

Biological activities and chemistry of saponins from *Panax ginseng* C. A. Meyer

Jong Dae Park^{1,*}, Dong Kwon Rhee² & You Hui Lee¹

¹Division of Ginseng Research, KT&G Central Research Institute, Taejeon, Korea; ²College of Pharmacy, Sung Kyun Kwan University, Suwon, Korea; *Author for correspondence (Tel.: +82-42-866-5534; Fax : +82-42-861-1949; E-mail: jdpark@ktng.com)

Key words: active constituents, anticancer, antidiabetic, antihypertensive activities, Araliaceae, biological activities, ginsenosides, metabolism, multidrug resistance, *Panax ginseng*, saponin

Abstract

The roots of *Panax ginseng* C. A. Meyer, known as Korean ginseng have been a valuable and important folk medicine in the East Asian countries, such as China, Korea and Japan for about 2000 years. *Panax* is derived from a word “panacea”, which means cure-all diseases and longevity as well as physical strength and resistance. As the use of traditional Chinese herbs as a food supply becomes more and more popular in the western countries, sales of *Panax ginseng* are increasing in North America and Europe as well as other parts of the world. Active constituents found in most ginseng species include ginsenosides, polysaccharides, peptides, polyacetylenic alcohols and fatty acids. Major active components in *Panax ginseng* are the ginsenosides, a group of saponins with triterpenoid dammarane structure. More than 30 ginsenosides have been isolated, and known compounds are identified but new compounds were elucidated. Pharmacological effects of ginseng have been demonstrated in cancer, diabetes mellitus, cardiovascular system, immune system and central nervous system including anti-stress and anti-oxidant activity. We have focused this review on the effect of ginseng on diabetes, anticancer activity and cardiovascular system and chemical structures of ginsenosides. In addition, our recent biological study on 20(S)-ginsenoside Rg₃ is also touched upon as follows. Multidrug resistance (MDR) has been a major problem in cancer chemotherapy. In this study *in vitro* and *in vivo* modulations of MDR by 20(S)-ginsenoside Rg₃ (Rg₃), a saponin characteristic of red ginseng, was investigated. In flow cytometric analysis using rhodamine 123 as an artificial substrate, Rg₃ promoted accumulation of rhodamine 123 in drug-resistant human fibrocarcinoma KBV 20C cells in a dose-dependent manner, but it had no effect on parental KB cells. Additionally Rg₃ inhibited [³H]-vinblastine efflux and reversed MDR to DOX (doxorubicin), COL, VCR (vincristine) and VP-16 in KBV20C cells. Reverse transcriptase-polymerase chain reaction and immuno-blot analysis after exposure of KBV20C cells to Rg₃ showed that inhibition of drug efflux by Rg₃ was due to neither repression of *MDR1* gene expression nor P-glycoprotein (Pgp) level. Photo-affinity labeling study with [³H]-azidopine, however, revealed that Rg₃ competed with [³H]-azidopine for binding to the Pgp demonstrating that G-Rg₃ competed with anticancer drug for binding to Pgp thereby blocking drug efflux. Furthermore, Rg₃ increased life span in mice implanted with DOX-resistant murine leukemia P388 cells *in vivo* and inhibited body weight increase significantly. Further clinical trial of Rg₃ in reversal of Pgp-associated MDR is highly feasible.

