Flavonoids: Promising Anticancer Agents

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DOI 10.1002/med.10033

Abstract: Flavonoids are polyphenolic compounds that are ubiquitously in plants. They have been shown to possess a variety of biological activities at nontoxic concentrations in organisms. The role of dietary flavonoids in cancer prevention is widely discussed. Compelling data from laboratory studies, epidemiological investigations, and human clinical trials indicate that flavonoids have important effects on cancer chemoprevention and chemotherapy. Many mechanisms of action have been identified, including carcinogen inactivation, antiproliferation, cell cycle arrest, induction of apoptosis and differentiation, inhibition of angiogenesis, antioxidation and reversal of multidrug resistance or a combination of these mechanisms. Based on these results, flavonoids may be promising anticancer agents. © 2003 Wiley Periodicals, Inc. Med Res Rev, 23, No. 4, 519–534, 2003

Key words: flavonoids; anticancer; mechanisms

1. INTRODUCTION

Flavonoids are a group of more than 4000 polyphenolic compounds that occur naturally in foods of plant origin. These compounds possess a common phenylbenzopyrone structure (C6-C3-C6), and they are categorized according to the saturation level and opening of the central pyran ring, mainly into flavones, flavanols, isoflavones, flavanols, flavanones, and flavanonols (Fig. 1).^{1,2}

Flavonoids have probably existed in the plant kingdom for over one billion years. They are present in practically all dietary plants, like fruits and vegetables (Table I). Therefore, they are consumed in considerable amounts and are also heat stable. It is estimated that the human intake of all flavonoids is a few hundreds of milligrams per day.³ Additionally, flavonoids are found in several medical plants, and herbal remedies containing flavonoids have been used in folk medicine around the world, especially in China.^{4–8} Licorice is the most used crude drug in Kampo medicines (traditional Chinese medicines modified in Japan). Flavonoids from licorice extract may be useful

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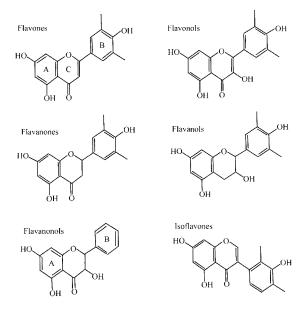


Figure 1. Chemical structures of the flavonoid family.

chemopreventive agents for peptic ulcer or gastric cancer in H. pylori-infected individuals.⁹ Sophoranone, extracted from a traditional Chinese medicine Shan Dou Gen, inhibited cell growth and induced apoptosis in various lines of cancer cells such as human stomach cancer MKN7 cells and human leukemia U937 cells.¹⁰ It was validated *in vivo* and *in vitro* that the *Crescentia alata* (Bignoniaceae), mainly containing flavonols rutin, kaempferol 3-*O*-rutinoside and kaempferol, was used in the traditional medicine of Guatemala as an anti-inflammatory remedy.¹¹ In folk medicine, the use of B. ferruginea stem bark for the treatment of rheumatic pains might be attributed to five of its constituents (3-*O*-methylquercetin, myricetin, ferrugin, quercetin 3-*O*-glucoside, and a biflavanol gallocatechin-[4'-*O*-7]-epigallocatechin) with xanthine oxidase inhibiting and superoxide scavenging activity.¹² Cirsimarin and cirsimaritin, flavonoids of Microtea debilis exhibiting adenosine antagonistic properties in rats, may partly explain the effectiveness of Microtea debilis against proteinuria in traditional medicine.¹³

These polyphenolic compounds display a remarkable spectrum of biological activities including those that might be able to influence processes that are dysregulated during cancer development. These include, for example, antiallergic, anti-inflammatory, antioxidant, antimutagenic, anticarcinogenic, and modulation of enzymatic activities.^{5,14–16} They may therefore have beneficial health

Flavonoid subgroup	Representative flavonoids	Major food sources	
Flavonols	Kaempherol, myricetin, quercetin, rutin	Onions, cherries, apples, broccoli, kale, tomato, berries, tea, red wine, tartary buckwheat	
Flavones	Apigenin, chrysin, luteolin	Parsley, thyme	
Isoflavones	Daidzein, genistein, glycitein, formononetin	Soya beans, legumes	
Flavanols	Catechin, gallocatechin	Apples, tea	
Flavanones	Eriodictyol, hesperitin, naringenin	Oranges, grapefruit	
Flavanonols	Taxifolin	Limon, aurantium	

Table I. Subclasses and Dietary Sources of Flavonoids

effects and can be considered possible chemopreventive or therapeutic agents against cancer.^{17,18} This review article will focus on the anticancer activity of flavonoids as well as their molecular mechanisms, since they are among the most promising anticancer agents.

2. EVIDENCE OF ANTICANCER EFFECTS

A. Epidemiological Data of Flavonoids

The weight of the epidemiological evidence for a protective effect of flavonoids against cancer is impressive. A growing number of epidemiological studies suggest that high flavonoid intake may be correlated with a decreased risk of cancer.¹⁹

More recently, in a population-based case-control study conducted in Shanghai from 1996–1998 which included 250 incident breast cancer cases and their individually matched controls, Dai et al.²⁰ reported that urinary excretion of total isoflavonoids and mammalian lignans was substantially lower in breast cancer cases than in controls (urine samples from breast cancer cases collected before cancer therapy). The median excretion rate of total isoflavonoids was 13.97 nmol/mg creatinine in cases and 23.09 in controls (P = 0.01), and that of total lignans was 1.77 in cases and 4.16 in controls (P < 0.01). This study strongly suggests a potential role of flavonoids in breast cancer preventing.

In a cohort study of 25-year follow-up on 9,959 Finnish men and women aged 15–99 years and initially cancer free, dietary intake of flavonoids was inversely associated with the incidence of cancer at all sites combined.²¹ The association was primarily due to the lower rates of lung cancer, with relative risk of 0.54 (highest vs. lowest quartiles), and was not attributed to the intake of vitamin E, vitamin C, beta-carotene, or total calories. Knekt and co-workers²² also estimated flavonoid intakes of 10,054 men and women mainly on the basis of the flavonoid concentrations in Finnish foods with a dietary history method. They found that men with higher quercetin intakes had a lower lung cancer incidence, and men with higher myricetin intakes had a lower prostate cancer risk. These data suggest a protective role of flavonoids against cancer.

A population-based case-control study in Hawaii further investigated the association between intake of flavonoids-powerful dietary and lung cancer risk. This study involved 582 patients with incident lung cancer and 582 age-, sex-, and ethnicity-matched control subjects. After adjusting for smoking and intake of saturated fat and beta-carotene, an inverse association was observed between lung cancer risk and the consumption of onions, apples, or white grapefruits as well as the calculated total intake of quercetin.²³ These results agree well with a former case-control study involving 541 cases of lung cancer and 540 hospitalized controls in Uruguay, but beta-carotene and vitamin E also associated with the reduction in risk of lung cancer.²⁴

In addition, the research group in Uruguay conducted a case-control study in the period of January 1996–December 1997, and found that flavonoids displayed a marked reduction by 70% in the risks of cancer of oral cavity, pharynx, larynx, and esophagus.²⁵ Another case-control study in Spain, including 354 cases of gastric cancer and 354 hospitalized controls, suggests that flavonoids such as quercetin and kaempferol may have protective effects against gastric cancer while specific carotenoids (alpha-carotene, beta-carotene, lutein, and lycopene) not.²⁶ A cohort of 34,651 postmenopausal cancer-free women aged 55–69 years were followed from 1986 to 1998. After adjustment for potential confounders, catechin intake was inversely associated with rectal cancer incidence only.²⁷ All these studies provide evidence for a protective role of flavonoids against cancer.

The intake of flavonoids is inversely associated with subsequent cancer in most but not all prospective epidemiological studies. There are few contrary reports²⁸⁻³⁰ that may be due to differences in bioavailability of the various flavonoids, and their effects on individual cancer sites cannot be excluded meriting further investigation.

