjunction with a latent diene-dienophile pair. Aryl ketone **500** was synthesized in 52% yield by *ortho*-lithiation of *p*-anisaldehyde dimethyl acetal (**499**), followed by nucleophilic addition to 2-chloro-*N*-methoxy-*N*-methylacetamide. Cyclic bis-acetal **501** was achieved by the replacement



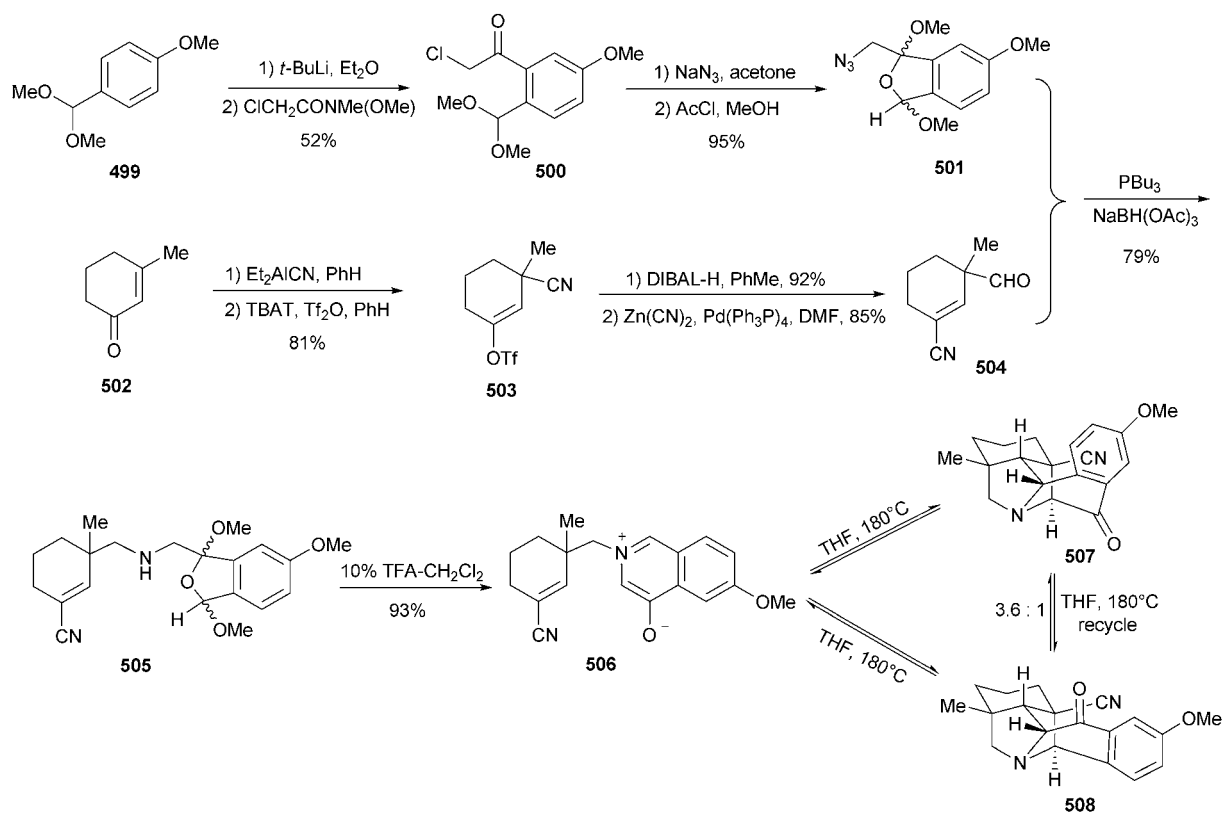
**Scheme 12** Conversion of deltaline (382) to taxane ABC core system.



**Scheme 13** Synthesis of intermediate 490.



**Scheme 14** The first total synthesis of (±)-nominine by Muratake and Natsume.



**Scheme 15** Synthesis of key intermediate **508**.

of the  $\alpha$ -chloro substituent in **500** with its  $\alpha$ -azido counterpart and acid-catalyzed rearrangement. Meanwhile, the ene–nitrile dipolarophile **504** was prepared from **502** in 63% total yield by cyanation, enolate trapping, and Pd<sup>0</sup>-catalyzed cross-coupling

with Zn(CN)<sub>2</sub>. Staudinger–aza-Wittig coupling of **501** with **504** followed by reduction of the corresponding imine afforded amine **505** (79%) as a mixture of four diastereomers. The mixture was then converted to 4-oxidoisquinolinium betaine **506** (93%),

which served as a suitable aza-1,3-dipole, *via* TFA-catalyzed MeOH extrusion and isomerization. The key intramolecular 1,3-dipolar cycloaddition of **506** at 180 °C provided a separable mixture of pyrrolidine isomers **507** and **508** with 97% conversion rate. The isomer **507** could be reconverted to the desired cycloadduct **508** through thermal equilibration.

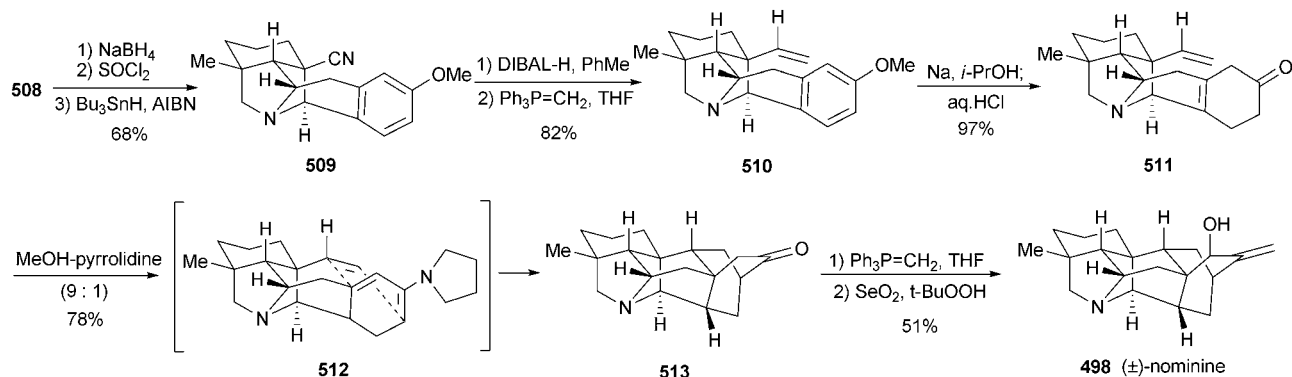
The  $\beta,\gamma$ -unsaturated cyclohexenone **511** was achieved from **508** in a six-step sequence (Scheme 16). With compound **511** in hand, the stage was set for the intramolecular Diels–Alder cycloaddition. Upon exposure of **511** to pyrrolidine in MeOH at 60 °C, the adduct **513** was generated in 78% yield, *via* the presumed dienamine intermediate **512**. The end game of the synthesis toward ( $\pm$ )-nominine (**498**) was completed by Wittig methylenation of the ketone **513** followed by diastereoselective SeO<sub>2</sub> allylic hydroxylation.

The first asymmetric total synthesis of (+)-nominine was also completed by the Gin group through an early-stage introduction of the desired chiral centers.<sup>262</sup> The ene–nitrile (+)-**504** served as a key chiral material, which was synthesized using a novel asymmetric conjugate addition methodology developed by Hoveyda *et al.*<sup>263</sup> This enantioselective transformation was applicable in the context of multi-step synthesis, culminating in the asymmetric synthesis of (+)-nominine.

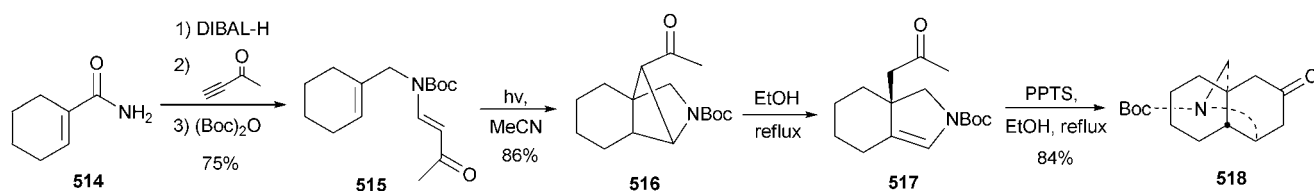
## 4.6 Syntheses directed towards hetisine-type C<sub>20</sub>-diterpenoid alkaloids

### 4.6.1 Synthetic studies towards hetisine by Kwak and Winkler.

Kwak and Winkler have disclosed a route to the bridged pyrrolidine **518** from **514**, employing a highly efficient intramolecular [2 + 2]-photocycloaddition of a vinylogous imide as a key reaction (**515** → **516**) (Scheme 17).<sup>264</sup> This methodology might be applicable to the synthesis of hetisine alkaloids.



Scheme 16 Completion of the total synthesis of ( $\pm$ )-nominine by Gin and Peese.



Scheme 17 Synthesis of intermediate **518** by Kwak and Winkler.

### 4.6.2 Synthetic studies towards hetisine by the Williams group.

The advanced intermediate **525** with an ABCE C<sub>20</sub>-diterpenoid ring system was prepared from **519** and **520** by Williams and co-workers, employing an intramolecular bridgehead arylation as a key reaction (Scheme 18).<sup>265,266</sup> Deprotonation of arylacetylene **520** with methylmagnesium bromide and subsequent addition to **519** in toluene afforded **521**. Catalytic hydrogenation of **521** gave the *cis*-alkene **522** in 74% yield. Elaboration of **522** using routine procedures afforded **523** and **524**. Intramolecular arylation of **524** with silver 2,4,6-trinitrobenzenesulfonate (AgTNBS) produced the target compound **525** in 53% yield.

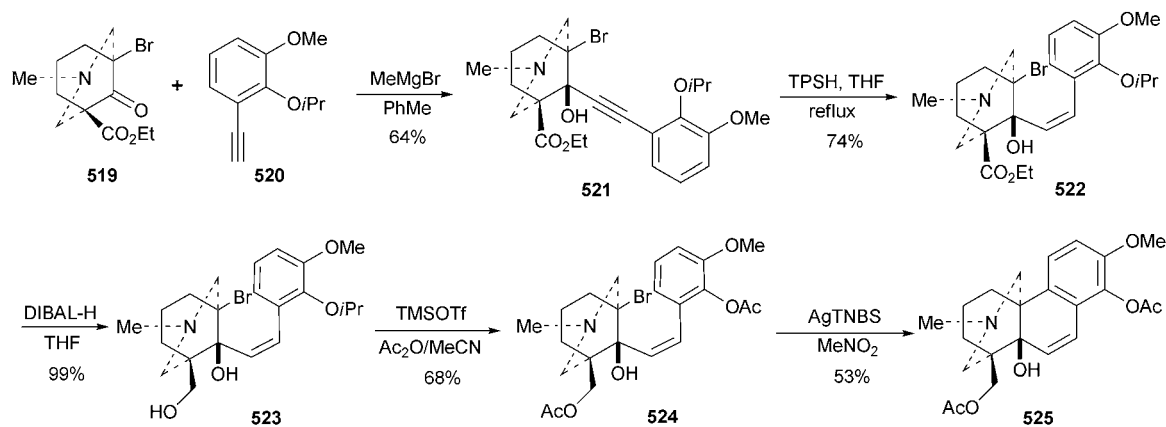
### 4.6.3 Synthetic studies towards nominine by Hutt and Mander.

Recently, attempts to synthesize nominine (**498**) by Hutt and Mander led to the preparation of the key intermediate **537** (Scheme 19).<sup>267</sup> The alcohol **527** was prepared in 27% overall yield from **526** in a six-step sequence. Protection of **527** using standard conditions followed by removal of the ketal functionality afforded the enone **528** in 42% yield, which was smoothly converted to the  $\beta$ -keto ester **529**. Treatment of **529** with NaH/MOMCl in HMPA followed by reduction delivered the alcohol **531**, which was converted into the nitrile **532** in three steps – oxidation, conversion of the aldehyde to the oxime, and dehydration in the presence of RuCl<sub>2</sub>(*p*-cymene)<sub>2</sub>. The construction of the key intermediate **537** was then achieved by a sequence of reactions involving alkylation (**532** → **533**), Birch reduction (**534** → **535**), DDQ oxidation (**535** → **536**), and intramolecular Lewis acid-catalyzed 1,6-addition (**536** → **537**).