Introduction

The root of *Panax ginseng* is one of traditional and folk medicines which has been used for many therapeutic purposes in the oriental countries such as Korea, China and Japan for thousands of years. The source plant of ginseng is *Panax ginseng* C. A. Meyer (Araliaceae), a herb with fleshy roots which grows wild in cool and shady forests extending from Korea and North eastern China to far eastern Siberia. However, because wild ginseng is relatively rare and very expensive, it has been cultivated in Korea, China and Japan. Most of commercially available ginseng is the root of *Panax ginseng* cultivated in Korea, the northeast district of China and Japan. The medical effects described in the related reference books categorize ginseng as a "mild" medication, a tonic that strengthens, invigorates a weakened body. The mainly physical ailments for which ginseng is said to be effective include headache, fatigue, dizziness, nausea, asthma, hemorrhage and impotence. It is further said to generally strengthen the viscera, improve resistance to external disease-causing agents, and improve the general physical conditions and mental capacity. Modern scientific data is substantiating many of these claims. Eminent scholars from around the world have focused their research their research into a question: which substances in ginseng manifest such miraculous effects? The research has made much progress, but still insufficient to fully uncover the mystery of ginseng. In a noteworthy development in the 1960s, Sibata group identified the structures of saponin and called them as ginsenosides (Shibata et al., 1963a, b). Latest surveys show that saponins account for about 3–4 % of Korean ginseng, and more than 30 kinds of ginsenosides have been found in it, double the number in ginseng of other countries (Park, 1996). Considering that each of these ginsenosides has different pharmacological activities, it becomes apparent that Korean ginseng might have a pharmacological effectiveness superior to those of any other. In this paper, we review recent studies on the effects of ginsenosides on various diseases, particularly on diabetes, hypertension and cancer. Ginseng improves glucose homeostasis and insulin sensitivity. It exerts cytotoxic and anti-metastatic activities against various kinds of cancer cell lines, and induces differentiation or apoptosis of several cancer cells.

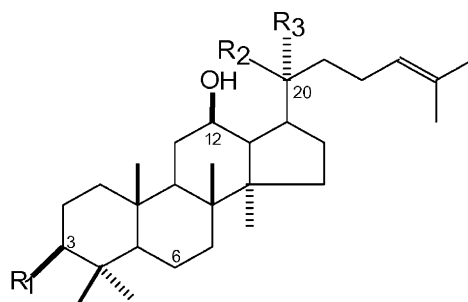
Furthermore, ginsenoside has been reported to have an anti-neoplastic effect by enhancing immune function. An anti-hypertensive effect of ginsenoside is shown to occur by the enhanced synthesis and release of nitric oxide. In addition, ginsenoside exerts anti-atherosclerotic and anti-platelet effects. This review provides useful information on the pharmacological and clinical usage, and also suggests a method to conduct further study on ginseng to identify its potential effects.

Structural property of ginsenoside

The history of scientific research on ginseng dates back to 1854 when Garriques, an American scientist, isolated a saponin from ginseng. However, the actual introduction of ginseng to the West happened after World War II. The chemical structures of several ginseng saponins were characterized in the late 1960s and further accelerated scientific research on ginseng. Saponin components are triterpenoidal dammarane glycosides, named ginsenosides Rx according to their mobility on TLC plates, with polarity decreasing from index "a" to "h". Ginsenosides, known as saponins, are the major components of ginseng (Figures 1, 2, 3 and 4). Ginsenosides possess dammarane triterpenoidal skeleton with a modified side chain at C-20 (Shibata et al., 1995; Huang, 1999). They differ from one another by the type of sugar moieties, their number and their site of attachment. Among the saponins, the genuine sapogenins, protopanaxadiol and protopanaxatriol have been identified as dammar-24-ene-3 β , 12 β , 20(S)-triol and dammar-24-ene-3 β , 6 α 12 β , 20(S)-tetrol, respectively (Shibata et al., 2001).

Ginsenosides of red and white ginsengs

The root of *Panax ginseng* is steamed and dried to prepare red ginseng, while the peeled roots dried without steaming are designated as white ginseng. The commercially available ginseng roots are classified into two forms, red and white ginsengs. It was reported that all of the saponins found in white ginseng were isolated in similar yields from red ginseng, while some partly deglycosylated saponins such as ginsenosides Rh₁, Rh₂ and Rg₃ are obtained from red ginseng as artifacts produced



	R ₁	R ₂	R ₃
20(S)-protopanaxadiol	OH	OH	CH ₃
Ginsenoside Rb ₁	O-glc(2→1)glc	O-glc(6→1)glc	CH ₃
Ginsenoside Rb ₂	O-glc(2→1)glc	O-glc(6→1)arap	CH ₃
Ginsenoside Rc	O-glc(2→1)glc	O-glc(6→1)araf	CH ₃
Ginsenoside Rd	O-glc(2→1)glc	O-glc	CH ₃
20(S)-Ginsenoside Rg ₃	O-glc(2→1)glc	OH	CH ₃
20(R)-Ginsenoside Rg ₃	O-glc(2→1)glc	CH ₃	OH
Ginsenoside Rs ₃	O-glc(2→1)glc(6)Ac	OH	CH ₃
Compound K	OH	O-glc	CH ₃
Ginsenoside Rh ₂	O-glc	OH	CH ₃