B. In Vitro Studies of Flavonoids

Many researchers have conducted *in vitro* studies on the potential anticancer activity of flavonoids in diverse cell systems. The collected reports on the inhibitory properties of flavonoids against carcinogenesis are summarized in Table II.

Hirano and co-workers examined anticancer efficacy of 28 flavonoids on human acute myeloid leukemia cell line HL-60, and compared differences between antiproliferative activity and cytotoxicity of these compounds with those of four clinical anticancer agents. Eight of the 28 flavonoids showed considerable suppressive effects on HL-60 cell growth with IC50s ranging from 10-940 ng/ml. The flavonoid genistein had the strongest effects almost equivalent to the effects of current anticancer agents with little cytotoxicity against HL-60 cells, whereas the regular anticancer agents had potent cytotoxicity.⁵⁶

Kuntz et al.⁴⁸ screened more than 30 flavonoids for their effects on cell proliferation and potential cytotoxicity in human colon cancer cell lines Caco-2 and HT-29. Almost all compounds displayed antiproliferative activity without cytotoxicity. There was no obvious structure-activity relationship in the antiproliferative effects either on basis of the subclasses (i.e., isoflavones, flavones, flavonols, and flavonones) or with respect to kind or position of substituents within a class.⁴⁸

An array of 55 flavones having a variety of substituents was evaluated by Cushman and Nagarathnam for cytotoxicity in five cancer cell cultures, A-549 lung carcinoma, MCF-7 breast carcinoma, HT-29 colon adenocarcinoma, SKMEL-5 melanoma, and MLM melanoma. Fifteen of the 55 flavone derivatives were significantly active against at least one of these cell cultures.⁵⁷ In addition, seven of the 27 examined Citrus flavonoids were observed to inhibit the proliferation of tumor cells, while less active against normal human cells.⁵⁸

C. In Vivo Studies of Flavonoids

Flavonoids have been demonstrated to inhibit carcinogenesis *in vitro* and substantial evidence indicates that they can also do so *in vivo*.^{59–61} They may inhibit carcinogenesis by affecting the molecular events in the initiation, promotion, and progression stages. Animal studies and

Cancer	Cell	Flavonoid	References
Human oral cancer	HSC-2, HSG, SCC-25	Flavanones, isoflavans, EGC, chalcones, EGCG, curcumin, genistein, ECG, quercetin, cisplatin	Refs. [31–35]
Human breast cancer	MCF-7	Flavanones, daidzein, genistein, quercetin, luteolin	Refs. [36,37]
Human thyroid cancer	ARO, NPA, WRO	Genistein, apigenin, kaempferol, chrysin, luteolin, biochanin A	Refs. [38,39]
Human lung cancer	SK-LU1, SW900, H441, H661, haGo-K-1, A549	Flavone, quercetin	Refs. [40,41]
Human prostate cancer	LNCaP, PC3, DU145	Catechin, epicatechin, quercetin, kaempferol, luteolin, genistein, apigenin, myricetin, silymarin	Refs. [42–45]
Human colon cancer	Caco-2, HT-29, IEC-6, HCT-15	Flavone, quercetin, genistein, anthocyanin	Refs. [46-50]
Human leukaemia	HL-60, K562, Jurkat	Apigenin, quercetin, myricetin, chalcones	Refs. [51–54]
B16 mouse melanoma	4A5	Chalcones	Ref. [55]

Table II. Anticancer Activities of Flavonoids in Various Cancer Cell Lines

investigations using different cellular models suggested that certain flavonoids could inhibit tumor initiation as well as tumor progression.^{62–68}

A recent study showed that fermented soy milk containing larger amounts of genistein and daidzein than unfermented one and isoflavone mixtures, given to rats starting at 7 weeks of age, inhibited mammary tumorigenesis induced by 2-amino-1-methyl-6-phenylimidazo [4,5-*b*] pyridine (PhIP).⁶⁹

Dietary quercetin inhibited DMBA-induced carcinogenesis in hamster buccal pouch⁷⁰ and in rat mammary gland.⁷¹ When given during the initiation stage, quercetin and ellagic acid, also inhibited DEN-induced lung tumorigenesis in mice.⁷² In a medium-term multiorgan carcinogenesis model in rats, quercetin (1% in the diet) inhibited tumor promotion in the small intestine.⁷³ Feeding rats with quercetin or chalcone and 2-hydroxychalcone (0.05% in the diet), during either the initiation or promotion stage, inhibited 4-NQO-induced carcinoma formation in the tongue. These compounds also decreased cell proliferation and polyamine levels.⁶⁷

Siess and co-workers investigated the effects of feeding rats with flavone, flavanone, tangeretin, and quercetin on two steps of aflatoxin B1 (AFB1)-induced hepatocarcinogenesis (initiation and promotion) and found that flavone, flavanone and tangeretin administered through the initiation period decreased the number of gamma-glutamyl transpeptidase-preneoplastic foci. Furthermore, feeding rats with flavanone during the phenobarbital-induced promotion step significantly reduced the areas of placental glutathione S-transferase preneoplastic foci. Therefore flavanone acts as an anti-initiator as well as an antipromotor.⁷⁴

Inhibition of lung tumorigenesis by tea preparations rich in catechin has been demonstrated in A/J mice.⁷⁵ Administration of decaffeinated green or black tea to mice (as the sole source of drinking fluid) for 3 weeks starting 2 weeks before the 4-(methylnitrosamine)-1-(3-pyridyl)-1 butanone (NNK) treatment, or for 15 weeks starting 1 week after the NNK treatment, markedly reduced the number of tumors formed in the mice. In mice that had already developed adenomas at 16 weeks after the NNK injection, the progression of adenomas to adenocarcinomas was significantly inhibited by the administration of black tea from weeks 16–52. These experiments indicate that tea has broad inhibitory activity against lung carcinogenesis, and it is effective when administered during the initiation, promotion, or progression stages of carcinogenesis. Moreover, there is evidence for the suppression of tumor invasion and metastasis by flavonoids.

Catechins, a group of flavonoid molecules, inhibit invasion of mouse MO4 cells into embryonic chick heart fragments *in vitro*.⁷⁶ A polymethoxy flavonoid, nobiletin, from Citrus depressa inhibited the tumor-invasive activity of human fibrosarcoma HT-1080 cells in the Matrigel model, which was likely through suppressing the expression of matrix metalloproteases (MMPs) and augmenting of tissue inhibitors of metalloproteinases (TIMPs) production in tumor cells.⁷⁷

When given i.p., quercetin and apigenin inhibited melanoma cell (B16-BL6) growth and metastatic potential in syngenetic mice, and interestingly, they significantly decreased the invasion of B16-BL6 cells *in vitro*.⁶¹ When given s.c., apigenin (0.75 or 1.5 mg/kg body weight) significantly decreased the incidence of lymphatic vessel invasion of intestinal adenocarcinomas induced by azoxymethane, and that of cancer peritoneal metastasis enhanced by bombesin in male Wistar rats. The inhibitory effect of apigenin on cancer metastasis may be through inhibition of phosphorylation of mitogen-activated protein kinase (MAPK).⁷⁸

D. Human Clinical Trials With Flavonoids

The encouraging results of anticancer effects in preclinical studies have stimulated the clinical trials of flavonoids in human.