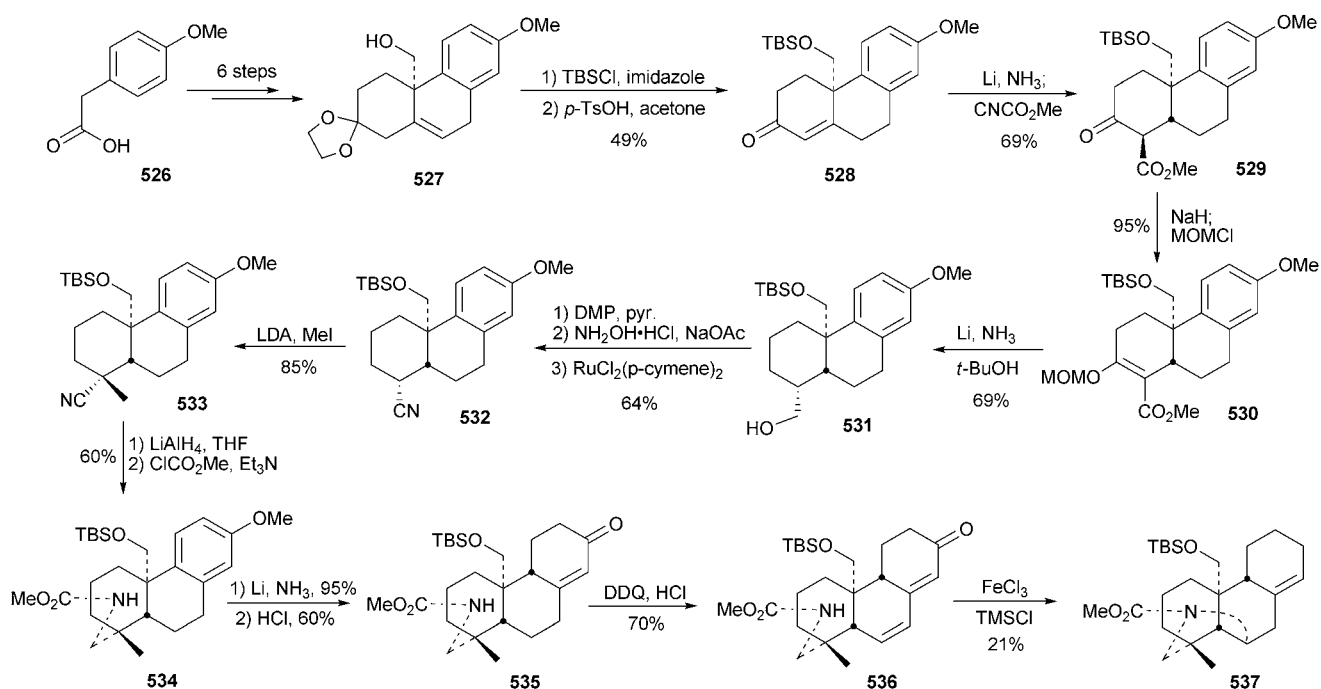
## 4.7 Synthetic studies towards C<sub>19</sub>-diterpenoid alkaloids

### 4.7.1 Synthetic studies towards *N*-deacetylappaconitine.

Taber *et al.* presented a model study leading to the preparation of the AEF rings of *N*-deacetylappaconitine, using conjugate



Scheme 18 Synthesis of ABCE ring system of hetisines by Williams.



Scheme 19 Synthetic studies towards nominine by Hutt and Mander.

addition and Mannich cyclization as key reactions (Scheme 20).<sup>268</sup> The synthesis of the substituted cyclohexanone intermediate **539** was successfully accomplished using an alkylidene carbene C–H insertion–ozonolysis–aldol condensation sequence. The conjugate addition to the **539** proceeded with high diastereoselectivity. The key intermediate **543** was constructed by Mannich cyclization of the intermediate **542** in the presence of Rexyn-300 and Na<sub>2</sub>SO<sub>4</sub>.

**4.7.2 Synthetic studies towards aconitine.** Conrad and Du Bois described a strategy for preparation of the BCD ring system of aconitine during their attempts to exploit chemoselective C–H amination for assembling the complex framework of natural products.<sup>269</sup> As shown in Scheme 21, treatment of sulfamate **544** with Rh<sub>2</sub>(esp)<sub>2</sub>, PhI(OAc)<sub>2</sub> and MgO furnished the corresponding *N,O*-acetal **545** in high yield and with exquisite

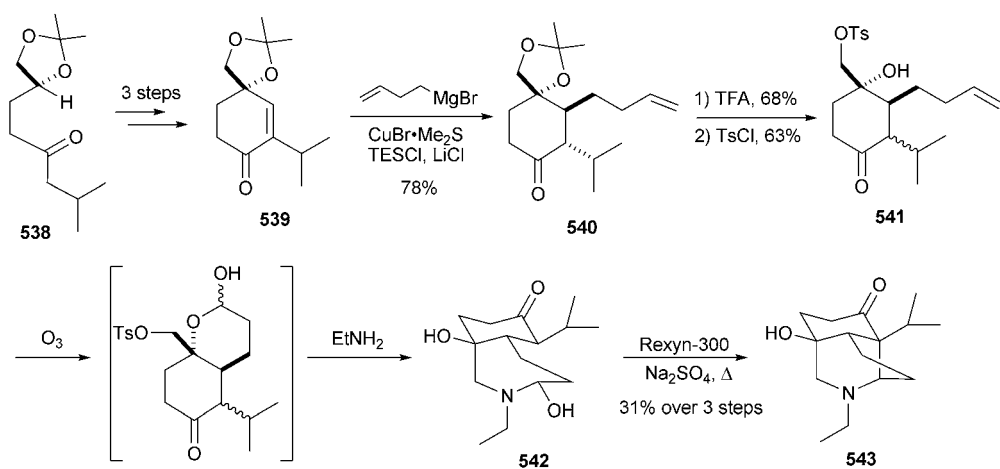
chemoselectivity. Subsequent Lewis acid-mediated cyclization failed to obtain the desired **546**; instead, the unexpected rearranged product **547** with a BCD ring system of typical C<sub>19</sub>-diterpenoid alkaloids was isolated.

**4.7.3 Synthesis of methyllycaconitines.** Since Blagbrough *et al.* reported that methyllycaconitine is a potential nAChR inhibitor,<sup>270</sup> the synthesis of methyllycaconitine and its analogues have attracted a lot of interest. An excellent review on the synthesis of methyllycaconitine and its analogues has been presented.<sup>12</sup>

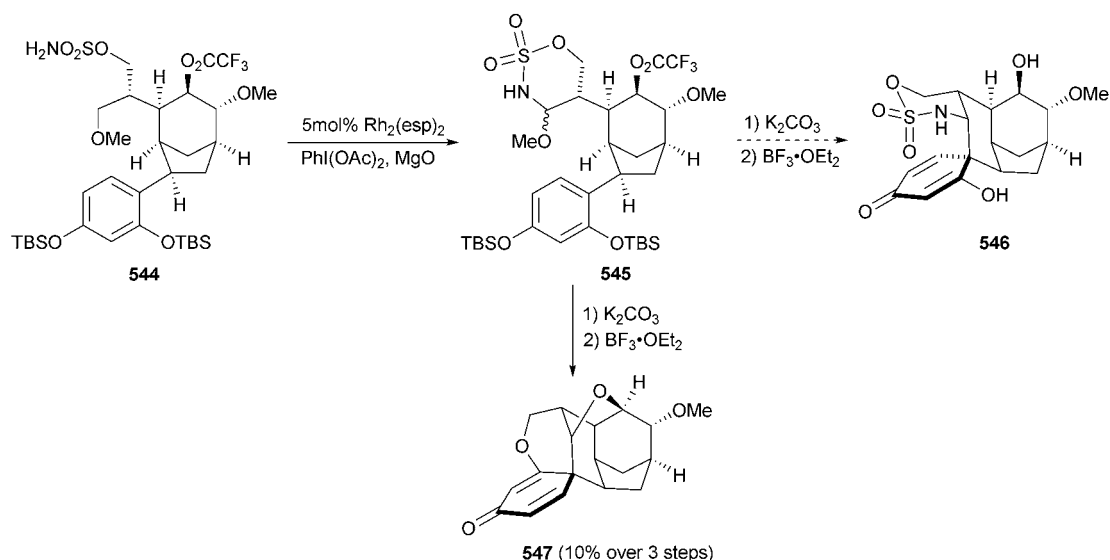
## 5 Studies of soft-ionization MS (ESI, ESI-MS<sup>n</sup>)

Since it was established in 1997 by Ohta *et al.* that soft-ionization mass spectrometry makes it possible to detect therapeutic levels





**Scheme 20** Construction of AEF rings of C<sub>19</sub>-diterpenoid alkaloids by Taber *et al.*



**Scheme 21** Synthetic efforts towards aconitine by Du Bois.

of diterpenoid alkaloids in blood and urine,<sup>271</sup> various soft-ionization techniques and their combinations with other techniques have been extensively applied to determine diterpenoid alkaloids in mixed samples (including extracts and metabolites).<sup>243–245,272–289</sup> This technique has been found to be superior to other analytical methods with regard to sample amount, speed, sensitivity, and convenience.

### 5.1 Aconitine-type alkaloids

“*Chuan wu*” (the roots of *Aconitum carmichaeli*), “*fu zi*” (radix of *Aconitum lateralis*), and “*cao wu*” (the roots of *Aconitum kusnezoffii*) have been widely used in clinical therapy as Traditional Chinese Medicines. They all have toxic and bioactive diester-type diterpenoid alkaloids, such as aconitine, mesaconitine, hypaconitine, deoxyaconitine, beiwutine and aconifine, together with their derivatives, lipo-alkaloids. Structurally, all of these alkaloids possess a hydroxyl group at C-15 $\alpha$ , a benzoyl group at

C-14, and an acetoxy or a long-chain fatty acid ester at C-8. Liu *et al.* have extensively investigated the analysis of diterpenoid alkaloids, including pure alkaloid or mixed samples (especially for the alkaloids from “*chuan wu*”, “*fu zi*”, and “*cao wu*”) by ESI-MS.<sup>243–245,272,274–279,283,284</sup>

The most important fragmentation features in the ESI-MS<sup>n</sup> of this kind of diterpenoid alkaloids are that the characteristic ion peaks come from the cleavage of substituents (such as OH, OMe, OAc, OBz, NMe, NEt); in a very few cases ring fragmentation is involved.

In addition, on the basis of the calculation of the stability of fragment ions by quantum chemistry, Chen *et al.* discovered that the cleavage order of the substituents in the ESI-MS<sup>n</sup> was as follows: 8-OAc  $\rightarrow$  14-OBz  $\rightarrow$  NET/NMe and that the methoxy group located at C-16 would be lost more readily than those at other positions (such as C-1, C-6 and C-18).<sup>290</sup>

ESI-MS<sup>n</sup> characteristics of three aconitine-type alkaloids, including isotalatizidine, neoline and senbusine A, have been

presented. The main fragments in ESI-MS<sup>n</sup> spectra of these alkaloids originate from the loss of H<sub>2</sub>O (–18 u), MeOH (–32 u), and HOAc (–60 u). The relative abundance of these fragments is influenced by the nature and position of the substituents.<sup>291</sup>

The lycoctonine-type alkaloid virescine possesses an  $\alpha$ -OH at C-6, while the aconitine-type alkaloid sensbusine A has a  $\beta$ -OH at C-7. The relative intensity of the fragments at  $m/z$  370 [P – 3H<sub>2</sub>O]<sup>+</sup> in the MS<sup>2</sup> of sensbusine A, as well as of the ions at  $m/z$  388 [P<sub>2</sub> – H<sub>2</sub>O]<sup>+</sup> and at  $m/z$  370 [P<sub>2</sub> – 2H<sub>2</sub>O]<sup>+</sup> in its MS<sup>3</sup>, is significantly higher than those of virescine. This suggests that the elimination of the hydroxyl group occurs more readily at C-6 than at C-7.<sup>291</sup>

## 5.2 Lipo-alkaloids

Most of the lipo-alkaloids belong to the aconitine-type group. The characteristic ion peak at [M + H – OR/C-8]<sup>+</sup> of the lipo-alkaloids in the MS<sup>2</sup> spectra is usually derived by the loss of the long-chain fatty acid ester at the C-8 position.<sup>272–274,277,278</sup> Except for this point, the other fragmentation patterns in their MS<sup>n</sup> spectra are similar to those of aconitine-type alkaloids.

Employing various ESI-MS<sup>n</sup> techniques, a few dozen lipo-alkaloids have been detected and determined from “*chuan wu*”, “*fu zi*”, and “*cao wu*”.<sup>271,274,276,279,280</sup>

## 5.3 Lycoctonine-type alkaloids

On the basis of careful analysis of fragmentation patterns in the mass spectra of some lycoctonine-type diterpenoid alkaloids by Gardner *et al.*,<sup>274</sup> it was found that the first two tandem mass experiments of lycoctonine-type alkaloids yield a significant fragment ion peak at [M + H – H<sub>2</sub>O]<sup>+</sup> from loss of water, or at [M + H – HOAc]<sup>+</sup> due to loss of acetic acid. It was also shown that the first three tandem mass experiments produce most abundant ions at (MH<sup>+</sup> – 32) derived from sequential losses of methanol, while MS<sup>4</sup> shows principal losses of water, methanol, and methanol–water. The generation of fragmentation ion peaks of the lycoctonine-type alkaloids is closely correlated with the pattern and position of substituents.

## 5.4 The stereochemistry of diterpenoid alkaloids

The Japanese scientists Wada *et al.* successfully applied HPLC–APCI-MS to the stereochemical investigation on C<sub>19</sub>-diterpenoid alkaloids.<sup>292</sup> It was shown that the abundance of fragment ions is significantly higher for C-1 $\beta$ -type alkaloids than for C-1 $\alpha$ -type alkaloids. The characteristic fragment ions are formed by the loss of a molecule of water, acetic acid or methanol at C-8. The reason for this kind of difference might be attributed to the stability of [M + H]<sup>+</sup>.<sup>293</sup> This method was also successfully applied to C-6 stereoisomeric C<sub>19</sub>-diterpenoid alkaloids by Wada and coworkers.<sup>293</sup> For the diterpenoid alkaloids with 6-OMe/8-OH moieties, comparison of the APCI spectra showed that the abundance of fragment ions at [M + H – H<sub>2</sub>O]<sup>+</sup> is significantly higher for C-6 $\beta$  alkaloids than for C-6 $\alpha$  alkaloids. However, for those with 6-OMe/8-OAc moieties, the abundance of fragment ions at [M + H – H<sub>2</sub>O]<sup>+</sup> is not correlated with its configuration at all.<sup>292</sup>

Similarly, HPLC–APCI-MS was also successfully applied to seven napelline-type stereoisomeric diterpenoid alkaloids at C-1

or C-12. The APCI spectra of alkaloids consist predominantly of the [M + H]<sup>+</sup> ion, the major fragment ion corresponding to the [M + H – H<sub>2</sub>O]<sup>+</sup> ion or the [M + H – CH<sub>3</sub>COOH]<sup>+</sup> ion. Comparison of the APCI spectra shows that the abundance of fragment ions is significantly higher for C-1 $\beta$ -type alkaloids than for C-1 $\alpha$ -type alkaloids, and for C-12 $\beta$ -type alkaloids than for C-12 $\alpha$ -type alkaloids.<sup>294</sup>

In addition, there are some other reports on the application of GC/SIM,<sup>295</sup> HPCE,<sup>296</sup> HPLC,<sup>297–301</sup> countercurrent chromatography,<sup>302–304</sup> and LC<sup>305</sup> for the analysis and separation of diterpenoid alkaloids.