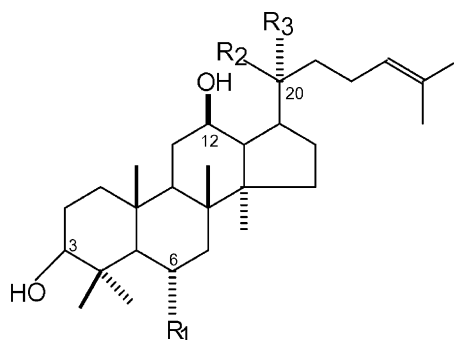
glc : β -D-glucopyranosyl arap : α -L-arabinopyranosyl

araf : α -L-arabinofuranosyl Ac : acetyl

Figure 1. Chemical structures of protopanaxadiol ginsenosides.

during steaming (Kitagawa et al., 1983). In addition, two minor saponins were also isolated only from red ginseng, two being designated as ginsenosides Rs₁ and Rs₂ (Kasai et al., 1983). Stepwise deglycosylated compounds such as compound K and 20(S)-protopanaxatriol can be generated through metabolic transformation by human intestinal bacteria (Hasegawa et al., 1996). Ginsenoside Rg₂ is converted into 20(S)-protopanaxatriol via ginsenoside Rh₁. The binding of the sugar has been shown to have influence on biological activity. Rh₁ and Rh₂ are structurally similar, but have different activity. Ginsenoside Rh₂ decreased growth of B16-BL6 melanoma cells, and stimulated melanogenesis and cell-to-cell adhesiveness. However, Rh₁ had no effect on cell growth and cell-to-cell adhesiveness, but stimulated melanogenesis (Odashima et al., 1985). Although both

Rh₂ and Rh₃ induced differentiation of promyelocytic leukemia HL-60 cells into morphological and functional granulocytes, the potency of Rh₂ was higher (Kim et al., 1998). Another factor that contributes to structural difference between ginsenosides is stereochemistry at C-20. Although both 20(S)- and 20(R)-ginsenoside Rg₂ inhibit acetylcholine-evoked secretion of catecholamines from cultured bovine adrenal chromaffin cells, the 20(S) isomer shows a greater inhibitory effect (Kudo et al., 1998). For new dammarane glycosides named ginsenosides Rg₅, Rh₄, Rs₃ and Rf₂ have been recently isolated from Korean red ginseng and their chemical structures have been elucidated by chemical and spectroscopic methods, as 3-O- $[\beta$ -D-glucopyranosyl (1 → 2)- β -D-glucopyranosyl]dammar-20 (22), 24-diene-3 β ,12 β -diol (ginsenoside Rg₅), 6-O- β -D-glucopyranosyl-dammar-20(22), 24-diene-3 β ,

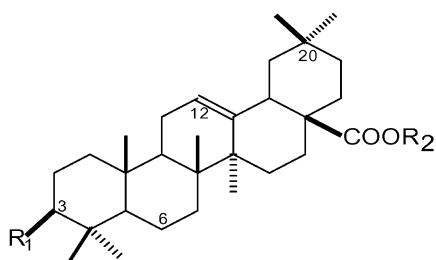


	R ₁	R ₂	R ₃
20(S)-protopanaxatriol	OH	OH	CH ₃
Ginsenoside Re	O-glc(2→1)rha	O-glc	CH ₃
Ginsenoside Rg ₁	O-glc	O-glc	CH ₃
20(S)-Ginsenoside Rg ₂	O-glc(2→1)rha	OH	CH ₃
20(R)-Ginsenoside Rg ₂	O-glc(2→1)rha	CH ₃	OH
20(S)-Ginsenoside Rh ₁	O-glc	OH	CH ₃

glc :β-D-glucopyranosyl rha : α-L-rhamnopyranosyl

Figure 2. Chemical structures of protopanaxatriol ginsenosides.