Early in 1988, Weiss et al. conducted a Phase I and pharmacological study of flavone acetic acid (FAA), one of a series of novel flavonoids.⁷⁹

A phase I, dose-escalation trial of quercetin (3,3',4',5,7-pentahydroxy-flavone), a naturally occurring flavonoid with many biological activities including inhibition of a number of tyrosine kinases, was performed by Ferry et al. Intravenous quercetin was found to inhibit lymphocyte tyrosine kinase in nine of 11 patients assayed. One hepatocellular carcinoma patient had a sustained (150 days) fall in serum alpha-fetoprotein and alkaline phosphatase during and after four low-dose, intravenous quercetin treatments (60 mg/m²) on a 3-week schedule. Another patient with stage four ovarian cancer who had not responded to six courses of cyclophosphamide/cisplatin chemotherapy had a fall in the CA125 tumor marker from 295 to 55 units/ml following two treatments of intravenous quercetin (420 mg/m²) 3 weeks apart. The authors recommend 1400 mg/m² as the bolus dose, which may be given either in 3-week or weekly intervals, for Phase II trials. They defined the maximum tolerated dose (MTD) as 1700 mg/m² three weekly, but the vehicle, dimethyl sulphoxide (DMSO) is unsuitable for further clinical development of quercetin.⁸⁰ After that, a synthetic watersoluble, pro-drug of quercetin (3'(N-carboxymethyl) carbomyl-3,4',5,7-tetrahydroxyflavone), QC12 was studied in an initial phase I trial by this research group. The authors suggested this water-soluble pro-drug warrant further clinical investigation, starting with a formal phase I, IV, dose-escalation study.81

Flavopiridol is a novel semisynthetic flavone analogue of rohitukine, a leading anticancer compound from an Indian tree. Flavopiridol inhibits most cyclin-dependent kinases (CDKs) and displays unique anticancer properties. It is the first CDKs inhibitor to be tested in human clinical trials by Aventis Pharma (formerly Hoechst Marion Roussel) and the National Cancer Institute (NCI) for the potential treatment of cancer and proliferative disorders. Initial human clinical trials with infusional flavopiridol demonstrated activity in some patients with non-Hodgkin's lymphoma, renal, prostate, colon, and gastric carcinomas. By July 1999, the compound had entered phase II trials for gastric cancer and leukemia (CLL), and phase I/II trials for esophageal tumor and non-small cell lung cancer (NSCLC). Phase II trials for colon and renal cancer were also reported.^{82,83}

Wang¹⁸ extensively reviewed the therapeutic potential in human of four most widely investigated flavonoids: flavopiridol, catechins, genistein, and quercetin. According to his another report,⁸⁴ by May 2001, flavopiridol was in phase IIa trials and had achieved proof-of-concept in phase I/IIa trials as a monotherapy. Moreover, it was expected that the product be launched by 2003.

3. MAJOR MOLECULAR MECHANISMS OF ACTION

A. Preventing Carcinogen Metabolic Activation

Studies *in vitro* and *in vivo* have shown that some flavonoids modulate the metabolism and disposition of carcinogens and can contribute to cancer prevention.^{85–89}

One important mechanism by which flavonoids may exert their effects is through their interaction with phase I metabolizing enzymes (e.g., cytochrome P450), which metabolically activate a large number of procarcinogens to reactive intermediates that can interact with cellular nucleophiles and ultimately trigger carcinogenesis. Flavonoids are demonstrated to inhibit the activities of certain P450 isozymes such as CYP1A1 and CYP1A2.^{23,90,91} Thus, they are likely to have a protective role against the induction of cellular damage by the activation of carcinogens.

Another mechanism of action is the induction of phase II metabolizing enzymes such as glutathione-S-transferase, quinone reductase, and UDP-glucuronyl transferase,^{92,93} by which carcinogens are detoxified and thus more readily eliminated from the body. This would also help explain the chemopreventive effects of flavonoids against carcinogenesis.

Moreover, some flavonoids have been reported as potent aromatase inhibitors.^{94–97} Substantial evidence supports the concept that estrogens be involved in mammary carcinomas. Estradiol, the most potent endogenous estrogen, is biosynthesized from androgens by the cytochrome P450 enzyme complex called aromatase. Inhibition of aromatase is an important approach for reducing growth

stimulatory effects of estrogens in hormone-dependent breast cancer.⁹⁴ Therefore, flavonoids could be considered potential agents against breast cancer through the inhibition of aromatase activity.

B. Antiproliferation

Dysregulated proliferation appears to be a hallmark of susceptibility to neoplasia. Cancer prevention is generally associated with inhibition, reversion or retardation of cellular hyperproliferation. Most flavonoids have been demonstrated to inhibit proliferation in many kinds of cultured human cancer cell lines, whereas less or no toxic to human normal cells.^{36,46,48,56,58}

The molecular mechanism of antiproliferation may involve the inhibition of the prooxidant process that causes tumor promotion. It is generally believed that the formation of growth promoting oxidants (reactive oxygen species, ROS) is a major "catalyst" of the tumor promotion and progression stages, which follow the initiation stage (carcinogen metabolic activation to mutagens). The prooxidant enzymes induced or activated by various tumor promoters, for example, phorbol esters, include the arachidonate metabolizing enzymes, cyclooxygenases (COX), and lipoxygenases (LOX). Flavonoids are particularly effective at inhibiting xanthine oxidase,^{98,99} COX or LOX^{55,100,101} and therefore inhibit tumor cell proliferation.

In addition, inhibition of polyamine biosynthesis could be a contributing mechanism to the antiproliferative activities of flavonoids. Ornithine decarboxylase is a rate-limiting enzyme in polyamine biosynthesis, which has been correlated with the rate of DNA synthesis and cell proliferation in several tissues. Several experiments show that flavonoids can inhibit ornithine decarboxylase induced by tumor promoters, and thus cause a subsequent decrease in polyamine and inhibition of DNA/protein synthesis.^{63,66,67}

Furthermore, flavonoids are also effective at inhibiting signal transduction enzymes, for example, protein tyrosine kinase (PTK),^{80,101} protein kinase C (PKC),¹⁰² and phosphoinositide 3-kinases (PIP₃),^{77,103} which are involved in the regulation of cell proliferation.

C. Cell Cycle Arrest

Perturbations in cell cycle progression may account for the anticarcinogenic effects of flavonoids. Mitogenic signals commit cells to entry into a series of regulated steps allowing traverse of the cell cycle. Synthesis of DNA (S phase) and separation of two daughter cells (M phase) are the main features of cell cycle progression. The time between the S and M phases is known as G2 phase. This phase is important to allow cells to repair errors that occur during DNA duplication, preventing the propagation of these errors to daughter cells. In contrast, the G1 phase represents the period of commitment to cell cycle progression that separates M and S phases as cells prepare for DNA duplication upon mitogenic signals.

CDKs have been recognized as key regulators of cell cycle progression. Alteration and deregulation of CDK activity are pathogenic hallmarks of neoplasia. A number of cancers are associated with hyperactivation of CDKs as a result of mutation of the CDK genes or CDK inhibitor genes. Therefore, inhibitors or modulators would be of interest to explore as novel therapeutic agents in cancer.^{82,83}

Checkpoints at both G1/S and G2/M of the cell cycle in cultured cancer cell lines have been found to be perturbed by flavonoids such as silymarin, genistein, quercetin, daidzein, luteolin, kaempferol, apigenin, and epigallocatechin 3-gallate.^{104–106} Studies from different laboratories revealed that flavopiridol could induce cell cycle arrest during either G1 or G2/M by the inhibition of all CDKs thus far examined.^{18,82}

D. Induction of Apoptosis

The significant anticancer properties observed of flavonoids may be due to frank apoptosis.^{33,38,46,48,51,54,55} Apoptosis is an active form of cell death that plays an essential role in the development and survival by eliminating damaged or otherwise unwanted cell. It is tightly regulated by a set of genes that either promote apoptosis or promote cell survival, and is mediated through a highly organized network of interacting protease and their inhibitors in response to noxious stimuli from either inside or outside of the cell. Dysregulation of apoptosis could play a critical role in oncogenesis. A series of recent studies have demonstrated that most, if not all, chemotherapeutic agents exert their tumoricidal effects by inducing apoptosis in target cells and tissues.