## 6 Phytotaxonomic studies

During the review period, only one paper on the phytotaxonomic characteristics of diterpenoid alkaloids was presented, by Xiao, Wang *et al.*<sup>17</sup> This report described the taxonomic characteristics of diterpenoid alkaloids from Chinese *Aconitum*, and a reliable taxonomic character of the genus based on the reported diterpenoid alkaloids of 84 species of *Aconitum* grown in China.

## 7 Biological activities

### 7.1 Pharmacological activities

**7.1.1 Anti-inflammatory, analgesic, and de-addictive activities.** Shaheen and co-workers have evaluated the anti-inflammatory activities of six C<sub>19</sub>-diterpenoid alkaloids in an *in vitro* assay.<sup>113</sup> The results showed that only lappaconitine and puberanine exhibited anti-inflammatory activity as good as indomethacin. Russian scientists have investigated on the effects of water–ethanol extracts and alkaloids extracted from tall *Delphinium* on acute inflammation induced by carrageenin, acetic acid, serotonin and histamine.<sup>306</sup> The bioassay results show that these plant extracts and alkaloids could have significant anti-inflammatory activity, comparable to that of non-steroid anti-inflammatory drugs.

The Wang group has recently published their research results on the structure–analgesic activity relationship of 28 C<sub>18</sub>- and C<sub>19</sub>-diterpenoid alkaloids.<sup>307</sup> It was found that a tertiary amine in ring A, an acetoxy or an ethoxy group at C-8, an aromatic ester at C-14, and the saturation state of the ring D are structural features necessary for their analgesic activities. The analgesic activity of several alkaloids of *Aconitum* was assumed to be attributed to the interference with voltage-gated Na<sup>+</sup> channels.<sup>308</sup> Aconitine, 3-acetylaconitine and hypaconitine seem to inhibit neuronal conduction by persistent depolarization, whereas lappaconitine might block Na<sup>+</sup> channels. A QSAR analysis of the effect of chemical substitutes in the analgesic potency of 12 diterpenoid alkaloids, which were previously evaluated in a model of acetic acid-induced writhing in rats, corroborates that analgesic effect is primarily peripheral and secondarily central.<sup>309</sup>

It has been reported that *Delphinium denudatum*, a rich source of diterpenoid alkaloids, might significantly reduce the aggregate scores for all parameters in morphine withdrawal syndrome in rats or mice by central action, and thus may prove to be an alternative remedy in morphine de-addiction.<sup>310–312</sup>

**7.1.2 Cardiovascular action.** Pelletier and co-workers have tested the *in vivo* cardiovascular action (hypotensive,

bradycardic, and ventricular arrhythmias) of 13 C<sub>19</sub>-diterpenoid alkaloids in male Sprague-Dawley rats.<sup>313</sup> It was found that 3,8-diacetylfalconerine can cause arrhythmias at doses of 200 and 400 µg kg<sup>-1</sup>, and that the following compounds exhibit prominent hypotensive and bradycardic activity without prominent arrhythmias: heteratisine *N*-oxide, 8-deacetyl-8-*p*-aminobenzoyldelphinine, 8-deacetyl-8-anthranoyldelphinine, 8-stearoylfalconerine, 8-linolenylfalconerine, pyrodelphinine, 16-*epi*-pyroaconitine, and 8,9-(methylenedioxy)-lappaconitine. It was shown that cardiotoxicity of lappaconitine and *N*-deacetylappaconitine is much lower than aconitine, and that lappaconitine is a naturally occurring compound with class-I antiarrhythmic action.<sup>314</sup> It was established that several napelline-type alkaloids produce a more-or-less significant antiarrhythmic effect. Among them, 1-*O*-benzoylnapelline showed the most potent activity, which markedly exceeded that of napelline itself and the reference class-I antiarrhythmic drugs novocainamide, quinidine and lidocaine.<sup>315</sup> 12-Acetyl-12-*epi*-napelline was also found to have better antiarrhythmic activity than quinidine and novocainamide.<sup>209</sup> As mentioned in the introduction, guan-fu base A has been developed by Liu *et al.* for the therapeutic treatment of arrhythmia in China.<sup>16</sup>

**7.1.3 Anticancer activity.** The bioassay-guided fractionation of the MeOH extract of the roots of *Aconitum pseudo-laeve* var. *erectum* led to the isolation of lycaconitine as an active compound (IC<sub>50</sub> = 74 µg mL<sup>-1</sup>) toward multidrug-resistant human fibrocarcinoma KB V20C.<sup>316</sup> Lycaconitine was found to have potent inhibitory activity on Pgp-MDR but not on MRP-MDR. Later, 8-*O*-azeloil-14-benzoylaconine was found to have *in vitro* cytotoxicity, with an IC<sub>50</sub> value of about 10–20 µM.<sup>13</sup> Two more reports on the effects of the diterpenoid alkaloids on cancer cells have appeared in recent years.<sup>317,318</sup> Several C<sub>19</sub>-diterpenoid alkaloids (such as neoline, pubescenine, 14-deacetylajadine, lycoctonine, dehydrotakaosamine and ajadelphinine) exhibited selective cytotoxicity to cancerous cells, and some of these had irreversible effects on SW480, HeLa and SkMel25 cell lines.<sup>319</sup> These cytotoxic effects were related to the inhibition of ATP production. Interestingly, it was reported that the C<sub>19</sub>-diterpenoid alkaloids showed a lower suppression effect against certain human tumor cell lines (such as A172, A549, HeLa and Raji) relative to the C<sub>20</sub>-diterpenoid alkaloids.<sup>320</sup>

**7.1.4 Anti-epileptiform activity.** It was reported that mesaconitine shows significant anti-epileptiform activity, which becomes obvious in the low Mg<sup>2+</sup>-model and is not blocked by the β-adrenoceptor antagonist timolol, but is blocked instead by the α-adrenoceptor antagonist yohimbine.<sup>321</sup> These findings indicate that noradrenergic inhibitory actions in the CA1 and CA3 subfield of the hippocampus are involved in the anti-epileptiform action of diterpenoid alkaloid mesaconitine, which are likely to be α-receptor-mediated. The alkaloid 6-benzoylheteratisine was also reported to be able to exert an inhibitory and anti-epileptiform effect in hippocampal slices by antagonizing the activation of Na<sup>+</sup> currents by aconitine.<sup>322</sup> The effect is likely to be mediated by a direct or indirect interaction at the neurotoxin binding site 2 of the voltage-dependent Na<sup>+</sup> channel. A subfraction (FS-1, 600 mg kg<sup>-1</sup> i.p.) from the aqueous fraction of the roots of *Delphinium denudatum* exhibits comparable

anticonvulsant activity in CF1 mice to that of phenytoin (the well-known anti-epileptic drug, 20 mg kg<sup>-1</sup>) in the maximal electroshock test, and it also protected 100% of animals from the hind limb tonic extension phase of this model.<sup>323</sup> Further studies suggest that FS-1 can block sustained repetitive firing in hippocampal neurons in a use-dependent and voltage-dependent manner similar to phenytoin (the prototype anticonvulsant drug).<sup>324</sup> The blockade of sustained repetitive firing is one of the basic mechanisms of anti-epileptic drugs at the cellular level.

**7.1.5 Antiparasite and insect repellent.** The diterpenoid alkaloids cardiopetamine and 15-acetylcardiopetamine were found to strongly inhibit the feeding activity of *Spodoptera littoralis* and *Leptinotarsa decemlineata*, respectively.<sup>325</sup> Structure–activity studies with *S. littoralis* showed that the C-13 and C-15 hydroxyl groups are essential features of the active molecule, while a C-13 hydroxyl and/or a C-15 acetate might determine their effect on *L. decemlineata*. The C-11 benzoate group may enhance the biological effect on both insect species. The repellent activities against *Tribolium castaneum* of 29 C<sub>19</sub>- and C<sub>20</sub>-diterpenoid alkaloids were assessed.<sup>326</sup> Twenty-one alkaloids among them showed promising insect repellent activity, with hetisine as the most potent compound (59.37%). The Spanish scientist González-Coloma and co-workers tested the insect antifeedant and toxic activity of 21 C<sub>20</sub>-diterpenoid alkaloids and 43 C<sub>19</sub>-diterpenoid alkaloids on *S. littoralis* and *L. decemlineata*.<sup>319,327,328</sup> The antifeedant effects of the test alkaloids were structure- and species-dependent. Overall, C<sub>19</sub>-diterpenoid alkaloids are better insect antifeedants and post-ingestive toxicants than the related C<sub>20</sub>-diterpenoid alkaloids. The most active antifeedants to *L. decemlineata* are 1,14-diacetylcardiopetaline and 18-hydroxy-14-*O*-methylgadesine (C<sub>19</sub>-diterpenoid alkaloid, EC<sub>50</sub> < 1 µg cm<sup>-2</sup>) and the rearranged form of hetisine (C<sub>20</sub>-diterpenoid alkaloid, EC<sub>50</sub> = 1.7 µg cm<sup>-2</sup>), while 19-oxodihydroatisine (EC<sub>50</sub> = 0.1 µg cm<sup>-2</sup>) is the most active against *S. littoralis*. The *in vitro* anti-proliferative effects were described for several atisine-type diterpenoid alkaloids against the protozoan parasite *Leishmania infantum*.<sup>329</sup> From a total of 43 compounds tested, including several classes of C<sub>19</sub>- and C<sub>20</sub>-diterpenoid alkaloids, only 15,22-*O*-diacetyl-19-oxodihydroatisine, azitine and isoazitine are highly active against cultures of the parasite (promastigote form), with IC<sub>50</sub> values within the range of the reference drug pentamidine-isothionate (7.39–12.80 mg L<sup>-1</sup> for the test compounds, 11.32 mg L<sup>-1</sup> for the positive control). González-Coloma and co-workers also screened anti-*Trypanosoma cruzi* activities of 64 C<sub>19</sub>- or C<sub>20</sub>-diterpenoid alkaloids.<sup>330</sup> It was found that five C<sub>20</sub>-diterpenoid alkaloids are active against *T. cruzi* epimastigotes: azitine, isoazitine, and 15,22-*O*-diacetyl-19-oxodihydroatisine have moderate effects on the parasite, while atisinium chloride and 13-oxocardiopetamine are potent *T. cruzi* epimastigote growth inhibitors, with activity levels similar to that of the reference drug benznidazole.

**7.1.6 Effects on cutaneous blood flow in mice.** Continuing their investigation on the effects of C<sub>20</sub>-diterpenoid alkaloid kobusine and pseudokobusin derivatives on cutaneous blood flow in mice, the Wada group established that the hydroxyl groups of these alkaloids, especially a free hydroxyl group at C-6,

are important for action on the peripheral vasculature leading to dilatation, and that the alkaloids with a 15-aromatic ester group (e.g. acetate, benzoate, anisoate, veratroate or *p*-nitrobenzoate) may enhance their activities relative to the parent alkaloids.<sup>331,332</sup>

**7.1.7 Other bioactivities.** Certain diterpenoid alkaloids were reported to have antibacterial activity,<sup>33,333</sup> antiviral activity,<sup>334</sup> or antioxidant ability.<sup>113</sup> Bulleyaconitine A was reported to display long-acting local anesthetic properties *in vitro* and *in vivo*.<sup>335</sup> The screening of various structural types of diterpenoid alkaloids and their derivatives for local anesthetic activity identified 26 compounds with distinct activity for surface anesthesia in rabbit eye cornea. Of these, 15 compounds have activities and durations of action that are greater than those of cocaine. However, their rates of onset of anesthesia are slower than that of cocaine.<sup>336</sup> The exceptions are 1-*O*-benzoylnappeline and tadzhaconine, which have similar rates of onset and durations of anesthesia, but higher activities, as compared with cocaine.