6α,12β-triol (ginsenoside Rh₄), 3-O-[6''-O-acetyl-β-D-glucopyranosyl(1 → 2)-β-D-glucopyranosyl] 20(S)-protopanaxadiol(ginsenoside Rs₃) and 6-O-[α-L-rhamnopyranosyl(1 → 2)-β-D-glucopyranosyl]dammarane-3β,6α,12β,20(R),25-pentol (ginsenoside Rf₂) (Park et al., 1998) (Figures 1 and 4).



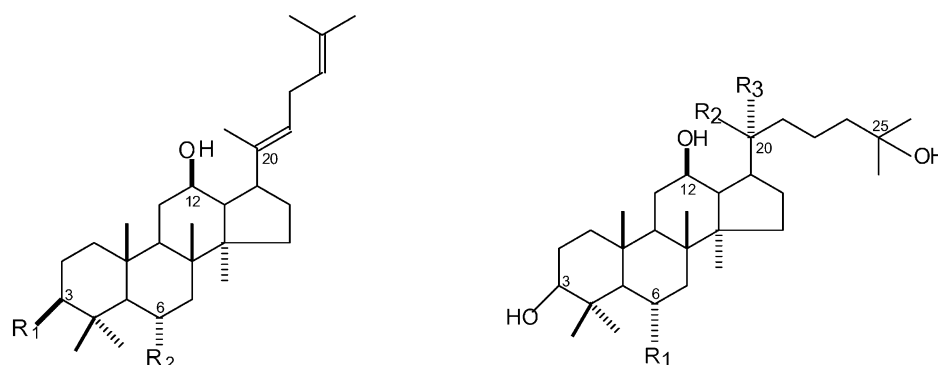
	R ₁	R ₂
Ginsenoside Ro	O-glcUA(2→1)rha	O-glc

glcUA : β-D-glucuronic acid rha : α-L-rhamnopyranosyl

Figure 3. Chemical structures of oleanane ginsenoside.

Metabolism of ginseng saponins by human intestinal bacteria

Crude drugs in herbal prescriptions contain many β-glycosides. Orally administered β-glycosides must meet gastric juice, digestive and bacterial enzymes in the gastrointestinal tract. Therefore, plant β-glycosides may act as natural prodrugs (Kobashi, 1998), which can be transformed to active metabolites after oral administration and finally induce biological activities. The decomposed products taken from the stomach of rats given a gastric bolus of ginsenoside Rb₂ differed from those produced by 0.1 N HCl (Han et al., 1982), and Rb₂ was found to be hardly decomposed by gastric juice with the exception of slight oxygenation (Karikura et al., 1991a, b). Certain ginsenosides such as Rb₁ and Rg₁ are poorly absorbed after injection (Odani et al., 1983). The metabolism of ginsenosides by human intestinal bacteria was examined. The oligosaccharides connected to the C-3 or C-20 hydroxy group of the aglycone are cleaved stepwise from the terminal sugar by bacterial hydrolysis. The



	R ₁	R ₂		R ₁	R ₂	R ₃
Ginsenoside Rh ₃	O-glc	H	Ginsenoside Rf ₂	O-glc(2→1)rha	CH ₃	OH
Ginsenoside Rh ₄	OH	O-glc				
Ginsenoside Rg ₅	O-glc(2→1)glc	H				

glc :β-D-glucopyranosyl
rha :α-L-rhamnopyranosyl

Figure 4. Chemical structures of ginsenosides with modified side chain.

main metabolic pathways are supposed as shown in Figure 5. Protopanaxadiol type saponins (ginsenosides Rb₁, Rb₂, Rc and Rd) were hydrolyzed to compound K(C-K) by intestinal flora (Karikura et al., 1991a, b) and the bacteria capable of hydrolyzing Rb₁ to C-K was identified as *Prevotella oris*. Whereas, protopanaxatriol type saponins (ginsenosides Re and Rg₁) were hydrolyzed to 20(S)-protopanaxatriol. C-K was shown to increase the cytotoxicity of antineoplastic drugs (Hasegawa et al., 1995) and to induce apoptosis in B16-BL6 melanoma cells (Wakabayashi et al., 1998). And also, the antiproliferative activity of C-K against tumor cells is primarily due to the induction of apoptosis *via* promotion of caspase-3 activity and regulation of apoptosis related proteins, which may lead to the anti-tumor activity (Suda et al., 2000). Moreover, in parallel with the antimetastatic effect of oral administration of C-K after the i.v. injection of colon 26-L5 cells, administration of C-K before tumor inoculation exerted a dose-dependent inhibition of liver metastasis (Hasegawa et al., 2000a). The pharmacokinetics of C-K and protopanaxatriol were investigated. C-K was selectively accumulated into the liver and mostly excreted as bile, but about 24% of C-K was