Flavonoids have been shown to induce apoptosis in some cancer cell lines, while sparing normal cells. The molecular mechanisms by which flavonoids induce apoptosis have not yet been clarified. Several mechanisms may be involved, including inhibition of DNA topoisomerase I/II activity,^{51,101,107,108} decrease of reactive oxygen species (ROS),¹⁰⁹ regulation of heat shock proteins expression,¹¹⁰ modulation of signaling pathways,³⁸ release of cytochrome c with a subsequent activation of caspase-9 and caspase-3,⁵¹ downregulation of Bcl-2 and Bcl-X(L) expression but promotion of Bax and Bak expression, nuclear transcription factor kappaB (NF-kappaB), activation of endonuclease, and suppression of Mcl-1 protein.^{46,55,109,111}

Preliminary evidence from our laboratory that apoptosis induced by tartary buckwheat flavonoid in HL-60 cells may be associated with early activation of caspase-3,¹¹² likely mediated through Fas and cytochrome c pathways, as well as regulated through the inactivation of NF-kappaB (manuscript submitted).

E. Promotion of Differentiation

In addition to the anticancer properties mentioned above, it is of interest that certain flavonoids cause undifferentiated cancer cell lines to differentiate into cells exhibiting mature phenotypic characteristics.^{113,114}

The flavones genistein, apigenin, luteolin, quercetin, and phloretin were found to induce differentiation of human acute myelogenous leukemia HL-60 cells into granulocytes and monocytes.¹¹⁵ The isoflavone daidzein was also capable of doing so.¹¹⁶ Erythroid differentiation of the human myelogenous leukemia K562 cell line and a multidrug-resistant subline (K562R) could also be induced by genistein.^{117,118} Moreover, flavone was shown to induce differentiation in HT-29 human colon cancer cells.⁴⁶

Cancers arise from cells harboring mutations that relinquish the need for exogenous growth factors. Deregulation of growth control ultimately leads to selection of clonal lines of cells that replicate at embryonic pace and yet fail to respond to differentiation and maturation signals. Non-physiological inducers of terminal differentiation have been used as novel therapies for the prevention and therapy of cancer. Induction of terminal differentiation by flavonoids may lead to the eventual elimination of tumorigenic cells and rebalance of normal cellular homeostasis. Thus, these compounds could be developed into promising anticancer agents.

F. Antioxidative Activity

Dietary flavonoids are natural antioxidants.¹¹⁹ They may be against cancer through limit of damaging oxidative reactions in cells, which may predispose to the development of cancer. Oxygen-derived free radicals appear to possess the propensity to initiate as well as to promote carcinogenesis. Lipid peroxidation products originating from dying cells could also exert a cancer promotional effect.^{120,121} Oxidation of DNA is likely to be an important cause of mutation that potentially can be reduced by antioxidants.¹²²

Flavonoids are chemically one-electron donors. They serve as derivatives of conjugated ring structures and hydroxyl groups that have the potential to function as antioxidants in *in vitro* cell culture or cell free systems by scavenging superoxide anion, singlet oxygen, lipid peroxy-radicals, and/or stabilizing free radicals involved in oxidative processes through hydrogenation or complexing with oxidizing species. *In vitro* studies are able to demonstrate for flavonols, flavones, and most recently

also for anthocyanins a considerable antioxidative activity, mainly based on scavenging of oxygen radicals.¹²² Theoretical underpinnings for the efficacy of flavonoids as antioxidants *in vivo* come from the inhibition of low-density lipoprotein (LDL) oxidation, likely due to their reductive capacity and protein-binding properties.¹²³

G. Inhibition of Angiogenic Process

Flavonoids are known as angiogenesis inhibitors derived from natural sources.¹²⁴ The abilities of particular flavonoids to block solid tumor growth may be due to their inhibition of the neoangiogenic process.

Angiogenesis is a strictly controlled process in the healthy adult human body, which is regulated by a variety of endogenous angiogenic and angiostatic factors. However, pathological angiogenesis can occur in cancer. When deprived of proper vascularization, the high proliferation rate in the tumor would be balanced by cell death due to the lack of diffusion of nutrients and oxygen. Angiogenesis inhibitors such as flavonoids, are able to interfere with various steps of angiogenesis, like basement destruction of blood vessels, proliferation and migration of endothelial cells, or the lumen formation. Therefore, these compounds may have potential for the treatment of solid tumors.^{125,126}

H. Modulation of Multidrug Resistance

Multidrug resistance due to P-glycoprotein (Pgp) or multidrug resistance associated protein (MRP) is a serious impediment to successful chemotherapy of cancer. Much effort has been spent to modulate multidrug resistance in the different species by using specific inhibitors, but generally with little success due to additional cellular targets and/or extrusion of the potential inhibitors.

Certain flavonoids have been reported to possess potent inhibitory activity against the drugexporting function of Pgp, a plasma membrane ATP-binding cassette (ABC) transporter that extrudes cytotoxic drugs at the expense of ATP hydrolysis. Pgp consists of two homologous halves each containing a transmembrane domain (TMD) involved in drug binding and efflux, and a cytosolic nucleotide-binding domain (NBD) involved in ATP binding and hydrolysis, with an overall (TMD-NBD)2 domain topology. Modulation by flavonoids of cell multidrug resistance mediated by Pgp may be through (i) inhibiting the overexpression of multidrug resistance gene-1 (MDR1),¹²⁷ (ii) direct binding to NBDs with high affinity,¹²⁸ (iii) inhibiting ATPase activity, nucleotide hydrolysis and energy-dependent drug interaction with transporter-enriched membranes.^{129,130} Acting through Pgp as a possible target, flavonoids are found to enhance doxorubicin (DOX) induced antitumor activity and increase the DOX concentrations in tumors. Thus, the unique property of reversal of multidrug resistance of these compounds might help protect against multidrug-resistant tumors.^{54,131–133}

4. CONCLUSIONS

Flavonoids are generally nontoxic and manifest a diverse range of beneficial biological activities. The role of dietary flavonoids in cancer prevention is widely discussed. There is much evidence that flavonoids have important effects on inhibiting carcinogenesis.

Epidemiological studies have provided data that high dietary intake of flavonoids with fruits and vegetables could be associated with a low cancer prevalence in humans. This is supported by a multitude of *in vitro* and *in vivo* studies, which show that flavonoids may inhibit various stages in the carcinogenesis process, namely tumor initiation, promotion, and progression. Based on the studies *in vivo* and *in vitro*, many mechanisms of action may be involved. These include carcinogen inactivation, antiproliferation, cell cycle arrest, induction of apoptosis and differentiation, inhibition of angiogenesis, antioxidation and reversal of multidrug resistance or a combination of these

mechanisms. Furthermore, the intriguing results from laboratory and epidemiological studies have stimulated the development of flavonoids in human clinical trials.

While these experiences strengthen the notion that flavonoids could be useful anticancer agents, to date few clinical studies have demonstrated that these bioflavonoids retain anticancer properties in humans *in vivo*. In addition, clinical trials available have required intravenously administered flavonoids at concentrations around 1400 mg/m² before effects are seen. These plasma concentrations are unlikely to be achieved using the dietary supplements currently available. Therefore, more focused clinical studies are required to establish whether such dietary effects of these compounds can be exploited to achieve cancer preventive or therapeutic effects in human.

In conclusion, considering that many chemotherapeutic agents against tumor cells without sparing normal cells remain a major obstacle and development of multidrug resistance further limits chemotherapy in cancer, the promising results will stimulate the development of flavonoids for cancer chemoprevention and chemotherapy.

A C K N O W L E D G M E N T S

We thank Prof. Chung S. Yang, Diane F. Birt, John D. Robertson, MH Siess, and Adrian M. Senderowicz for sending their reprints to us. We thank Mr. Quan Shen for his excellent drawing of the figures.