## 7.2 Toxicity

*Aconitum* species have been used since ancient times as poisons for spears and arrows, and later in homicides. *Aconitum* alkaloids (diterpenoid alkaloids) and especially aconitine (a C<sub>19</sub>-diterpenoid alkaloid) are neurotoxins that cause conduction block and paralysis by activating voltage-dependent sodium channels. Aconitine poisoning occurs mainly because of confusion with an edible plant,<sup>337</sup> deliberate or accidental ingestion by children,<sup>297</sup> and ingestion for a desired phytotherapeutic effect.<sup>338</sup> It was reported that the toxicity of *A. coreanum* is mainly due to alkaloids that have myorelaxant activity based on the experimental data, and the myorelaxant activity for hetisine, atisine and coryphine diterpenoid alkaloids was found for the first time.<sup>339</sup>

Larkspurs (*Delphinium* spp.) are toxic plants that are responsible for the majority of cattle deaths in western North America. Cattle deaths may exceed 15% in areas where larkspurs are abundant.<sup>340</sup> C<sub>19</sub>-diterpenoid alkaloids occur as prominent constituents of poisonous larkspurs in one of two structural types: the lycoctonine and the 7,8-methylenedioxylycoctonine (MDL) types. Of the 40 different diterpenoid larkspur alkaloids, the one that is thought to be responsible for much of the toxicity was identified as methyllycaconitine (MLA). MLA is a potent neuromuscular blocker, causing paralysis and rapid death from respiratory failure.<sup>341</sup> The study on the toxicokinetics of MLA excretion indicates that the MLA is rapidly distributed and excreted.<sup>342</sup> Four *N*-(methylsuccinimido)anthranoyllycoctonine C<sub>19</sub>-diterpenoid (MSAL) alkaloids – geyerline, grandiflorine, bearline and 14-acetylbearline – were recently reported to possess toxicity comparable to that of MLA.<sup>136,343</sup> Evaluation of the toxicity data establishes two structural features (an *N*-ethyl bicyclo-substituted tertiary alkaloid nitrogen atom and a C-18 anthranilic acid ester) to be necessary to impart toxicity to the lycoctonine-type C<sub>19</sub>-diterpenoid alkaloids. Additionally, the toxicity data establish that two other structural features (functionality at the anthranilic acid amine nitrogen and at C-14) can enhance that toxicity of the lycoctonine-type C<sub>19</sub>-diterpenoid alkaloids. Toxic alkaloid levels above 3 mg g<sup>-1</sup> pose a threat to grazing cattle. Diterpenoid alkaloid concentrations in larkspur plants vary with environment, plant and location.<sup>136,341,344,345</sup>

Alkaloids in tall larkspur appear to be synthesized during the first three to four weeks of early growth. Pools of alkaloids increase during this period, peak at four to six weeks, then decline as the alkaloids are apparently catabolized or translocated back to the roots.<sup>346</sup> Significantly, three competitive inhibition enzyme-linked immunosorbent assays (CIELISA) for toxic larkspur alkaloids were developed.<sup>347</sup> One assay is class-specific toward the MSAL alkaloids, and two assays are specific for individual alkaloids. It was reported that vaccinating mice with larkspur toxin–protein carrier conjugates appeared to provide a mild protective effect against MLA toxicity.<sup>348</sup>

QSAR analysis performed for the diterpenoid alkaloids showed that a linear relationship and high correlation coefficients were observed for: (1) LD<sub>50</sub> and analgesic activity ( $r = 0.96$ ); (2) LD<sub>50</sub> and local anesthetic activity ( $r = 0.71$ ); (3) toxicity and antiarrhythmic activity data ( $r = 0.88$ ); (4) toxicity and three therapeutic actions ( $r = 0.88–0.97$ ).<sup>309,349–353</sup> In addition, descriptors related to “drug-likeness” of molecules were selected to discriminate between “drugs” and “non-drugs” amongst diterpenoid alkaloids studied. A list of boundaries calculated for seven conventional “drug-likeness” parameters has shown that curariform and antiarrhythmic alkaloids are more drug-like compounds, while arrhythmogenic alkaloids are all likely to be classified as “non-drugs”. This is in a good agreement with the experimental data. When deciding whether a particular compound is a potential drug, a therapeutic index should also be considered. Thus, therapeutic indexes reported for some of “drug-like” diterpenoid alkaloids from the investigated series were in the range 100–230, mostly for those with benzoyl group. These findings also suggest that “drug-like” series of alkaloids should be further pursued as promising therapeutic agents.

## 7.3 Mechanisms of action

In 1998, Ameri summarized the effects of diterpenoid alkaloids on the central nervous system.<sup>3</sup> He subdivided the diterpenoid alkaloids into three subgroups based on the chemical features as well as their mechanisms of action: (1) diester diterpenoid alkaloids (with analgesic activity and high toxicity) that are able to activate voltage-dependent sodium channels even at resting potential and inhibit noradrenaline re-uptake; (2) monoester diterpenoid alkaloids (less toxic) which have been shown to possess strong antinociceptive, antiarrhythmic and anti-epileptiform properties due to a blockade of the voltage-dependent sodium channel; (3) de-esterified diterpenoid alkaloids which possess markedly reduced toxicity when compared with the two other groups, and fail to affect neuronal activity, but are reported to have antiarrhythmic actions. It was also demonstrated that 14-benzoyltalatisamine and talatisamine are strongly different in their mode of action even though their structures differ only in the presence or absence of a benzoyloxy group at the C-14 position.<sup>354</sup> It was shown that aconitine and 3-acetylaconitine, which are known to activate sodium channels, have comparable inhibitory potencies toward [<sup>3</sup>H]noradrenaline uptake.<sup>355</sup> In contrast, lappaconitine and *N*-desacetylappaconitine were found to fail to inhibit [<sup>3</sup>H]noradrenaline uptake. When either lappaconitine or *N*-desacetylappaconitine was applied in combination with aconitine, [<sup>3</sup>H]noradrenaline uptake was not affected. The

inhibitory and anti-epileptiform effect of ajacine and lappaconitine were shown to be mediated by a frequency-dependent inhibition of the voltage-dependent sodium channel.<sup>356</sup> Four diterpenoid alkaloids – songorine, 14-benzoyltalatisamine, pyrochasmaconitine and talatisamine – were shown to be able to depress markedly delayed rectifier K<sup>+</sup> current (*I<sub>K</sub>*) and fast transient K<sup>+</sup> current (*I<sub>A</sub>*) during the investigation using the whole-cell voltage-clamp recording in rat dissociated hippocampal neurons.<sup>357</sup> Very recently, it was further identified that talatisamine is a specific blocker for the delayed rectifier K<sup>+</sup> channel in rat hippocampal neurons.<sup>14</sup> An antagonist modulation of voltage-gated sodium channels exhibited by diterpenoid alkaloids from *Aconitum* and *Delphinium* species were investigated by means of two computational approaches: analysis of frontier MOs generated at B3LYP/6-31G(d, p) level and QSAR study.<sup>358</sup> The research results confirmed the experimental findings that neurotoxins acting at type 2 receptor site of voltage-dependent sodium channel are activators and blockers with common structural features and differ only in efficacy. Five alkaloids, methyllycaconitine, nudicauline, 14-deacetylnudicauline, barbinine and deltaline, were demonstrated to be likely nicotinic receptor antagonists that reduce synaptic efficacy and block neuromuscular transmission.<sup>359</sup> Songorine was suggested to be a novel non-competitive antagonist at the GABA<sub>A</sub> receptor in rat brain.<sup>15</sup>

MLA is an  $\alpha 7$ -type nAChR subtype selective antagonist.<sup>12,270,360</sup> Unlike the  $\alpha$ -conotoxins, MLA shows consistent affinity for the  $\alpha 7$  subtype across species. Unlike  $\alpha$ -Bgt, MLA can discriminate between muscular and neuronal nAChRs, and the association and dissociation kinetics are very rapid. MLA is a very potent antagonist, binding to  $\alpha 7$  nAChRs with nanomolar affinity. Although methyllycaconitine is itself too toxic for clinical therapeutic use as a drug in humans, several semi-synthetic efforts have been undertaken to increase the understanding of the structure–activity relationship. The synthesis of small analogues has also been undertaken in the search for less toxic but equally potent drugs.<sup>12,361,362</sup>

It was reported that lappaconitine and puberanine can exhibit mild inhibition against the tyrosinase.<sup>113</sup> Further investigation carried out by Sultankhodzhaev *et al.* on the tyrosinase inhibition of 15 lycocotone-type diterpenoid alkaloids and six napelline-type alkaloids led to the discovery that lappaconitine HBr is the most potent member of the series (IC<sub>50</sub> = 13.30  $\mu$ M), with a comparable inhibitory activity to that of kojic acid (IC<sub>50</sub> = 16.67  $\mu$ M).<sup>363</sup>

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## 9 References

- Atta-ur-Rahman and M. I. Choudhary, *Nat. Prod. Rep.*, 1999, **16**, 619.
- K. J. Hale and S. Manaviazar, in *Second Supplements to the 2nd Edition of Rodd's Chemistry of Carbon Compounds*, ed. M. Sainsbury, Elsevier Science, New York, 1998, vol. 4, part G (partial)/H, p. 1.
- A. Ameri, *Prog. Neurobiol.*, 1998, **56**, 211.
- X. W. Wang and H. Xie, *Drugs Future*, 1999, **24**, 877.
- B. S. Joshi and S. W. Pelletier, in *Alkaloids: Chemical and Biological Perspectives*, ed. S. W. Pelletier, Pergamon, New York, 1999, vol. 13, p. 289.
- B. S. Joshi, S. W. Pelletier and S. K. Srivastava, in *Alkaloids: Chemical and Biological Perspectives*, ed. S. W. Pelletier, Pergamon, New York, 2001, vol. 15, p. 1.
- I. A. Bessonova and Sh. A. Saidkhodzhaeva, *Khim. Prir. Soedin.*, 2000, **36**, 345.
- F. P. Wang and X. T. Liang, in *The Alkaloids: Chemistry and Biology*, ed. G. A. Cordell, Elsevier Science, New York, 2002, vol. 59, p. 1.
- C. S. Peng and F. P. Wang, *Acta Pharm. Sin.*, 2000, **35**, 932.
- P. A. Q. H. Chen and F. P. Wang, *Chem. Res. Appl.*, 2004, **16**, 173.
- L. Y. Lin, Q. H. Chen and F. P. Wang, *West China J. Pharm. Sci.*, 2004, **19**, 200.
- K. J. Goodall, D. Barker and M. A. Brimble, *Synlett*, 2005, **12**, 1809.
- A. Chodoeva, J. J. Bosc, J. Guillon, A. Decendit, M. Petraud, C. Absalon, C. Vitry, C. Jarry and J. Robert, *Bioorg. Med. Chem.*, 2005, **13**, 6493.
- M. K. Song, H. Liu, H. L. Jiang, J. M. Yue, G. Y. Hu and H. Z. Chen, *Neuroscience*, 2008, **155**, 469.
- X. Y. Zhao, Y. Wang, Y. Li, X. Q. Chen, H. H. Yang, J. M. Yue and G. Y. Hu, *Neurosci. Lett.*, 2003, **337**, 33.
- X. J. Yang, G. J. Wang, S. S. Ling, N. Y. Qiu, G. G. Wang, J. Zhu and J. H. Liu, *J. Chromatogr., B: Biomed. Sci. Appl.*, 2000, **740**, 273. See also the announcement: *Chin. J. Nat. Med.*, 2006, **4**, 1.
- P. G. Xiao, F. P. Wang, F. Gao, L. P. Yan, D. L. Chen and Y. Liu, *Acta Phytotaxon. Sin.*, 2006, **44**, 1.
- C. L. Zou, L. Cai, H. Ji, G. B. Xie, F. P. Wang, X. X. Jian, L. Song, X. Y. Liu, D. L. Chen and Q. H. Chen, *Tetrahedron*, 2008, **64**, 7594.
- H. Muratake and M. Natsume, *Angew. Chem., Int. Ed.*, 2004, **43**, 4646.
- K. M. Peese and D. Y. Gin, *J. Am. Chem. Soc.*, 2006, **128**, 8734.
- F. P. Wang, Q. H. Chen and X. T. Liang, in *The Alkaloids: Chemistry and Biology*, ed. G. A. Cordell, Elsevier Science, New York, 2009, vol. 67, p. 1.
- L. Cai, D. L. Chen and F. P. Wang, *Nat. Prod. Commun.*, 2006, **1**, 191.
- S. H. Jiang, H. Q. Wang, Y. M. Li, S. J. Lin, J. J. Tan and D. Y. Zhu, *Chin. Chem. Lett.*, 2007, **18**, 409.
- D. Csupor, P. Forgo, E. M. Wenzig, R. Bauer and J. Hohmann, *J. Nat. Prod.*, 2008, **71**, 1779.
- U. T. Teshebaeva, M. N. Sultankhodzhaev and A. A. Nishanov, *Khim. Prir. Soedin.*, 1999, **35**, 498.
- U. T. Teshebaeva, M. N. Sultankhodzhaev and A. A. Nishanov, *Khim. Prir. Soedin.*, 1999, **35**, 774.
- C. S. Peng, F. P. Wang, J. Z. Wang and X. X. Jian, *Acta Pharm. Sin.*, 2000, **35**, 201.
- U. T. Teshebaeva, M. N. Sultankhodzhaev and A. A. Nishanov, *Khim. Prir. Soedin.*, 1999, **35**, 811.
- F. Feng, W. C. Ye, J. H. Liu, S. X. Zhao, I. D. Williams and C. T. Che, *J. Chin. Pharm. Sci.*, 2000, **9**, 167.
- A. Ulubelen, A. H. Meriçli, F. Meriçli, U. Kolak, R. Ilarslan and W. Voelter, *Phytochemistry*, 1999, **50**, 513.
- F. P. Wang, C. S. Peng, X. X. Jian and D. L. Chen, *J. Asian Nat. Prod. Res.*, 2001, **3**, 15.
- J. J. Tan, C. H. Tan, B. Q. Ruan, S. H. Jiang and D. Y. Zhu, *J. Asian Nat. Prod. Res.*, 2006, **8**, 535.
- M. Ahmad, W. Ahmad, M. Ahmad, M. Zeeshan, Obaidullah and F. Shaheen, *J. Enzyme Inhib. Med. Chem.*, 2008, **23**, 1018.
- S. X. Zhang and S. S. Jia, *Chin. Chem. Lett.*, 1999, **10**, 133.
- S. X. Zhang and S. S. Jia, *Acta Pharm. Sin.*, 1999, **34**, 762.
- L. Song, X. X. Liang, D. L. Chen, X. X. Jian and F. P. Wang, *Chem. Pharm. Bull.*, 2007, **55**, 918.
- U. Kolak, M. Öztürk, F. Özgökçe and A. Ulubelen, *Phytochemistry*, 2006, **67**, 2170.
- P. M. Shrestha and A. Katz, *J. Nat. Prod.*, 2000, **63**, 2.
- A. Alva, M. Grandez, A. Madinaveitia, G. de la Fuente and J. A. Gavin, *Helv. Chim. Acta*, 2004, **87**, 2110.
- J. Li, D. L. Chen, X. X. Jian and F. P. Wang, *Molecules*, 2007, **12**, 353.
- C. S. Peng, D. L. Chen, Q. H. Chen and F. P. Wang, *Chin. J. Org. Chem.*, 2005, **25**, 1235.
- Z. S. Boronova and M. N. Sultankhodzhaev, *Khim. Prir. Soedin.*, 2000, **36**, 320.