esterified with fatty acids in the liver and accumulated in the liver longer than C-K. In case of protopanaxatriol, it was absorbed from the small intestine into the mesenteric lymphatics followed by the esterification with fatty acids and its spreading to the organ and excretion as bile (Hasegawa et al., 2000b). The fatty acid esterification of intestinal bacterial metabolites of ginsenosides was found to potentiate the antitumor activity of the parental metabolites through delay of the clearance of C-K from the liver and immunostimulation of lymphocyte-mediated tumor cytotoxicity by the esterified form (Hasegawa et al., 2000c, 2002). These results definitely indicate that the pharmacological studies accompanied by metabolic elucidation of ginsenosides and other biologically active compounds lead to a better understanding of the real active ingredients in the body.

Reduction of blood glucose level and improvement of diabetes treatment by ginsenosides

Diabetes, affecting almost 3% of the world population, is one of the major global health problems and there is especially high number of incidence in

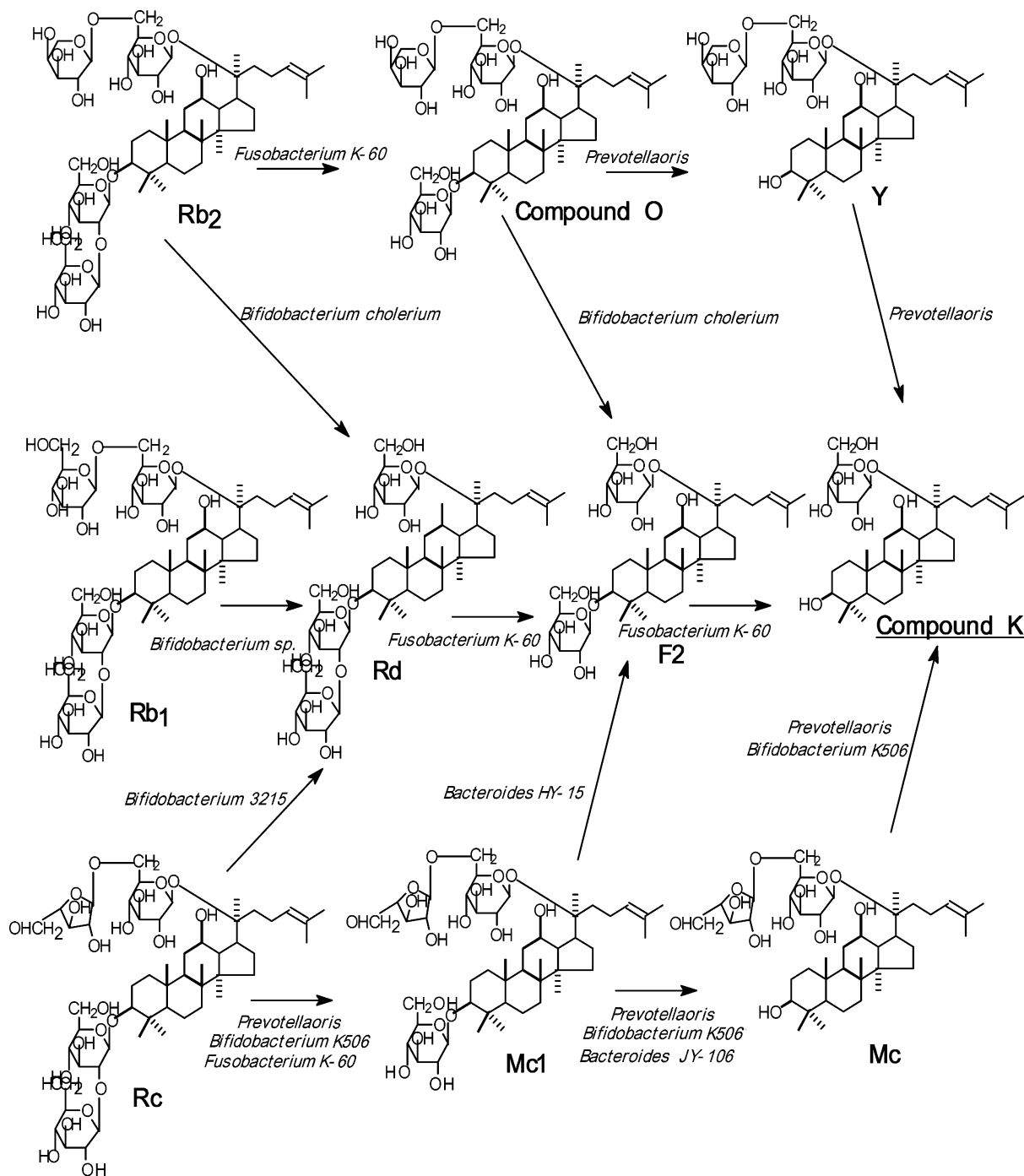


Figure 5. Metabolic pathway of ginsenosides Rb₁, Rb₂, Rc and Rd to compound K by intestinal bacterial flora after oral administration.

elderly population. It has been shown that the root of *Panax ginseng* and other ginseng species possess anti-hyperglycemic activity *in vitro* (Kimura et al.,

1980, 1981a, b) and *in vivo* (Kimura et al., 1981, 1981a, b, 1999; Yokozawa et al., 1985). More than 90% of patients with diabetes have type 2 diabetes,

which is related to aging and diet. Although type 2 diabetes is more common and has serious complications, even reducing life expectancy by 8–10 years (Astrup and Finer, 2000), most *in vivo* animal studies using ginseng have been conducted in type 1 rather than type 2 diabetes models. Here, we would like to focus on the effects of ginseng on type 2 diabetes.