REFERENCES

- Middleton E, Jr., Kandaswami C. The impact of plant flavonoids on mammalian biology: Implications for immunity, inflammation and cancer. In: Harborne JB, editor. The flavonoids advances in research since 1986. 1st edition. London: Chapman and Hall; 1994. p 619–652.
- Harborne JB, Williams CA. Advances in flavonoid research since 1992. Phytochemistry 2000;55:481– 504.
- 3. Hollman PC, Katan MB. Dietary flavonoids: Intake, health effects and bioavailability. Food Chem Toxicol 1999;37:937–942.
- Di Carlo G, Mascolo N, Izzo AA, Capasso F. Flavonoids: Old and new aspects of a class of natural therapeutic drugs. Life Sci 1999;65:337–353.
- 5. Craig WJ. Health-promoting properties of common herbs. Am J Clin Nutr 1999;70:491S–499S.
- Kadarian C, Broussalis AM, Mino J, Lopez P, Gorzalczany S, Ferraro G, Acevedo C. Hepatoprotective activity of Achyrocline satureioides(Lam) D. C. Pharmacol Res 2002;45:57–61.
- 7. Pascual ME, Slowing K, Carretero E, Sanchez Mata D, Villar A. Lippia: Traditional uses, chemistry and pharmacology: A review. J Ethnopharmacol 2001;76:201–214.
- Samuelsen AB. The traditional uses, chemical constituents and biological activities of Plantago major L. A review. J Ethnopharmacol 2000;71:1–21.
- 9. Fukai T, Marumo A, Kaitou K, Kanda T, Terada S, Nomura T. Anti-helicobacter pylori flavonoids from licorice extract. Life Sci 2002;71:1449–1463.
- Kajimoto S, Takanashi N, Kajimoto T, Xu M, Cao J, Masuda Y, Aiuchi T, Nakajo S, Ida Y, Nakaya K. Sophoranone, extracted from a traditional Chinese medicine Shan Dou Gen, induces apoptosis in human leukemia U937 cells via formation of reactive oxygen species and opening of mitochondrial permeability transition pores. Int J Cancer 2002;99:879–890.
- Autore G, Rastrelli L, Lauro MR, Marzocco S, Sorrentino R, Sorrentino U, Pinto A, Aquino R. Inhibition of nitric oxide synthase expression by a methanolic extract of Crescentia alata and its derived flavonols. Life Sci 2001;70:523–534.
- 12. Cimanga K, Ying L, De Bruyne T, Apers S, Cos P, Hermans N, Bakana P, Tona L, Kambu K, Kalenda DT, Pieters L, Vanden Berghe D, Vlietinck AJ. Radical scavenging and xanthine oxidase inhibitory activity of phenolic compounds from Bridelia ferruginea stem bark. J Pharm Pharmacol 2001;53:757–761.
- Hasrat JA, De Bruyne T, De Backer JP, Vauquelin G, Vlietinck AJ. Cirsimarin and cirsimaritin, flavonoids of Microtea debilis (Phytolaccaceae) with adenosine antagonistic properties in rats: Leads for new therapeutics in acute renal failure. J Pharm Pharmacol 1997;49:1150–1156.

- Middleton E, Jr., Kandaswami C, Theoharides TC. The effects of plant flavonoids on mammalian cells: Implications for inflammation, heart disease, and cancer. Pharmacol Rev 2000;52:673–751.
- Galati G, Teng S, Moridani MY, Chan TS, O'Brien PJ. Cancer chemoprevention and apoptosis mechanisms induced by dietary polyphenolics. Drug Metabol Drug Interact 2000;17:311–349.
- Yang CS, Landau JM, Huang MT, Newmark HL. Inhibition of carcinogenesis by dietary polyphenolic compounds. Annu Rev Nutr 2001;21:381–406.
- Birt DF, Hendrich S, Wang W. Dietary agents in cancer prevention: Flavonoids and isoflavonoids. Pharmacol Ther 2001;90:157–177.
- 18. Wang HK. The therapeutic potential of flavonoids. Expert Opin Invest Drugs 2000;9:2103–2119.
- Le Marchand L. Cancer preventive effects of flavonoids—A review. Biomed Pharmacother 2002;56:296– 301.
- Dai Q, Franke AA, Jin F, Shu XO, Hebert JR, Custer LJ, Cheng J, Gao YT, Zheng W. Urinary excretion of phytoestrogens and risk of breast cancer among Chinese women in Shanghai. Cancer Epidemiol Biomarkers Prev 2002;11:815–821.
- 21. Knekt P, Jarvinen R, Seppanen R, Hellovaara M, Teppo L, Pukkala E, Aromaa A. Dietary flavonoids and the risk of lung cancer and other malignant neoplasms. Am J Epidemiol 1997;146:223–230.
- 22. Knekt P, Kumpulainen J, Jarvinen R, Rissanen H, Heliovaara M, Reunanen A, Hakulinen T, Aromaa A. Flavonoid intake and risk of chronic diseases. Am J Clin Nutr 2002;76:560–568.
- Le Marchand L, Murphy SP, Hankin JH, Wilkens LR, Kolonel LN. Intake of flavonoids and lung cancer. J Natl Cancer Inst 2000;92:154–160.
- Stefani ED, Boffetta P, Deneo-Pellegrini H, Mendilaharsu M, Carzoglio JC, Ronco A, Olivera L. Dietary antioxidants and lung cancer risk: A case-control study in Uruguay. Nutr Cancer 1999;34:100–110.
- 25. De Stefani E, Ronco A, Mendilaharsu M, Deneo-Pellegrini H. Diet and risk of cancer of the upper aerodigestive tract-II. Nutrients. Oral Oncol 1999;35:22–26.
- Garcia-Closas R, Gonzalez CA, Agudo A, Riboli E. Intake of specific carotenoids and flavonoids and the risk of gastric cancer in Spain. Cancer Causes Control 1999;10:71–75.
- Arts IC, Jacobs DR, Jr., Gross M, Harnack LJ, Folsom AR. Dietary catechins and cancer incidence among postmenopausal women: The Iowa Women's Health Study (United States). Cancer Causes Control 2002; 13:373–382.
- 28. Garcia R, Gonzalez CA, Agudo A, Riboli E. High intake of specific carotenoids and flavonoids does not reduce the risk of bladder cancer. Nutr Cancer 1999;35:212–214.
- 29. Arts IC, Hollman PC, Bueno De Mesquita HB, Feskens EJ, Kromhout D. Dietary catechins and epithelial cancer incidence: The Zutphen elderly study. Int J Cancer 2001;92:298–302.
- Garcia-Closas R, Agudo A, Gonzalez CA, Riboli E. Intake of specific carotenoids and flavonoids and the risk of lung cancer in women in Barcelona, Spain. Nutr Cancer 1998;32:154–158.
- 31. Shi YQ, Fukai T, Sakagami H, Chang WJ, Yang PQ, Wang FP, Nomura T. Cytotoxic flavonoids with isoprenoid groups from Morus mongolica. J Nat Prod 2001;64:181–188.
- Fukai T, Sakagami H, Toguchi M, Takayama F, Iwakura I, Atsumi T, Ueha T, Nakashima H, Nomura T. Cytotoxic activity of low molecular weight polyphenols against human oral tumor cell lines. Anticancer Res 2000;20:2525–2536.
- 33. Sakagami H, Jiang Y, Kusama K, Atsumi T, Ueha T, Toguchi M, Iwakura I, Satoh K, Fukai T, Nomura T. Induction of apoptosis by flavones, flavonols (3-hydroxyflavones) and isoprenoid-substituted flavonoids in human oral tumor cell lines. Anticancer Res 2000;20:271–277.
- Elattar TM, Virji AS. Effect of tea polyphenols on growth of oral squamous carcinoma cells in vitro. Anticancer Res 2000;20:3459–3465.
- Elattar TM, Virji AS. The inhibitory effect of curcumin, genistein, quercetin and cisplatin on the growth of oral cancer cells in vitro. Anticancer Res 2000;20:1733–1738.
- Pouget C, Lauthier F, Simon A, Fagnere C, Basly JP, Delage C, Chulia AJ. Flavonoids: Structural requirements for antiproliferative activity on breast cancer cells. Bioorg Med Chem Lett 2001;11:3095– 3097.
- 37. Han D, Tachibana H, Yamada K. Inhibition of environmental estrogen-induced proliferation of human breast carcinoma MCF-7 cells by flavonoids. In Vitro Cell Dev Biol Anim 2001;37:275–282.
- Yin F, Giuliano AE, Van Herle AJ. Signal pathways involved in apigenin inhibition of growth and induction of apoptosis of human anaplastic thyroid cancer cells (ARO). Anticancer Res 1999;19:4297–4303.
- Yin F, Giuliano AE, Van Herle AJ. Growth inhibitory effects of flavonoids in human thyroid cancer cell lines. Thyroid 1999;9:369–376.
- 40. Bai F, Matsui T, Ohtani-Fujita N, Matsukawa Y, Ding Y, Sakai T. Promoter activation and following induction of the p21/WAF1 gene by flavone is involved in G1 phase arrest in A549 lung adenocarcinoma cells. FEBS Lett 1998;437:61–64.