- 43 J. Y. Dong, Z. Y. Li and L. Li, *Chin. Chem. Lett.*, 2000, **11**, 1005.
- 44 J. G. Diaz, J. G. Ruiza and W. Herz, *Phytochemistry*, 2005, **66**, 837.
- 45 L. He, Y. J. Pan, X. Pan, B. G. Li and Y. Z. Chen, *Chin. Chem. Lett.*, 1999, **10**, 395.
- 46 L. He, Y. J. Pan and Y. Z. Chen, *Indian J. Chem., Sect. B: Org. Chem. Incl. Med. Chem.*, 2001, **40**, 1285.
- 47 M. Reina, R. Mancha, A. Gonzalez-Coloma, M. Bailen, M. L. Rodriguez and R. A. Martinez-Diaz, *Nat. Prod. Res.*, 2007, **21**, 1048.
- 48 K. Zhang, L. He, X. Pan and Y. Z. Chen, *Planta Med.*, 1998, **64**, 580.
- 49 F. P. Wang, Z. B. Li, J. L. Wang, X. X. Jian and D. L. Chen, *Acta Chim. Sin.*, 2000, **58**, 576.
- 50 C. S. Peng, Q. H. Chen, D. L. Chen, X. X. Jian and F. P. Wang, *Heterocycles*, 2002, **57**, 1117.
- 51 X. L. Zhou, Q. H. Chen and F. P. Wang, *Chem. Pharm. Bull.*, 2004, **52**, 456.
- 52 Z. B. Li, L. Xu, X. X. Jian and F. P. Wang, *J. Asian Nat. Prod. Res.*, 2001, **3**, 131.
- 53 M. Grandez, A. Madinaveitia, J. A. Gavín, A. Alva and G. de la Fuente, *J. Nat. Prod.*, 2002, **65**, 513.
- 54 L. Cai, D. L. Chen, S. Y. Liu and F. P. Wang, *Chem. Pharm. Bull.*, 2006, **54**, 779.
- 55 A. Ulubelen, M. Arfan, U. Sönmez, A. H. Meriçli and F. Meriçli, *Phytochemistry*, 1998, **48**, 385.
- 56 L. Y. Li, Q. H. Chen, X. L. Zhou, D. L. Chen, F. P. Wang and C. T. Che, *J. Nat. Prod.*, 2003, **66**, 269.
- 57 S. H. Jiang, P. M. Yang, H. Zhou and D. Y. Zhu, *Planta Med.*, 2002, **68**, 1147.
- 58 G. B. Xie, Q. H. Chen, D. L. Chen, X. X. Jian and F. P. Wang, *Heterocycles*, 2003, **60**, 631.
- 59 G. B. Xie and F. P. Wang, *Chem. J. Chin. Univ.*, 2004, **25**, 482.
- 60 F. Gao, D. L. Chen and F. P. Wang, *Arch. Pharmacol Res.*, 2007, **30**, 1497.
- 61 F. Gao, Q. H. Chen and F. P. Wang, *J. Nat. Prod.*, 2007, **70**, 876.
- 62 F. Gao, D. L. Chen and F. P. Wang, *Chem. Pharm. Bull.*, 2006, **54**, 117.
- 63 F. Gao and F. P. Wang, *Heterocycles*, 2005, **65**, 365.
- 64 A. H. Meriçli, F. Meriçli, G. V. Seyhan, H. Özçelik, N. Kilinçer, A. G. Ferizli and A. Ulubelen, *Heterocycles*, 1999, **51**, 1843.
- 65 S. H. Shim, J. S. Kim, K. H. Son, K. H. Bae and S. S. Kang, *J. Nat. Prod.*, 2006, **69**, 400.
- 66 U. Kolak and M. Ulusoylu, *Sci. Pharm.*, 1998, **66**, 381.
- 67 D. L. Chen, X. X. Jian, Q. H. Chen and F. P. Wang, *Acta Chim. Sin.*, 2003, **61**, 901.
- 68 J. H. Yang, Z. Y. Li, L. Li and Y. S. Wang, *Phytochemistry*, 1999, **50**, 345.
- 69 H. Yan, D. L. Chen, X. X. Jian and F. P. Wang, *Helv. Chim. Acta*, 2007, **90**, 1133.
- 70 A. Ulubelen, A. H. Meriçli, F. Meriçli, H. Özçelik, B. Şener, H. Becker, J. Zapp, I. Choudhary and Atta-ur-Rahman, *Phytochemistry*, 1999, **50**, 909.
- 71 S. H. Shim, J. S. Kim and S. S. Kang, *Chem. Pharm. Bull.*, 2003, **51**, 999.
- 72 M. Taki, K. Niitu, Y. Omiya, M. Noguchi, M. Fukuchi, M. Aburada and M. Okada, *Planta Med.*, 2003, **69**, 800.
- 73 D. Csupor, P. Forgo, K. Csedő and J. Hohmann, *Helv. Chim. Acta*, 2006, **89**, 2981.
- 74 A. H. Meriçli, S. Pirildar, S. Süzgeç, L. Bitiş, F. Meriçli, H. Özçelik, J. Zapp and H. Becker, *Helv. Chim. Acta*, 2006, **89**, 210.
- 75 C. S. Peng, F. P. Wang and X. X. Jian, *J. Asian Nat. Prod. Res.*, 2000, **2**, 245.
- 76 Atta-ur-Rahman, N. Fatima, F. Akhtar, M. I. Choudhary and A. Khalid, *J. Nat. Prod.*, 2000, **63**, 1393.
- 77 J. Y. Dong and L. Li, *Acta Bot. Yunnanica*, 2001, **23**, 381.
- 78 L. H. Hou, D. L. Chen, X. X. Jian and F. P. Wang, *Chem. Pharm. Bull.*, 2007, **55**, 1090.
- 79 P. Tang, D. L. Chen, X. X. Jian and F. P. Wang, *Chin. Chem. Lett.*, 2007, **18**, 704.
- 80 A. Ulubelen, A. H. Meriçli, F. Meriçli, U. Kolak, M. Arfan, M. Ahmad and H. Ahmad, *Heterocycles*, 2000, **53**, 2279.
- 81 S. Yang, X. D. Yang, J. F. Zhao, H. B. Zhang and L. Li, *Helv. Chim. Acta*, 2007, **90**, 1160.
- 82 F. P. Wang, Z. B. Li, J. J. Chen and J. S. Yang, *Chin. Chem. Lett.*, 2000, **11**, 1003.
- 83 P. A. Q. H. Chen, D. L. Chen, X. X. Jian and F. P. Wang, *J. Asian Nat. Prod. Res.*, 2004, **6**, 151.
- 84 K. S. Khetwal and S. Pande, *Nat. Prod. Res.*, 2004, **18**, 129.
- 85 H. Ji and F. P. Wang, *J. Asian Nat. Prod. Res.*, 2006, **8**, 619.
- 86 X. Pan, L. He, B. G. Li and Y. Z. Chen, *Chin. Chem. Lett.*, 1998, **9**, 57.
- 87 A. H. Meriçli, F. Meriçli, H. K. Desai, R. Ilarslan, A. Ulubelen and S. W. Pelletier, *Pharmazie*, 2001, **56**, 418.
- 88 U. Kolak, A. Türkekul, F. Özgökçe and A. Ulubelen, *Pharmazie*, 2005, **60**, 953.
- 89 X. Y. Wei, H. H. Xie, M. F. Liu and X. J. Ge, *Heterocycles*, 2000, **53**, 2027.
- 90 Atta-ur-Rahman, Atia-tul-Wahab, M. N. Sultankhodzhaev, U. T. Teshebaeva and M. I. Choudhary, *Nat. Prod. Res.*, 2005, **19**, 713.
- 91 S. Y. Lee, S. H. Shim, J. S. Kim, J. H. Lee, H. Y. Lee, D. Y. Jung, H. Ha, C. Kim and S. S. Kang, *Arch. Pharmacol Res.*, 2007, **30**, 691.
- 92 H. L. Yu and S. S. Jia, *Acta Pharm. Sin.*, 2000, **35**, 232.
- 93 L. M. Gao, X. M. Wei and L. Yang, *Chin. Chem. Lett.*, 2005, **16**, 475.
- 94 Y. L. Ren, Z. H. Huang and S. S. Jia, *Acta Pharm. Sin.*, 1999, **34**, 873.
- 95 M. N. Sultankhodzhaev and Z. S. Boronova, *Khim. Prir. Soedin.*, 1999, **35**, 226.
- 96 F. Zhang, S. L. Peng, F. Luo and L. S. Ding, *Chin. Chem. Lett.*, 2005, **16**, 1043.
- 97 K. Ishimi, M. Makino, Y. Asada, Y. Ichinohe and Y. Fujimoto, *J. Nat. Med.*, 2006, **60**, 255.
- 98 Z. S. Boronova and M. N. Sultankhodzhaev, *Khim. Prir. Soedin.*, 2001, **37**, 228.
- 99 X. L. Zhou, Q. H. Chen, D. L. Chen and F. P. Wang, *Chem. Pharm. Bull.*, 2003, **51**, 592-4.
- 100 H. Ji, D. L. Chen and F. P. Wang, *Heterocycles*, 2004, **63**, 2363.
- 101 I. Kitagawa, M. Yoshikawa, Z. L. Chen and K. Kobayashi, *Chem. Pharm. Bull.*, 1982, **30**, 758.
- 102 S. H. Shim, S. Y. Lee, J. S. Kim, K. H. Son and S. S. Kang, *Arch. Pharmacol Res.*, 2005, **28**, 1239.
- 103 C. S. Peng, F. P. Wang and X. X. Jian, *Chin. Chem. Lett.*, 2002, **13**, 233.
- 104 X. D. Yang, S. Yang, J. Yang, J. F. Zhao, H. B. Zhang and L. Li, *Helv. Chim. Acta*, 2008, **91**, 569.
- 105 H. K. Desai, L. P. Silverman and S. W. Pelletier, *Heterocycles*, 1998, **48**, 1107.
- 106 S. Yang, X. D. Yang, J. F. Zhao, Y. Jin, H. B. Zhang and L. Li, *Khim. Prir. Soedin.*, 2008, **44**, 265.
- 107 A. H. Meriçli, F. Meriçli, A. Ulubelen, H. K. Desai, B. S. Joshi, S. W. Pelletier, S. Özden and M. Küçükislamoglu, *Heterocycles*, 1998, **47**, 329.
- 108 Y. Chen and A. Katz, *J. Nat. Prod.*, 1999, **62**, 798.
- 109 S. M. Zhang, G. L. Zhao and G. Q. Lin, *Phytochemistry*, 1999, **51**, 333.
- 110 X. L. Zhou, Q. H. Chen, D. L. Chen and F. P. Wang, *Chin. J. Chem.*, 2003, **21**, 871.
- 111 X. L. Zhou, Q. H. Chen and F. P. Wang, *Heterocycles*, 2004, **63**, 123.
- 112 N. Batbayar, S. Enkhzaya, J. Tunsag, D. Batsuren, D. S. Rycroft, S. Sproll and F. Bracher, *Phytochemistry*, 2003, **62**, 543.
- 113 F. Shaheen, M. Ahmad, M. T. H. Khan, S. Jallil, A. Ejaz, M. N. Sultankhodjaev, M. Arfan, M. I. Choudhary and Atta-ur-Rahman, *Phytochemistry*, 2005, **66**, 935.
- 114 A. Alva, M. Grandez, A. Madinaveitia, G. de la Fuente and J. Gavín, *Chem. Pharm. Bull.*, 2004, **52**, 530.
- 115 X. X. Liang, D. L. Chen and F. P. Wang, *Chin. Chem. Lett.*, 2006, **17**, 1473.
- 116 H. Y. Pu, Q. Y. Xu, F. P. Wang and C. T. Che, *Planta Med.*, 1996, **62**, 462.
- 117 J. Hohmann, P. Forgo, Z. Hajdú, E. Varga and I. Máthé, *J. Nat. Prod.*, 2002, **65**, 1069.
- 118 Zs. Hajdú, P. Forgo, B. Löffler and J. Hohmann, *Biochem. Syst. Ecol.*, 2005, **33**, 1081.
- 119 X. L. Shen and F. P. Wang, *Chin. J. Nat. Med.*, 2004, **2**, 152.
- 120 S. W. Pelletier, F. M. Harraz, M. M. Badawi, S. Tantiraksachai, F. P. Wang and S. Y. Chen, *Heterocycles*, 1986, **24**, 1853.
- 121 M. X. Yu, Y. J. Pan and Y. Z. Chen, *Chin. J. Org. Chem.*, 2003, **23**, 563.
- 122 Y. Q. He, Z. Y. Ma, Q. Yang, B. H. Yao and L. M. Gao, *Acta Pharm. Sin.*, 2008, **43**, 934.