The root of *Panax ginseng* has been used to improve glucose homeostasis and insulin sensitivity (Sonnenborn and Proppert, 1990) and further clinically to treat type 2 diabetes (Bensky et al., 1993; Huang, 1999). It has been observed that blood glucose level falls significantly in genetically obese diabetic mice after treatment with a single 90 mg/kg of ginseng root extract at an intraperitoneal (i.p.) dose (Kimura et al., 1999). It has also been demonstrated that 3 g of American ginseng root given 40 min before the test meal significantly lowers blood glucose level in non-diabetic subjects and type 2 diabetic patients (Vuksan et al., 2000). Oral administration of *Panax ginseng* root to diabetic KKAY mice for 4 weeks reduced blood glucose levels similar to that of an insulin sensitizer (rosiglitazone)-treated group (Chung et al., 2001). Moreover, ginseng therapy for type 2 diabetes elevates mood, improves psychophysical performance, and reduces fasting blood glucose and body weight. A 200 mg dose of ginseng improves glycated hemoglobin, serum lipid, amino-terminal propeptide concentration, and physical activity. These observations suggest that ginseng is beneficial for the people with type 2 diabetes and to prevent development of diabetes in non-diabetic subjects.

The main component of *Panax ginseng* is ginsenosides (ginseng saponins). Ginsenoside Rb₂ was found to be the most effective component of ginsenosides for streptozotocin-diabetic rats (Yokozawa et al., 1985). Rats treated with ginsenoside Rb₂ showed a significant decrease in blood glucose level with increased activity of glucokinase and decreased activity of glucose 6-phosphatase. Recently, anti-hyperglycemic and antiobese effects of *Panax ginseng* berry extract have been demonstrated, and its major constituent, ginsenoside Re has been observed (Attele et al., 2002). Treatment of the berry extract by daily i.p. injection for 12 days in obese diabetic C56BL/6J mice reduced glucose to levels similar to normal control value; after treatment the mice also has significantly improved glucose tolerance. The improvement of

blood glucose level in the extract-treated *ob/ob* mice is associated with a significant reduction in serum insulin level in fed and fasting mice. A hyperinsulinemic–euglycemic clamp study reveals more than two-fold increase in the rate of insulin-stimulated glucose disposal in treated *ob/ob* mice. In addition, the extract-treated *ob/ob* mice lost a significant amount of weight, which was associated with a significant reduction in food intake and very significant increase in energy expenditure and body temperature.