- 530 REN ET AL.
- 41. Caltagirone S, Ranelletti FO, Rinelli A, Maggiano N, Colasante A, Musiani P, Aiello FB, Piantelli M. Interaction with type II estrogen binding sites and antiproliferative activity of tamoxifen and quercetin in human non-small-cell lung cancer. Am J Respir Cell Mol Biol 1997;17:51–59.
- 42. Knowles LM, Zigrossi DA, Tauber RA, Hightower C, Milner JA. Flavonoids suppress androgenindependent human prostate tumor proliferation. Nutr Cancer 2000;38:116–122.
- 43. Bhatia N, Agarwal R. Detrimental effect of cancer preventive phytochemicals silymarin, genistein and epigallocatechin 3-gallate on epigenetic events in human prostate carcinoma DU145 cells. Prostate 2001;46:98–107.
- 44. Kampa M, Hatzoglou A, Notas G, Damianaki A, Bakogeorgou E, Gemetzi C, Kouroumalis E, Martin PM, Castanas E. Wine antioxidant polyphenols inhibit the proliferation of human prostate cancer cell lines. Nutr Cancer 2000;37:223–233.
- 45. Agarwal R. Cell signaling and regulators of cell cycle as molecular targets for prostate cancer prevention by dietary agents. Biochem Pharmacol 2000;60:1051–1059.
- 46. Wenzel U, Kuntz S, Brendel MD, Daniel H. Dietary flavone is a potent apoptosis inducer in human colon carcinoma cells. Cancer Res 2000;60:3823–3831.
- 47. Kamei H, Hashimoto Y, Koide T, Kojima T, Hasegawa M. Anti-tumor effect of methanol extracts from red and white wines. Cancer Biother Radiopharm 1998;13:447–452.
- 48. Kuntz S, Wenzel U, Daniel H. Comparative analysis of the effects of flavonoids on proliferation, cytotoxicity, and apoptosis in human colon cancer cell lines. Eur J Nutr 1999;38:133–142.
- 49. Kuo SM, Morehouse HF, Jr., Lin CP. Effect of antiproliferative flavonoids on ascorbic acid accumulation in human colon adenocarcinoma cells. Cancer Lett 1997;116:131–137.
- Kuo SM. Antiproliferative potency of structurally distinct dietary flavonoids on human colon cancer cells. Cancer Lett 1996;110:41–48.
- Wang IK, Lin-Shiau SY, Lin JK. Induction of apoptosis by apigenin and related flavonoids through cytochrome c release and activation of caspase-9 and caspase-3 in leukaemia HL-60 cells. Eur J Cancer 1999;35:1517–1525.
- 52. Chung HS, Chang LC, Lee SK, Shamon LA, van Breemen RB, Mehta RG, Farnsworth NR, Pezzuto JM, Kinghorn AD. Flavonoid constituents of Chorizanthe diffusa with potential cancer chemopreventive activity. J Agric Food Chem 1999;47:36–41.
- 53. Csokay B, Prajda N, Weber G, Olah E. Molecular mechanisms in the antiproliferative action of quercetin. Life Sci 1997;60:2157–2163.
- 54. De Vincenzo R, Ferlini C, Distefano M, Gaggini C, Riva A, Bombardelli E, Morazzoni P, Valenti P, Belluti F, Ranelletti FO, Mancuso S, Scambia G. In vitro evaluation of newly developed chalcone analogues in human cancer cells. Cancer Chemother Pharmacol 2000;46:305–312.
- Iwashita K, Kobori M, Yamaki K, Tsushida T. Flavonoids inhibit cell growth and induce apoptosis in B16 melanoma 4A5 cells. Biosci Biotechnol Biochem 2000;64:1813–1820.
- 56. Hirano T, Gotoh M, Oka K. Natural flavonoids and lignans are potent cytostatic agents against human leukemic HL-60 cells. Life Sci 1994;55:1061–1069.
- 57. Cushman M, Nagarathnam D. Cytotoxicities of some flavonoid analogues. J Nat Prod 1991;54:1656– 1660.
- Kawaii S, Tomono Y, Katase E, Ogawa K, Yano M. Antiproliferative activity of flavonoids on several cancer cell lines. Biosci Biotechnol Biochem 1999;63:896–899.
- Miyagi Y, Om AS, Chee KM, Bennink MR. Inhibition of azoxymethane-induced colon cancer by orange juice. Nutr Cancer 2000;36:224–249.
- Kamei H, Koide T, Kojimam T, Hasegawa M, Terabe K, Umeda T, Hashimoto Y. Flavonoid-mediated tumor growth suppression demonstrated by in vivo study. Cancer Biother Radiopharm 1996;11:193– 196.
- Caltagirone S, Rossi C, Poggi A, Ranelletti FO, Natali PG, Brunetti M, Aiello FB, Piantelli M. Flavonoids apigenin and quercetin inhibit melanoma growth and metastatic potential. Int J Cancer 2000;87:595– 600.
- 62. Deschner EE, Ruperto J, Wong G, Newmark HL. Quercetin and rutin as inhibitors of azoxymethanolinduced colonic neoplasia. Carcinogenesis 1991;12:1193–1196.
- 63. Tanaka T, Makita H, Kawabata K, Mori H, Kakumoto M, Satoh K, Hara A, Sumida T, Tanaka T, Ogawa H. Chemoprevention of azoxymethane-induced rat colon carcinogenesis by the naturally occurring flavonoids, diosmin and hesperidin. Carcinogenesis 1997;18:957–965.
- 64. Mukhtar H, Agarwal R. Skin cancer chemoprevention. J Investig Dermatol Symp Proc 1996;1:209-214.
- 65. Yang M, Tanaka T, Hirose Y, Deguchi T, Mori H, Kawada Y. Chemopreventive effects of diosmin and hesperidin on N-butyl-N-(4-hydroxybutyl)nitrosamine-induced urinary-bladder carcinogenesis in male ICR mice. Int J Cancer 1997;73:719–724.