- 123 B. T. Salimov, *Khim. Prir. Soedin.*, 2001, **37**, 231.
- 124 A. Ulubelen, H. K. Desai, Q. Teng, A. H. Meriçli, F. Meriçli, U. Kolak, M. Arfan, C. K. Lee and S. W. Pelletier, *Heterocycles*, 1999, **51**, 1897.
- 125 P. Tang, D. L. Chen, Q. H. Chen, X. X. Jian and F. P. Wang, *Chin. Chem. Lett.*, 2007, **18**, 700.
- 126 T. M. Gabbasov, E. M. Tsyrlina, L. V. Spirikhin, N. I. Fedorov and M. S. Yunusov, *Khim. Prir. Soedin.*, 2008, **44**, 380.
- 127 L. S. Ding, J. Wang, S. L. Peng and N. Y. Chen, *Acta Bot. Sin.*, 2000, **42**, 523.
- 128 L. P. Yan, D. L. Chen and F. P. Wang, *Chin. J. Org. Chem.*, 2007, **27**, 976.
- 129 S. M. Zhang and Q. Y. Ou, *Phytochemistry*, 1998, **48**, 191.
- 130 L. He, Y. J. Pan, B. G. Li and Y. Z. Chen, *Chin. Chem. Lett.*, 1999, **10**, 1027.
- 131 F. Shaheen, M. Zeeshan, M. Ahmad, S. Anjum, S. Ali, H. K. Fun, M. I. Choudhary and Atta-ur-Rahman, *J. Nat. Prod.*, 2006, **69**, 823.
- 132 Y. He, D. L. Chen and F. P. Wang, *Nat. Prod. Commun.*, 2006, **1**, 357.
- 133 X. L. Shen, X. L. Zhou, Q. H. Chen, D. L. Chen and F. P. Wang, *Chem. Pharm. Bull.*, 2002, **50**, 1265.
- 134 X. L. Zhou, Q. H. Chen and F. P. Wang, *Chem. Pharm. Bull.*, 2004, **52**, 381.
- 135 A. Ulubelen, A. H. Meriçli, F. Meriçli, U. Kolak, M. Arfan, M. Ahmad and H. Ahmad, *Pharmazie*, 2002, **57**, 427.
- 136 D. R. Gardner, G. D. Manners, K. E. Panter, S. T. Lee and J. A. Pfister, *J. Nat. Prod.*, 2000, **63**, 1127.
- 137 E. D. Khairitdinova, E. M. Tsyrlina, L. V. Spirikhin, N. I. Fedorov, Yu. Ya. Efremov and M. S. Yunusov, *Russ. Chem. Bull.*, 2003, **52**, 2078.
- 138 T. M. Gabbasov, E. M. Tsyrlina, L. V. Spirikhin, V. T. Danilov and M. S. Yunusov, *Russ. J. Bioorg. Chem.*, 2005, **31**, 383.
- 139 E. D. Khairitdinova, E. M. Tsyrlina, L. V. Spirikhin, N. I. Fedorov and M. S. Yunusov, *Khim. Prir. Soedin.*, 2005, **41**, 467.
- 140 D. Csupor, P. Forgo, K. Csédő, I. Máthé and J. Hohmann, *Acta Pharm. Hung.*, 2006, **76**, 181.
- 141 D. Y. Gu, H. A. Aisa and S. K. Usmanova, *Khim. Prir. Soedin.*, 2007, **43**, 248.
- 142 D. L. Chen, L. Y. Lin, Q. H. Chen, X. X. Jian and F. P. Wang, *J. Asian Nat. Prod. Res.*, 2003, **5**, 209.
- 143 E. D. Khairitdinova, E. M. Tsyrlina, L. V. Spirikhin, N. I. Fedorov and M. S. Yunusov, *Khim. Prir. Soedin.*, 2005, **41**, 469.
- 144 L. He, X. Pan, B. G. Li and Y. Z. Chen, *Indian J. Chem., Sect. B: Org. Chem. Incl. Med. Chem.*, 1998, **37**, 612.
- 145 D. Csupor, P. Forgo, I. Máthé and J. Hohmann, *Helv. Chim. Acta*, 2004, **87**, 2125.
- 146 J. G. Díaz, J. G. Ruiz and W. Herz, *Phytochemistry*, 2004, **65**, 2123.
- 147 L. Bitiş, S. Süzgeç, U. Sözer, H. Özçelik, J. Zapp, A. K. Kiemer, F. Meriçli and A. H. Meriçli, *Helv. Chim. Acta*, 2007, **90**, 2217.
- 148 A. Katz and H. Rudin, *Helv. Chim. Acta*, 1984, **67**, 2017.
- 149 T. Mori, T. Ohsawa, M. Murayama, H. Bando, K. Wada and T. Amiya, *Heterocycles*, 1989, **29**, 873.
- 150 D. H. Chen and W. L. Sung, *Acta Chim. Sin.*, 1983, **41**, 843.
- 151 F. P. Wang, Z. B. Li, X. P. Dai and C. S. Peng, *Phytochemistry*, 1997, **45**, 1539.
- 152 P. A. D. L. Chen, Q. H. Chen, X. X. Jian and F. P. Wang, *Nat. Prod. Res. Dev.*, 2002, **14**(5), 6.
- 153 F. P. Wang, Z. B. Li and C. T. Che, *J. Nat. Prod.*, 1998, **61**, 1555.
- 154 E. G. Zinurova, T. V. Khakimova, L. V. Spirikhin, M. S. Yunusov, P. G. Gorovoi and G. A. Tolstikov, *Russ. Chem. Bull.*, 2001, **50**, 311.
- 155 M. N. Sultankhodzaev, B. Tashkhodzhaev, B. B. Averkiev and M. Yu. Antipin, *Khim. Prir. Soedin.*, 2002, **38**, 63.
- 156 M. N. Sultankhodzaev, Atia-tul-Wahab, M. I. Choudhary and Atta-ur-Rahman, *Khim. Prir. Soedin.*, 2003, **39**, 421.
- 157 Z. Y. Li, J. F. Zhao, J. H. Yang, H. B. Zhang and L. Li, *Helv. Chim. Acta*, 2004, **87**, 2085.
- 158 S. K. Usmanova, I. A. Bessonova, N. D. Abdullaev and M. G. Levkovich, *Khim. Prir. Soedin.*, 1999, **35**, 113.
- 159 M. S. Yunusov, E. M. Tsyrlina, E. D. Khairitdinova, L. V. Spirikhin, A. Yu. Kovalevsky and M. Yu. Antipin, *Russ. Chem. Bull.*, 2000, **49**, 1629.
- 160 O. E. Edwards, *Can. J. Chem.*, 1981, **59**, 3039.
- 161 A. S. Narzullaev and M. S. Yunusov, *Khim. Prir. Soedin.*, 1991, **27**, 545.
- 162 E. D. Khairitdinova, E. M. Tsyrlina, L. V. Spirikhin, A. A. Balandina, Sh. K. Latypov and M. S. Yunusov, *Russ. J. Org. Chem.*, 2008, **44**, 536.
- 163 J. Xiong, N. H. Tan, C. J. Ji, Y. Lu and N. B. Gong, *Tetrahedron Lett.*, 2008, **49**, 4851.
- 164 Z. B. Li and F. P. Wang, *J. Asian Nat. Prod. Res.*, 1998, **1**, 87.
- 165 H. P. He, Y. M. Shen, X. S. Yang, B. G. Wang, L. Li and X. J. Hao, *Chin. Chem. Lett.*, 2001, **12**, 67.
- 166 H. P. He, Y. M. Shen, J. X. Zhang, G. Y. Zuo and X. J. Hao, *J. Nat. Prod.*, 2001, **64**, 379.
- 167 F. Meriçli, A. H. Meriçli, A. Ulubelen, H. K. Desai and S. W. Pelletier, *J. Nat. Prod.*, 2001, **64**, 787.
- 168 A. Ulubelen, M. Arfan, U. Sönmez, A. H. Meriçli and F. Meriçli, *Phytochemistry*, 1998, **47**, 1141.
- 169 G. Y. Zuo, H. P. He, X. Hong, W. M. Zhu, Y. M. Hu, X. S. Yang and X. J. Hao, *Chin. Chem. Lett.*, 2001, **12**, 147.
- 170 G. Y. Zuo, H. P. He, X. Hong, W. M. Zhu, X. S. Yang and X. J. Hao, *Heterocycles*, 2001, **55**, 487.
- 171 Y. M. Shen, H. P. He, Y. S. Zhang, B. G. Wang and X. J. Hao, *Chin. Chem. Lett.*, 2000, **11**, 789.
- 172 Y. Q. He, X. M. Wei, Y. L. Han and L. M. Gao, *Chin. Chem. Lett.*, 2007, **18**, 545.
- 173 T. S. Wu, C. C. Hwang, P. C. Kuo, A. G. Damu, C. J. Chou and C. F. Chen, *Heterocycles*, 2002, **57**, 1495.
- 174 B. G. Wang, L. Li, X. S. Yang, Z. H. Chen and X. J. Hao, *Heterocycles*, 2000, **53**, 1343.
- 175 B. G. Wang, X. Hong, G. Y. Zuo and X. J. Hao, *J. Asian Nat. Prod. Res.*, 2000, **2**, 271.
- 176 B. G. Wang, B. Liu, G. Y. Zuo and X. J. Hao, *Acta Bot. Yunnanica*, 2000, **22**, 209.
- 177 J. G. Díaz, J. G. Ruiz and G. de la Fuente, *J. Nat. Prod.*, 2000, **63**, 1136.
- 178 P. M. Shrestha and A. Katz, *J. Nat. Prod.*, 2004, **67**, 1574.
- 179 F. Zhang, S. L. Peng, X. Liao, K. B. Yu and L. S. Ding, *Planta Med.*, 2005, **71**, 1073.
- 180 A. H. Meriçli, S. Süzgeç, L. Bitiş, F. Meriçli, H. Özçelik, J. Zapp and H. Becker, *Pharmazie*, 2006, **61**, 483.
- 181 A. H. Meriçli, F. Meriçli, G. V. Seyhan, M. Bahar, H. K. Desai, H. Özçelik and A. Ulubelen, *Pharmazie*, 2002, **57**, 761.
- 182 Z. C. Miao, R. Feng, Y. X. Zhou, G. Y. Li, Z. G. Hao and J. H. Liu, *Chin. J. Magn. Reson.*, 1999, **16**, 547.
- 183 F. Feng, J. H. Liu and S. X. Zhao, *Phytochemistry*, 1998, **49**, 2557.
- 184 A. H. Meriçli, F. Meriçli, E. Doğru, H. Özçelik, Atta-ur-Rahman and A. Ulubelen, *Phytochemistry*, 1999, **51**, 337.
- 185 C. S. Peng, X. X. Jian, F. P. Wang and D. L. Chen, *Chin. Chem. Lett.*, 2000, **11**, 411.
- 186 M. Li, X. B. Du, Y. M. Shen, B. G. Wang and X. J. Hao, *Chin. Chem. Lett.*, 1999, **10**, 827.
- 187 A. H. Meriçli, F. Meriçli, H. K. Desai, B. S. Joshi, Q. Teng, K. Bhattacharyya, G. Melikoglu, M. Küçükislamoglu, A. Ulubelen and S. W. Pelletier, *Heterocycles*, 2000, **53**, 1987.
- 188 H. Q. Wang, S. H. Jiang, P. M. Yang, M. H. Ying, S. J. Lin and D. Y. Zhu, *Nat. Prod. Res. Dev.*, 2002, **14**(4), 13.
- 189 J. X. Cao, L. B. Li, J. Ren, S. P. Jiang, R. R. Tian, X. L. Chen, S. L. Peng, J. Zhang and H. J. Zhu, *Helv. Chim. Acta*, 2008, **91**, 1954.
- 190 L. M. Gao, X. M. Wei and L. Yang, *J. Chem. Res. (S)*, 2004, 307.
- 191 Z. C. Miao, R. Feng, Y. X. Zhou, G. Y. Li, J. H. Liu and Z. G. Hao, *Chin. J. Magn. Reson.*, 1999, **16**, 403.
- 192 C. H. Yang, H. J. Zhang and J. H. Liu, *Chin. Tradit. Herbal Drugs*, 2004, **35**, 1328.
- 193 K. Jiang, C. H. Yang, J. H. Liu and Q. F. Tang, *Acta Pharm. Sin.*, 2006, **41**, 128.
- 194 Q. F. Tang, C. H. Yang, J. H. Liu, W. C. Ye, S. X. Zhao, Y. Lu, L. Mao and Q. T. Zheng, *Acta Pharm. Sin.*, 2005, **40**, 640.
- 195 Q. F. Tang, J. H. Liu, J. Xue, W. C. Ye, Z. J. Zhang and C. H. Yang, *J. Chromatogr., B*, 2008, **872**, 181.
- 196 C. H. Yang, X. C. Wang, Q. F. Tang, W. Y. Liu and J. H. Liu, *Helv. Chim. Acta*, 2008, **91**, 759.
- 197 S. H. Shim, J. S. Kim, S. S. Kang, K. H. Son and K. Bae, *J. Asian Nat. Prod. Res.*, 2006, **8**, 451.
- 198 P. Wangchuk, J. B. Bremner and S. Samosorn, *J. Nat. Prod.*, 2007, **70**, 1808.
- 199 S. K. Srivastava, H. K. Desai, V. Vobalaboina and S. W. Pelletier, *J. Liq. Chromatogr. Relat. Technol.*, 1999, **22**, 1687.