Anticancer activities of ginsenosides

The main weapons in the war against cancer have been early detection and surgical removal, radiotherapy, chemotherapy, and attempts to develop gene therapy. However, the results are less than ideal, and the strategy is now changing from therapeutic approaches to prevention of cancer by identifying effective natural products as chemopreventive agents. One promising candidate with cancer-preventive effects is *Panax ginseng*. Its usefulness in cancer has been shown by extensive preclinical and epidemiological studies. Here, we describe the anti-carcinogenic effect of ginseng based on its diverse mechanisms.

Effect on tumor cell cytotoxicity and differentiation

Saponin and non-saponin compounds have been reported to show cytotoxic activities against various kinds of cancer cell lines in culture. Major active components are ginsenoside Rh₂, a peculiar component of red ginseng, and also polyacetylenes, panaxydol, panaxynol and panaxytriol. Ginseng was found to have the ability to induce neoplastic cells into normal cells. Ginsenoside Rh₂ inhibited the growth and colony forming ability of Morris hepatoma cells in soft agar suspension culture, and stimulated serum protein synthesis of these cells, thus converting the cell characteristics both functionally and morphologically to those resembling original normal liver cells, a process known as “redifferentiation or reverse transformation” (Odashima et al., 1989). Similarly, ginsenosides Rh₁ and Rh₂ have been shown to cause differentiation of F9 teratocarcinoma stem cells via binding to a steroid receptor (Lee et al., 1993). Therefore, recent studies on therapy for various

types of cancers have focused on drugs, which induce differentiation of maturation-resistant cells causing the disease. Furthermore, ginsenoside Rh₂ has been found to significantly induce B16 cell differentiation and to increase melanin synthesis in B16 cells (Xia and Han, 1996). The cytotoxicity of natural glycosides from ginseng, semisynthetic analogues of the genus *Betula* was evaluated to elucidate structure–activity relationships. Cytotoxic effects of the dammarane glucosides were inversely proportional both to the number of sugars attached to the aglycones and to the number of hydroxyl groups of the aglycones. The type of side chain and the configuration of the hydroxyl group at C-3 in aglycones was not found to have a significant influence on the cytotoxicity (Atopkina et al., 1999). Ginsenosides Rg₃ and Rh₂ has been found to inhibit the proliferation of prostate cancer cells, which may be associated with modulation of three modules of MAP kinases (Kim et al., 2004). In addition, 20(S)-protopanaxadiol and ginsenoside Rh₂ reduce cell proliferation and increase sub-G1 cells in two cultured intestinal cell lines, Int-407 and Caco-2, indicating a specific structure–function relationship for bioactive ginsenosides in two contrasting intestinal cell types (Popovich et al., 2004).

Anti-metastatic effects

Generally, primary tumor is not fatal. Instead, most cancer patients succumb to metastases – multiple, widespread tumor colonies established by malignant cells that detach themselves from the original tumor and travel through the body often to distant sites. Some invading cells penetrate into a body cavity or the blood, lymph or spinal fluid and then are released (Friedberg, 1986). Ginseng saponin has recently received a great deal of attention to the effects to inhibit invasion and metastasis of cancer cells. 20(R)- and 20(S)-ginsenoside Rg₃ was found to possess an ability to inhibit the lung metastasis of tumor cells such as B16-BL-6 melanoma and colon 26M3.1, when they were orally administered at a dose of 100–1000 µg/mouse, and the mechanism of their anti-metastatic effect is supposed to be related to inhibition of the adhesion and invasion of tumor cells, and also to anti-angiogenesis activity (Mochizuki et al., 1995). Glucocorticoid receptor-induced down-regulation of matrix metalloproteinase