- 66. Tanaka T, Makita H, Ohnishi M, Mori H, Satoh K, Hara A, Sumida T, Fukutani K, Tanaka T, Ogawa H. Chemoprevention of 4-nitroquinoline 1-oxide-induced oral carcinogenesis in rats by flavonoids diosmin and hesperidin, each alone and in combination. Cancer Res 1997;57:246–252.
- 67. Makita H, Tanaka T, Fujitsuka H, Tatematsu N, Satoh K, Hara A, Mori H. Chemoprevention of 4-nitroquinoline 1-oxide-induced rat oral carcinogenesis by the dietary flavonoids chalcone, 2-hydroxychalcone, and quercetin. Cancer Res 1996;56:4904–4909.
- 68. Elangovan V, Sekar N, Govindasamy S. Chemopreventive potential of dietary bioflavonoids against 20-methylcholanthrene-induced tumorigenesis. Cancer Lett 1994;87:107–113.
- 69. Ohta T, Nakatsugi S, Watanabe K, Kawamori T, Ishikawa F, Morotomi M, Sugie S, Toda T, Sugimura T, Wakabayashi K. Inhibitory effects of bifidobacterium-fermented soy milk on 2-amino-1-methyl-6-phenylimidaz[4,5-b]pyridine-induced rat mammary carcinogenesis, with a partial contribution of its component isoflavones. Carcinogenesis 2000;21:937–941.
- Balasubramanian S, Govindasamy S. Inhibitory effect of dietary flavonol quercetin on 7,12dimethylbanz[a]anthracene-induced hamster buccal pouch carcingoenesis. Carcinogenesis 1996;17: 877–879.
- 71. Verma AK, Johnson JA, Gould MN, Tanner MA. Inhibition of 7,12 dimethylbenz(a) anthracene and N-nitrosomethylurea induced rat mammary cancer by dietary flavonol quercetin. Cancer Res 1988;48:5754–5758.
- 72. Khanduja KL, Gandhi RK, Pathania V, Syal N. Prevention of N-nitrosodiethylamine-induced lung tumorigenesis by ellagic acid and quercetin in mice. Food Chem Toxicol 1999;37:313–318.
- 73. Akagi K, Hirose M, Hoshiya T, Mizoguchi Y, Ito N, Shirai T. Modulating effects of ellagic acid, vanillin and quercetin in a rat medium term multi-organ carcingoenesis model. Cancer Lett 1995;94:113–121.
- 74. Siess MH, Le Bon AM, Canivenc-Lavier MC, Suschetet M. Mechanisms involved in the chemoprevention of flavonoids. Biofactors 2000;12:193–199.
- 75. Yang CS, Chung JY, Yang GY, Chhabra SK, Lee MJ. Tea and tea polyphenols in cancer prevention. J Nutr 2000;130:472S-478S.
- 76. Bracke M, Vyncke B, Opdenakker G, Foidart JM, De Pestel G, Mareel M. Effect of catechins and citrus flavonoids on invasion in vitro. Clin Exp Metastasis 1991;9:13–25.
- 77. Sato T, Koike L, Miyata Y, Hirata M, Mimaki Y, Sashida Y, Yano M, Ito A. Inhibition of activator protein-1 binding activity and phosphatidylinositol 3-kinase pathway by nobiletin, a polymethoxy flavonoid, results in augmentation of tissue inhibitor of metalloproteinases-1 production and suppression of production of matrix metalloproteinases-1 and -9 in human fibrosarcoma HT-1080 cells. Cancer Res 2002;62:1025–1029.
- Tatsuta A, Iishi H, Baba M, Yano H, Murata K, Mukai M, Akedo H. Suppression by apigenin of peritoneal metastasis of intestinal adenocarcinomas induced by azoxymethane in Wistar rats. Clin Exp Metastasis 2000;18:657–662.
- 79. Weiss RB, Greene RF, Knight RD, Collins JM, Pelosi JJ, Sulkes A, Curt GA. Phase I and clinical pharmacology study of intravenous flavone acetic acid (NSC 347512). Cancer Res 1988;48:5878–5882.
- Ferry DR, Smith A, Malkhandi J, Fyfe DW, deTakats PG, Anderson D, Baker J, Kerr DJ. Phase I clinical trial of the flavonoid quercetin: Pharmacokinetics and evidence for in vivo tyrosine kinase inhibition. Clin Cancer Res 1996;2:659–668.
- Mulholland PJ, Ferry DR, Anderson D, Hussain SA, Young AM, Cook JE, Hodgkin E, Seymour LW, Kerr DJ. Pre-clinical and clinical study of QC12, a water-soluble, pro-drug of quercetin. Ann Oncol 2001;12:245–248.
- Senderowicz AM. Flavopiridol: The first cyclin-dependent kinase inhibitor in human clinical trials. Invest New Drugs 1999;17:313–320.
- 83. Senderowicz AM. Development of cyclin-dependent kinase modulators as novel therapeutic approaches for hematological malignancies. Leukemia 2001;15:1–9.
- 84. Wang HK. Flavopiridol. National Cancer Institute. Curr Opin Investig Drugs 2001;2:1149–1155.
- Wattenberg LW. Inhibition of carcinogenesis by minor dietary constituents. Cancer Res 1992;52:2085s-2091s.
- Calomme M, Pieters L, Vlietinck A, Vanden Berghe D. Inhibition of bacterial mutagenesis by Citrus flavonoids. Planta Med 1996;62:222–226.
- Dashwood RH, Xu M, Hernaez JF, Hasaniya N, Youn K, Razzuk A. Cancer chemopreventive mechanisms of tea against heterocyclic amine mutagens from cooked meat. Proc Soc Exp Biol Med 1999;220:239– 243.
- Williamson G, Faulkner K, Plumb GW. Glucosinolates and phenolics as antioxidants from plant foods. Eur J Cancer Prev 1998;7:17–21.

- 532 REN ET AL.
 - Carroll KK, Guthrie N, So FV, Chambers AF. Anticancer properties of flavonoids, with emphasis on citrus flavonoids. In: Rice-Evans CA, Packer L, editors. Flavonoids in health and disease. New York: Marcel Dekker Inc.; 1998. p 437–446.
- Lahiri-Chatterjee M, Katiyar SK, Mohan RR, Agarwal R. A flavonoid antioxidant, silymarin, affords exceptionally high protection against tumor promotion in the SENCAR mouse skin tumorigenesis model. Cancer Res 1999;59:622–632.
- Tsyrlov IB, Mikhailenko VM, Gelboin HV. Isozyme- and species-specific susceptibility of cDNAexpressed CYP1A P-450s to different flavonoids. Biochim Biophys Acta 1994;1205:325–335.
- 92. Bu-Abbas A, Clifford MN, Walker R, Ioannides C. Contribution of caffeine and flavanols in the induction of hepatic Phase II activities by green tea. Food Chem Toxicol 1998;36:617–621.
- Sun XY, Plouzek CA, Henry JP, Wang TT, Phang JM. Increased UDP-glucuronosyltransferase activity and decreased prostate specific antigen production by biochanin A in prostate cancer cells. Cancer Res 1998;58:2379–2384.
- 94. Brueggemeier RW. Aromatase, aromatase inhibitors, and breast cancer. Am J Ther 2001;8:333–344.
- 95. Pouget C, Fagnere C, Basly JP, Besson AE, Champavier Y, Habrioux G, Chulia AJ. Synthesis and aromatase inhibitory activity of flavanones. Pharm Res 2002;19:286–291.
- Jeong HJ, Shin YG, Kim IH, Pezzuto JM. Inhibition of aromatase activity by flavonoids. Arch Pharm Res 1999;22:309–312.
- Wang C, Makela T, Hase T, Adlercreutz H, Kurzer MS. Lignans and flavonoids inhibit aromatase enzyme in human preadipocytes. J Steroid Biochem Mol Biol 1994;50:205–212.
- Chang WS, Lee YJ, Lu FJ, Chiang HC. Inhibitory effects of flavonoids on xanthine oxidase. Anticancer Res 1993;13:2165–2170.
- 99. Chan WS, Wen PC, Chiang HC. Structure-activity relationship of caffeic acid analogues on xanthine oxidase inhibition. Anticancer Res 1995;15:703–707.
- 100. Mutoh M, Takahashi M, Fukuda K, Komatsu H, Enya T, Matsushima-Hibiya Y, Mutoh H, Sugimura T, Wakabayashi K. Suppression by flavonoids of cyclooxygenase-2 promoter-dependent transcriptional activity in colon cancer cells: Structure-activity relationship. Jpn J Cancer Res 2000;91:686–691.
- 101. Markovits J, Linassier C, Fosse P, Couprie J, Pierre J, Jacquemin-Sablon A, Saucier JM, Le Pecq JB, Larsen AK. Inhibitory effects of the tyrosine kinase inhibitor genistein on mammalian DNA topoisomerase II. Cancer Res 1989;49:5111–5117.
- 102. Lin JK, Chen YC, Huang YT, Lin-Shiau SY. Suppression of protein kinase C and nuclear oncogene expression as possible molecular mechanisms of cancer chemoprevention by apigenin and curcumin. J Cell Biochem Suppl 1997;28–29:39–48.
- 103. Weber G, Shen F, Prajda N, Yang H, Li W, Yeh A, Csokay B, Olah E, Look KY. Regulation of the signal transduction program by drugs. Adv Enzyme Regul 1997;37:35–55.
- 104. Zi X, Feyes DK, Agarwal R. Anticarcinogenic effect of a flavonoid antioxidant, silymarin, in human breast cancer cells MDA-MB 468: Induction of G1 arrest through an increase in Cip1/p21 concomitant with a decrease in kinase activity of cyclin-dependent kinases and associated cyclins. Clin Cancer Res 1998;4:1055–1064.
- 105. Choi JA, Kim JY, Lee JY, Kang CM, Kwon HJ, Yoo YD, Kim TW, Lee YS, Lee SJ. Induction of cell cycle arrest and apoptosis in human breast cancer cells by quercetin. Int J Oncol 2001;19:837–844.
- 106. Casagrande F, Darbon JM. Effects of structurally related flavonoids on cell cycle progression of human melanoma cells: Regulation of cyclin-dependent kinases CDK2 and CDK1. Biochem Pharmacol 2001;61:1205–1215.
- 107. Bailly C. Topoisomerase I poisons and suppressors as anticancer drugs. Curr Med Chem 2000;7:39–58.
- Sukardiman, Darwanto A, Tanjung M, Darmadi MO. Cytotoxic mechanism of flavonoid from Temu Kunci (Kaempferia pandurata) in cell culture of human mammary carcinoma. Clin Hemorheol Microcirc 2000;23:185–190.
- 109. Lee WR, Shen SC, Lin HY, Hou WC, Yang LL, Chen YC. Wogonin and fisetin induce apoptosis in human promyeloleukemic cells, accompanied by a decrease of reactive oxygen species, and activation of caspase 3 and Ca(2 +)-dependent endonuclease. Biochem Pharmacol 2002;63:225–236.
- 110. Rong Y, Yang EB, Zhang K, Mack P. Quercetin-induced apoptosis in the monoblastoid cell line U937 in vitro and the regulation of heat shock proteins expression. Anticancer Res 2000;20:4339–4345.
- 111. Konig A, Schwartz GK, Mohammad RM, Al-Katib A, Gabrilove JL. The novel cyclin-dependent kinase inhibitor flavopiridol downregulates Bcl-2 and induces growth arrest and apoptosis in chronic B-cell leukemia lines. Blood 1997;90:4307–4312.
- 112. Ren W, Qiao Z, Wang H, Zhu L, Zhang L, Lu Y, Cui Y, Zhang Z, Wang Z. Tartary buckwheat flavonoid activates caspase 3 and induces HL-60 cell apoptosis. Methods Find Exp Clin Pharmacol 2001;23:427– 432.