- 200 L. Ruiz-Mesía, A. Madinaveitia, M. Reina, M. L. Rodriguez, G. de la Fuente and W. Ruiz-Mesía, *J. Nat. Prod.*, 2002, **65**, 496.
- 201 L. M. Fan, H. P. He, Y. M. Shen and X. J. Hao, *J. Integr. Plant Biol.*, 2005, **47**, 120.
- 202 N. Singh and A. Singh, *J. Indian Chem. Soc.*, 1965, **42**, 49.
- 203 N. Singh and S. S. Jaswal, *Tetrahedron Lett.*, 1968, **9**, 2219.
- 204 S. W. Pelletier, K. N. Iyer, L. H. Wright, M. G. Newton and N. Singh, *J. Am. Chem. Soc.*, 1971, **93**, 5942.
- 205 L. Li, J. F. Zhao, Y. B. Wang and H. B. Zhang, *Helv. Chim. Acta*, 2004, **87**, 866.
- 206 Y. B. Wang, R. Huang, H. B. Zhang and L. Li, *Helv. Chim. Acta*, 2005, **88**, 1081.
- 207 X. L. Zhou, D. L. Chen, Q. H. Chen and F. P. Wang, *J. Nat. Prod.*, 2005, **68**, 1076.
- 208 Q. P. Jiang and S. W. Pelletier, *J. Nat. Prod.*, 1991, **54**, 525.
- 209 B. T. Salimov, K. K. Turgunov, B. Tashkhodzhaev and F. N. Dzhakhangirov, *Khim. Prir. Soedin.*, 2004, **40**, 129.
- 210 L. M. Gao, X. M. Wei, X. L. Jin and L. Yang, *Heterocycles*, 2004, **63**, 1181.
- 211 C. S. Peng, X. X. Jian and F. P. Wang, *J. Asian Nat. Prod. Res.*, 2001, **3**, 49.
- 212 S. W. Pelletier, N. V. Mody, J. Finer-Moore, A. M. Aceyai and L. C. Schramm, *J. Chem. Soc., Chem. Commun.*, 1981, 327.
- 213 S. W. Pelletier, J. A. Glinski, K. I. Varughese, J. Maddry and N. V. Mody, *Heterocycles*, 1983, **20**, 413.
- 214 Z. B. Li, G. H. Lü, D. L. Chen and F. P. Wang, *Nat. Prod. Res. Dev.*, 1997, **9**(1), 9.
- 215 Z. B. Li and F. P. Wang, *J. Asian Nat. Prod. Res.*, 1998, **1**, 87.
- 216 X. Z. Zhen and F. P. Wang, *Nat. Prod. Res. Dev.*, 2002, **14**(1), 13.
- 217 D. L. Chen, Z. B. Li, C. S. Peng, X. Z. Zheng, Q. H. Chen and F. P. Wang, *Chin. J. Org. Chem.*, 2003, **23**, 674.
- 218 C. H. Yang, J. H. Liu, B. R. Xiang, Y. Lu, C. Wang and Q. T. Zheng, *Chin. Tradit. Herbal Drugs*, 2002, **33**, 201.
- 219 F. P. Wang, C. S. Peng and K. B. Yu, *Tetrahedron*, 2000, **56**, 7443.
- 220 B. Tashkhodzhaev, Sh. A. Saidkhodzhaeva, I. A. Bessonova and M. Yu. Antipin, *Khim. Prir. Soedin.*, 2000, **36**, 62.
- 221 Sh. A. Saidkhodzhaeva, I. A. Bessonova and N. D. Abdullaev, *Khim. Prir. Soedin.*, 2001, **37**, 397.
- 222 K. Nishimura, Y. Hitotsuyanagi, N. Sugeta, H. Fukaya, Y. Aoyagi, T. Hasuda, T. Kinoshita and K. Takeya, *J. Nat. Prod.*, 2007, **70**, 758.
- 223 A. Arnone, A. Bava, G. Fronza, G. Nasini and E. Ragg, *Tetrahedron Lett.*, 2005, **46**, 8037.
- 224 Y. M. Zhang, N. H. Tan, Y. Lu, Y. Chang and R. R. Jia, *Org. Lett.*, 2007, **9**, 4579.
- 225 M. J. Uddin, S. Kokubo, K. Suenaga, K. Ueda and D. Uemura, *Heterocycles*, 2001, **54**, 1039.
- 226 M. J. Uddin, S. Kokubo, K. Ueda, K. Suenaga and D. Uemura, *J. Nat. Prod.*, 2001, **64**, 1169.
- 227 M. J. Uddin, S. Kokubo, K. Ueda, K. Suenaga and D. Uemura, *Chem. Lett.*, 2002, 1028.
- 228 C. S. Li, Y. T. Di, S. Z. Mu, H. P. He, Q. Zhang, X. Fang, Y. Zhang, S. L. Li, Y. Lu, Y. Q. Gong and X. J. Hao, *J. Nat. Prod.*, 2008, **71**, 1202.
- 229 J. Z. Fan, Z. B. Li, Q. H. Chen, F. P. Wang and B. G. Li, *Chin. Chem. Lett.*, 2000, **11**, 417.
- 230 C. L. Zou, H. Ji, G. B. Xie, D. L. Chen and F. P. Wang, *J. Asian Nat. Prod. Res.*, 2008, **10**, 1063.
- 231 F. P. Wang, J. Z. Fan, Z. B. Li, J. S. Yang and B. G. Li, *Chin. Chem. Lett.*, 1999, **10**, 375.
- 232 F. P. Wang, Z. B. Li, J. S. Yang and B. G. Li, *Chin. Chem. Lett.*, 1999, **10**, 453.
- 233 Z. B. Li, Q. H. Chen, F. P. Wang and B. G. Li, *Chin. Chem. Lett.*, 2000, **11**, 421.
- 234 E. G. Zinurova, N. N. Kabal'nov, V. V. Shereshevets, E. V. Ivanova, E. E. Shults, G. A. Tolstikov and M. S. Yunusov, *Russ. Chem. Bull.*, 2001, **50**, 720.
- 235 S. A. Osadchii, N. A. Pankrushina, M. M. Shakirov, E. E. Shults and G. A. Tolstikov, *Russ. Chem. Lett.*, 2000, **49**, 557.
- 236 Y. M. He, C. L. Zou, Q. H. Chen and F. P. Wang, *J. Asian Nat. Prod. Res.*, 2007, **9**, 713.
- 237 X. L. Shen and F. P. Wang, *Chem. Pharm. Bull.*, 2005, **53**, 267.
- 238 X. L. Shen and F. P. Wang, *Chem. Pharm. Bull.*, 2004, **52**, 1095.
- 239 S. A. Osadchii, E. E. Shults and G. A. Tolstikov, *Russ. Chem. Bull.*, 2001, **50**, 907.
- 240 Q. H. Chen, F. P. Wang and K. B. Yu, *Chin. Chem. Lett.*, 2000, **11**, 689.
- 241 Q. H. Chen and F. P. Wang, *Chin. Chem. Lett.*, 2001, **12**, 421.
- 242 F. P. Wang, Q. H. Chen, Z. B. Li and B. G. Li, *Chem. Pharm. Bull.*, 2001, **49**, 689.
- 243 Y. Wang, L. Shi, F. R. Song, Z. Q. Liu and S. Y. Liu, *Rapid Commun. Mass Spectrom.*, 2003, **17**, 279.
- 244 Y. F. Zhao, F. R. Song, Y. H. Guo and S. Y. Liu, *Chem. J. Chin. Univ.*, 2008, **29**, 55.
- 245 H. Yue, Z. P. Pi, H. L. Li, F. R. Song, Z. Q. Liu and S. Y. Liu, *Phytochem. Anal.*, 2008, **19**, 141.
- 246 Q. H. Chen, L. Xu and F. P. Wang, *Heterocycles*, 2002, **57**, 2357.
- 247 F. W. Bachelor, R. F. Brown and G. Büchi, *Tetrahedron Lett.*, 1960, **1**(31), 1.
- 248 Q. H. Chen, L. Xu and F. P. Wang, *Chin. Chem. Lett.*, 2003, **14**, 147.
- 249 Q. H. Chen and F. P. Wang, *Chem. Pharm. Bull.*, 2002, **50**, 1310.
- 250 F. P. Wang, J. Z. Fan, X. X. Jian and B. G. Li, *Chin. Chem. Lett.*, 1999, **10**, 379.
- 251 F. P. Wang, J. S. Yang, Q. H. Chen and L. Xu, *Chem. Pharm. Bull.*, 2000, **48**, 1912.
- 252 Q. H. Chen and F. P. Wang, *J. Asian Nat. Prod. Res.*, 2003, **5**, 43.
- 253 L. Xu, Q. H. Chen and F. P. Wang, *Tetrahedron*, 2002, **58**, 4267.
- 254 F. P. Wang, Q. H. Chen and B. G. Li, *Tetrahedron*, 2001, **57**, 4705.
- 255 Q. H. Chen, L. Xu and F. P. Wang, *Tetrahedron*, 2002, **58**, 9431.
- 256 F. P. Wang and L. Xu, *Tetrahedron*, 2005, **61**, 2149.
- 257 E. Ochiai, T. Okamoto, S. Sakai and A. Saito, *Yakugaku Zasshi*, 1956, **76**, 1414.
- 258 H. Muratake and M. Natsume, *Tetrahedron*, 2006, **62**, 7056.
- 259 H. Muratake and M. Natsume, *Tetrahedron*, 2006, **62**, 7071.
- 260 H. Muratake and M. Natsume, *Tetrahedron*, 2006, **62**, 7093.
- 261 H. Muratake and M. Natsume, *Tetrahedron Lett.*, 2002, **43**, 2913.
- 262 K. M. Peese and D. Y. Gin, *Chem. Eur. J.*, 2008, **14**, 1654.
- 263 M. K. Brown, T. L. May, C. A. Baxter and A. H. Hoveyda, *Angew. Chem., Int. Ed.*, 2007, **46**, 1097.
- 264 Y. S. Kwak and J. D. Winkler, *J. Am. Chem. Soc.*, 2001, **123**, 7429.
- 265 C. M. Williams and L. N. Mander, *Org. Lett.*, 2003, **5**, 3499.
- 266 C. M. Williams, L. N. Mander, P. V. Bernhardt and A. C. Willis, *Tetrahedron*, 2005, **61**, 3759.
- 267 O. E. Hutt and L. N. Mander, *J. Org. Chem.*, 2007, **72**, 10130.
- 268 D. F. Taber, J. L. Liang, B. Chen and L. S. Cai, *J. Org. Chem.*, 2005, **70**, 8739.
- 269 R. M. Conrad and J. Du Bois, *Org. Lett.*, 2007, **9**, 5465.
- 270 P. A. Coates, I. S. Blagbrough, M. G. Rowan, B. V. L. Potter, D. P. J. Pearson and T. Lewis, *Tetrahedron Lett.*, 1994, **35**, 8709.
- 271 H. Ohat, Y. Seto and N. Tsunoda, *J. Chromatogr., B: Biomed. Sci. Appl.*, 1997, **691**, 351.
- 272 W. X. Sun, S. Y. Liu, Z. Q. Liu, F. R. Song and S. P. Fang, *Rapid Commun. Mass Spectrom.*, 1998, **12**, 821.
- 273 D. R. Gardner, K. E. Panter, J. A. Pfister and A. P. Knight, *J. Agric. Food Chem.*, 1999, **47**, 5049.
- 274 W. X. Sun, F. R. Song, M. Cui and S. Y. Liu, *Planta Med.*, 1999, **65**, 432.
- 275 Y. Wang, Z. Q. Liu, F. R. Song and S. Y. Liu, *Rapid Commun. Mass Spectrom.*, 2002, **16**, 2075.
- 276 A. M. Sun, H. Li, Z. M. Huang, P. P. H. But and X. Q. Ding, *Chin. Chem. Lett.*, 2004, **15**, 1071.
- 277 H. Yue, P. Z. Pi, Y. F. Zhao, F. R. Song, Z. Q. Liu and S. Y. Liu, *Chem. Res. Chin. Univ.*, 2007, **23**, 625.
- 278 Y. Wang, F. R. Song, Q. X. Xu, Z. Q. Liu and S. Y. Liu, *J. Mass Spectrom.*, 2003, **38**, 962.
- 279 Y. Wang, Z. Q. Liu, F. R. Song and S. Y. Liu, *Acta Pharm. Sin.*, 2003, **38**, 290.
- 280 Y. Tu, G. J. Zhang, S. M. Wang and Z. Q. Liu, *J. Chin. Mater. Med.*, 2008, **33**, 789.
- 281 Y. Wang, S. Y. Liu, Z. Q. Liu, F. R. Song, D. M. Jin and C. M. Liu, *Chin. J. Anal. Chem.*, 2003, **31**, 139.
- 282 S. K. Wong, S. K. Tsui and S. Y. Kwan, *J. Pharm. Biomed. Anal.*, 2002, **30**, 161.
- 283 H. Yue, Z. F. Pi, F. R. Song, Z. Q. Liu and S. Y. Liu, *Acta Chim. Sin.*, 2008, **66**, 211.
- 284 Y. Wang, F. R. Song, D. M. Jin, Z. Q. Liu and S. Y. Liu, *Chem. J. Chin. Univ.*, 2004, **25**, 85.



- 285 H. G. Zhang, Y. Sun, M. Y. Duan, Y. J. Chen, D. F. Zhong and H. Q. Zhang, *Toxicol.*, 2005, **46**, 500.
- 286 J. Beike, L. Frommherz, M. Wood, B. Brinkmann and H. Köhler, *Int. J. Legal Med.*, 2004, **118**, 289.
- 287 M. Hayashida, H. Hayakawa, K. Wada, T. Yamada, M. Nihira and Y. Ohno, *Legal Med.*, 2003, **5**, S101.
- 288 Z. H. Wang, Z. P. Wang, J. Wen and Y. He, *J. Pharm. Biomed. Anal.*, 2007, **45**, 145.
- 289 F. Zhang, M. H. Tang, L. J. Chen, R. Li, X. H. Wang, J. G. Duan, X. Zhao and Y. Q. Wei, *J. Chromatogr. B*, 2008, **873**, 173.
- 290 L. H. Chen, L. J. Jin, Z. M. Sun, Y. Q. Qiu, Y. Wang and S. Y. Liu, *Chem. J. Chin. Univ.*, 2005, **26**, 2340.
- 291 Y. Chen, S. Koelliker, M. Oehme and A. Katz, *J. Nat. Prod.*, 1999, **62**, 701.
- 292 K. Wada, T. Mori and N. Kawahara, *Chem. Pharm. Bull.*, 2000, **48**, 660.
- 293 K. Wada, T. Mori and N. Kawahara, *J. Mass Spectrom.*, 2000, **35**, 432.
- 294 K. Wada, T. Mori and N. Kawahara, *Chem. Pharm. Bull.*, 2000, **48**, 1065.
- 295 K. Ito, S. Tanaka, M. Funayama and M. Mizugaki, *J. Anal. Toxicol.*, 2000, **24**, 348.
- 296 A. M. Sun, D. H. Chen and P. X. Bi, *Chin. J. Chromatogr.*, 1999, **17**, 67.
- 297 S. P. Elliott, *Sci. Justice*, 2002, **42**, 111.
- 298 Z. H. Wang, D. A. Guo, Y. He, C. H. Ju and J. Z. Zhang, *Phytochem. Anal.*, 2004, **15**, 16.
- 299 F. X. Xiao, L. L. Zhou and R. Li, *Food and Drug*, 2005, **7**, 47.
- 300 F. Liu, X. H. Yu, F. Li, Y. Y. Tan and Y. J. Qiao, *China J. Chin. Mat. Med.*, 2006, **31**, 1160.
- 301 Z. H. Wang, J. Wen, J. B. Xing and Y. He, *J. Pharm. Biomed. Anal.*, 2006, **40**, 1031.
- 302 F. Q. Yang and Y. Ito, *J. Chromatogr., A*, 2001, **923**, 281.
- 303 K. Jiang, C. H. Yang, J. H. Liu and Q. F. Tang, *Acta Pharm. Sin.*, 2006, **41**, 128.
- 304 Q. F. Tang, C. H. Yang, W. C. Ye, J. H. Liu and S. X. Zhao, *J. Chromatogr., A*, 2007, **1144**, 203.
- 305 M. Liu, H. Zhang, L. Zhao, B. Y. Zhao, L. L. Dong, Z. Y. Zhu and Y. F. Chai, *Chromatographia*, 2008, **67**, 1003.
- 306 Yu. V. Nesterova, T. N. Poveteva, Yu. G. Nagorniyak, T. I. Andreeva and N. I. Suslov, *Bull. Exp. Biol. Med.*, 2008, **145**, 724.
- 307 J. L. Wang, X. L. Shen, Q. H. Chen, G. Qi, W. Wang and F. P. Wang, *Chem. Pharm. Bull.*, 2009, **57**, 801.
- 308 U. T. Gutser, J. Friese, J. F. Heubach, T. Matthiesen, N. Selve, B. Willfert and J. Gleitz, *Naunyn-Schmiedeberg's Arch. Pharmacol.*, 1998, **357**, 39.
- 309 A. M. Bello-Ramírez, J. Buendía-Orozco and A. A. Nava-Ocampo, *Fundam. Clin. Pharmacol.*, 2003, **17**, 575.
- 310 S. Zafar, M. A. Ahmad and T. A. Siddiqui, *J. Ethnopharmacol.*, 2001, **78**, 95.
- 311 S. Rahman, R. A. Khan and A. Kumar, *BMC Complementary Altern. Med.*, 2002, **2**, 6.
- 312 S. Zafar, M. A. Ahmad and T. A. Siddiqui, *Fitoterapia*, 2002, **73**, 553.
- 313 H. K. Desai, B. P. Hart, R. W. Caldwell, J. Z. Huang and S. W. Pelletier, *J. Nat. Prod.*, 1998, **61**, 743.
- 314 J. F. Heubach and A. Schüle, *Planta Med.*, 1998, **64**, 22.
- 315 N. Kh. Shakhidoyatova, F. N. Dzhakhangirov and M. N. Sultankhodzaev, *Pharm. Chem. J.*, 2001, **35**, 266.
- 316 D. K. Kim, H. Y. Kwon, K. R. Lee, D. K. Rhee and O. P. Zee, *Arch. Pharmacol. Res.*, 1998, **21**, 344.
- 317 C. de Ines, M. Reina, J. A. Gavin and A. Gonzalez-Coloma, *Z. Naturforsch., C: J. Biosci.*, 2006, **61**, 11.
- 318 K. Wada, M. Hazawa, K. Takahashi, T. Mori, N. Kawahara and I. Kashiwakura, *J. Nat. Prod.*, 2007, **70**, 1854.
- 319 M. Reina and A. González-Coloma, *Phytochem. Rev.*, 2007, **6**, 81.
- 320 M. Hazawa, K. Wada, K. Takahashi, T. Mori, N. Kawahara and I. Kashiwakura, *Invest. New Drugs*, 2009, **27**, 111.
- 321 A. Ameri, *Eur. J. Pharmacol.*, 1998, **342**, 183.
- 322 A. Ameri and T. Simmet, *Eur. J. Pharmacol.*, 1999, **386**, 187.
- 323 M. Raza, F. Shaheen, M. I. Choudhary, A. Suria, A. U. Rahman, S. Sombati and R. J. DeLorenzo, *Phytother. Res.*, 2001, **15**, 426.
- 324 M. Raza, F. Shaheen, M. I. Choudhary, A. U. Rahman, S. Sombati, A. Suria, A. Rafiq and R. J. DeLorenzo, *Phytother. Res.*, 2003, **17**, 38.
- 325 A. González-Coloma, A. Guadaño, C. Gutiérrez, R. Cabrera, E. de La Peña, G. deLa Fuente and M. Reina, *J. Agric. Food Chem.*, 1998, **46**, 286.
- 326 A. Ulubelen, A. H. Meriçli, F. Meriçli, N. Kilinçer, A. G. Ferizli, M. Emekci and S. W. Pelletier, *Phytother. Res.*, 2001, **15**, 170.
- 327 A. González-Coloma, M. Reina, A. Guadaño, R. Martínez-Díaz, J. G. Díaz, J. García-Rodríguez, A. Alva and M. Grandez, *Chem. Biodiversity*, 2004, **1**, 1327.
- 328 A. González-Coloma, M. Reina, A. Madinaveitia, A. Guadaño, O. Santana, R. Martínez-Díaz, L. Ruiz-Mesia, A. Alva, M. Grandez, R. Diaz, J. A. Gavin and G. de la Fuente, *J. Chem. Ecol.*, 2004, **30**, 1393.
- 329 P. González, C. Marín, I. Rodríguez-González, A. B. Hitos, M. J. Rosales, M. Reina, J. G. Díaz, A. González-Coloma and M. Sánchez-Moreno, *Int. J. Antimicrob. Agents*, 2005, **25**, 136.
- 330 P. González, C. Marín, I. Rodríguez-González, A. Illana, H. Mateo, S. S. Longoni, M. J. Rosales and M. Sánchez-Moreno, *Pharmacology*, 2006, **76**, 123.
- 331 K. Wada, S. Ishizuki, T. Mori, E. Fujihira and N. Kawahara, *Biol. Pharm. Bull.*, 1998, **21**, 140.
- 332 K. Wada, S. Ishizuki, T. Mori, E. Fujihira and N. Kawahara, *Biol. Pharm. Bull.*, 2000, **23**, 607.
- 333 F. N. Ngounou, R. N. Manfouo, L. A. Taponjdjou, D. Lontsi, V. Kuate, V. Penlap, F. X. Etoa, M.-A. L. Dubois and B. L. Sondengam, *Bull. Chem. Soc. Ethiop.*, 2005, **19**, 221.
- 334 B. Şener, I. Orhan and B. Özçelik, *ARKIVOC*, 2007, 265.
- 335 C. F. Wang, P. Gerner, S. Y. Wang and G. K. Wang, *Anesthesiology*, 2007, **107**, 82.
- 336 F. N. Dzhakhangirov, K. R. Kasymova, M. N. Sultankhodzaev, B. T. Salimov, S. K. Usmanova and R. Sh. Shakirov, *Khim. Prir. Soedin.*, 2007, **43**, 477.
- 337 N. Gaibazzi, G. Montresor, D. Canel, T. Comini, M. Fracalossi, M. L. Poeta and V. Ziacchi, *Ital. Heart J.*, 2002, **3**, 874.
- 338 F. Moritz, P. Compagnon, I. G. Kaliszczak, Y. Kaliszczak, V. Caliskan and C. Girault, *Clin. Toxicol.*, 2005, **43**, 873.
- 339 F. N. Dzhakhangirov and I. A. Bessonova, *Khim. Prir. Soedin.*, 2002, **38**, 60.
- 340 J. A. Pfister, D. R. Gardner, K. E. Panter, G. D. Manners, M. H. Ralphs, B. L. Stegelmeier and T. K. Schoch, *J. Nat. Toxins*, 1999, **8**, 81.
- 341 M. H. Ralphs, D. R. Gardner, D. L. Turner, J. A. Pfister and E. Thacker, *J. Chem. Ecol.*, 2002, **28**, 2327.
- 342 B. L. Stegelmeier, J. O. Hall, D. R. Gardner and K. E. Panter, *J. Anim. Sci.*, 2003, **81**, 1237.
- 343 G. D. Manners, K. E. Panter, J. A. Pfister, M. H. Ralphs and L. F. James, *J. Nat. Prod.*, 1998, **61**, 1086.
- 344 D. R. Gardner and J. A. Pfister, *J. Range Manage.*, 2000, **53**, 329.
- 345 D. R. Gardner and J. A. Pfister, *Rangeland Ecol. Manage.*, 2007, **60**, 441.
- 346 M. H. Ralphs, D. R. Gardner and J. A. Pfister, *J. Chem. Ecol.*, 2000, **26**, 1595.
- 347 S. T. Lee, D. R. Gardner and B. L. Stegelmeier, *J. Agric. Food Chem.*, 2000, **48**, 4520.
- 348 S. T. Lee, B. L. Stegelmeier, K. E. Panter, J. A. Pfister, D. R. Gardner, T. K. Schoch and L. F. James, *J. Anim. Sci.*, 2003, **81**, 232.
- 349 A. M. Bello-Ramírez and A. A. Nava-Ocampo, *Fundam. Clin. Pharmacol.*, 2004, **18**, 699.
- 350 M. A. Turabekova and R. B. Rasulev, *Molecules*, 2004, **9**, 1194.
- 351 M. A. Turabekova and R. B. Rasulev, *Khim. Prir. Soedin.*, 5, 41, p. 170.
- 352 M. A. Turabekova, B. F. Rasulev, F. N. Dzhakhangirov and S. I. Salikhov, *Environ. Toxicol. Pharmacol.*, 2008, **25**, 310.
- 353 A. M. Bello-Ramírez and A. A. Nava-Ocampo, *Fundam. Clin. Pharmacol.*, 2004, **18**, 157.
- 354 A. Ameri, *Naunyn-Schmiedeberg's Arch. Pharmacol.*, 1998, **357**, 585.
- 355 U. Seitz and A. Ameri, *Biochem. Pharmacol.*, 1998, **55**, 883.
- 356 A. Ameri and T. Simmet, *Brain Res.*, 1999, **842**, 332.
- 357 H. Liu, Y. Li, M. K. Song, X. J. Tan, F. Cheng, S. X. Zheng, J. H. Shen, X. M. Luo, R. Y. Ji, J. M. Yue, G. Y. Hu, H. L. Jiang and K. X. Chen, *Chem. Biol.*, 2003, **10**, 1103.

- 
- 358 M. A. Turabekova, B. F. Rasulev, M. G. Levkovich, N. D. Abdullaev and J. Leszczynski, *Comput. Biol. Chem.*, 2008, **32**, 88.
- 359 P. Dobelis, J. E. Madl, J. A. Pfister, G. D. Manners and J. P. Walrond, *J. Pharmacol. Exp. Ther.*, 1999, **291**, 538.
- 360 A. R. L. Davies, D. J. Hardick, I. S. Blagbrough, B. V. L. Potter, A. J. Wolstenholme and S. Wonnacott, *Neuropharmacology*, 1999, **38**, 679.
- 361 S. C. Bergmeier, D. J. Lapinsky, R. B. Free and D. B. McKay, *Bioorg. Med. Chem. Lett.*, 1999, **9**, 2263.
- 362 S. C. Bergmeier, K. A. Ismail, K. M. Arason, S. McKay, D. L. Bryant and D. B. McKay, *Bioorg. Med. Chem. Lett.*, 2004, **14**, 3739.
- 363 M. N. Sultankhodzhaev, M. T. H. Khan, M. Moin, M. I. Choudhary and Atta-ur-Rahman, *Nat. Prod. Res.*, 2005, **19**, 517.