- 113. Kim SY, Gao JJ, Kang HK. Two flavonoids from the leaves of Morus alba induce differentiation of the human promyelocytic leukemia (HL-60) cell line. Biol Pharm Bull 2000;23:451–455.
- 114. Mata-Greenwood E, Ito A, Westenburg H, Cui B, Mehta RG, Kinghorn AD, Pezzuto JM. Discovery of novel inducers of cellular differentiation using HL-60 promyelocytic cells. Anticancer Res 2001;21:1763–1770.
- 115. Takahashi T, Kobori M, Shinmoto H, Tsushida T. Structure-activity relationships of flavonoids and the induction of granulocytic- or monocytic-differentiation in HL60 human myeloid leukemia cells. Biosci Biotechnol Biochem 1998;62:2199–2204.
- 116. Jing Y, Nakaya K, Han R. Differentiation of promyelocytic leukemia cells HL-60 induced by daidzein in vitro and in vivo. Anticancer Res 1993;13:1049–1054.
- 117. Honma Y, Okabe-Kado J, Kasukabe T, Hozumi M, Umezawa K. Inhibition of abl oncogene tyrosine kinase induces erythroid differentiation of human myelogenous leukemia K562 cells. Jpn J Cancer Res 1990;81: 1132–1136.
- Constantinou A, Kiguchi K, Huberman E. Induction of differentiation and DNA strand breakage in human HL-60 and K-562 leukemia cells by genistein. Cancer Res 1990;50:2618–2624.
- 119. Kandaswami C, Middleton E, Jr. Free radical scavenging and antioxidant activity of plant flavonoids. Adv Exp Med Biol 1994;366:351–376.
- Halliwell B, Gutteridge JM. Role of free radicals and catalytic metal ions in human disease: An overview. Methods Enzymol 1990;186:1–85.
- 121. Halliwell B, Gutteridge JMC, Cross CE. Free radicals, antioxidants, and human disease: Where are we now? J Lab Clin Med 1992;119:598–620.
- 122. Duthie SJ, Dobson VL. Dietary flavonoids protect human colonocyte DNA from oxidative attack in vitro. Eur J Nutr 1999;38:28–34.
- 123. Wang W, Goodman MT. Antioxidant property of dietary phenolic agents in a human LDL-oxidation ex vivo model: Interaction of protein binding activity. Nutr Res 1999;19:191–202.
- 124. Paper DH. Natural products as angiogenesis inhibitors. Planta Med 1998;64:686-695.
- 125. Tosetti F, Ferrari N, De Flora S, Albini A. Angioprevention': Angiogenesis is a common and key target for cancer chemopreventive agents. FASEB J 2002;16:2–14.
- 126. Fotsis T, Pepper MS, Montesano R, Aktas E, Breit S, Schweigerer L, Rasku S, Wahala K, Adlercreutz H. Phytoestrogens and inhibition of angiogenesis. Baillieres Clin Endocrinol Metab 1998;12:649–666.
- 127. Kioka M, Hosokawa N, Komano T, Hirayoshi K, Nagata K, Ueda K. Quercetin, a bioflavonoid, inhibits the increase of human multidrug resistance gene (MDR1) expression caused by arsenite. FEBS Lett 1992;301: 307–309.
- 128. Di Pietro A, Dayan G, Conseil G, Steinfels E, Krell T, Trompier D, Baubichon-Cortay H, Jault J. P-glycoprotein-mediated resistance to chemotherapy in cancer cells: Using recombinant cytosolic domains to establish structure-function relationships. Braz J Med Biol Res 1999;32:925–939.
- 129. Shapiro AB, Ling V. Effect of quercetin on Hoechst 33342 transport by purified and reconstituted P-glycoprotein. Biochem Pharmacol 1997;53:587–596.
- 130. Di PA, Conseil G, Perez-Victoria JM, Dayan G, Baubichon-Cortaya H, Trompiera D, Steinfels E, Jault JM, de WH, Maitrejean M, Comte G, Boumendjel A, Mariotte AM, Dumontet C, McIntosh DB, Goffeau A, Castanys S, Gamarro F, Barron D. Modulation by flavonoids of cell multidrug resistance mediated by P-glycoprotein and related ABC transporters. Cell Mol Life Sci 2002;59:307–322.
- 131. Blagosklonny MV. Treatment with inhibitors of caspases, that are substrates of drug transporters, selectively permits chemotherapy-induced apoptosis in multidrug-resistant cells but protects normal cells. Leukemia 2001;15:936–941.
- 132. Sadzuka Y, Sugiyama T, Sonobe T. Efficacies of tea components on doxorubicin induced antitumor activity and reversal of multidrug resistance. Toxicol Lett 2000;114:155–162.
- 133. Choi SU, Ryu SY, Yoon SK, Jung NP, Park SH, Kim KH, Choi EJ, Lee CO. Effects of flavonoids on the growth and cell cycle of cancer cells. Anticancer Res 1999;19:5229–5233.

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