(MMP)-9 by panaxadiol (PD) and panaxatriol (PT) appear to be associated with the reduced invasive capacity of HT1080, a highly metastatic human fibrosarcoma cell line (Park et al., 1999). Multiple administration of ginsenoside Rb₂ after the intravenous inoculation of B16-BL6 melanoma cells resulted in a significant inhibition of tumor-associated angiogenesis responsible for the inhibition of lung tumor metastasis (Sato et al., 1994). Ginsenoside Rb₂ has been reported to inhibit invasion to the basement membrane via MMP-2 suppression in some endometrial cancers such as HHUA and HEC-1-A cells, and is supposed to be used as a medicine for inhibition of secondary spreading of uterine endometrial cancers (Fujimoto et al., 2001). It was shown that ginsenoside Rg₃ inhibited experimental pulmonary metastasis by highly metastatic mouse melanoma B16F7 cells, which was mediated by inhibiting the 1-oleoyl-lysophosphatidic acid (LPA)-triggered rise of intracellular Ca⁺² (Shinkai et al., 1996). While these results suggest the usefulness of ginsenoside Rg₃ in preventing cancer spread, further studies on *in vivo* will be necessary to clarify anti-metastatic effects of ginseng.

Anticarcinogenic activities and synergistic effects in combination with chemical therapeutic agents

Several investigations were carried out to evaluate the inhibitory or preventive effects of ginseng on carcinogenesis induced by various chemical carcinogens. The prolonged administration of Korean red ginseng extract inhibited the incidence and the proliferation of tumors induced by 7,12-dimethylbenz(a)anthracene (DMBA), urethane, and aflatoxin B1 (Yun et al., 1983). The chemopreventive potential of ginseng was evaluated using DMBA-induced skin tumorigenesis (papillomagenesis) in male Swiss albino mice, and there was a marked reduction not only in tumor incidence but also in cumulative tumor frequency at the initiation phase of tumorigenesis, while a little reduction at the promotional stage, suggesting the anti-carcinogenic activities of ginseng (Kumar, 1993). Ginsenosides Rg₃ and Rg₅ were found to show statistically significant reduction of lung tumor incidence in the newly established 9 weeks' medium-term anti-carcinogenicity test model of lung tumors in mice, and ginsenoside Rh₂ showed a tendency of decreasing the incidence, indicating

them to be active anti-carcinogenic compounds (Yun et al., 2001). Subsequent studies have confirmed that *Panax ginseng* C. A. Meyer cultivated in Korea is a non-organ specific cancer preventive against human cancers and the anticarcinogenicity or human cancer preventive effects is due to ginsenosides Rg₃, Rg₅ and Rh₂ (Yun, 2003). The inhibitory effects of ginseng on the development of 1,2-dimethylhydrazine (DMH)-induced aberrant crypt foci (ACF) in the colon were investigated in rats. Dietary administration of red ginseng in combination with DMH suppresses colon carcinogenesis in rats associated partly with inhibition of cell proliferation, acting on ACF in the colonic mucosa (Fukushima et al., 2001). In addition, anti-carcinogenic effect of red ginseng on the development of liver cancer induced by diethylnitrosamine (DEN) in rats was identified in preventive and curative groups (Wu et al., 2001). More recently, it has been reported that less glycosylated protopanaxadiol derivatives are effective in cancer prevention and some oleanane-type pentacyclic triterpenoid compounds show anti-carcinogenic activities in two-stage anti-cancer promotion experiments *in vitro* and *in vivo* (Shibata, 2001). Ginsenoside Rh₂ has been described to have diverse effects on the expression of the transformed phenotype in BALB/c3T3 cells, and augments the metastatic potential in an experimental metastasis assay (Tatsuka et al., 2001). The estrogenic potential of American ginseng extract to induce the expression of pS2, an estrogen-regulated gene, was evaluated in breast cancer cell lines MCF-7, T-47D and BT-20. It is found that American ginseng exhibits estrogen-like effects on estrogen receptor-positive breast cancer cells by inducing pS2 expression, suggesting that it may play a protective role against breast cancer (Duda et al., 1996). It is further supported by the fact that American ginseng inhibits breast cancer cell growth by transcriptional activation of the p21 gene, a universal cell cycle inhibitor, independent of p53 (Duda et al., 2001). It is also reported that concurrent use of American ginseng extract and breast cancer therapeutic agents results in a significant suppression of cancer cell growth, suggesting its synergistic effects on breast cancer therapeutics (Duda et al., 1999). Compound K, an intestinal bacterial metabolite derived from protopanaxadiol saponin, has been found to show anti-inflammatory effects by inhibiting TPA-in-

duced COX-2 expression, which may contribute to its antitumor-promoting effects on mouse skin carcinogenesis (Lee et al., 2005).

Antitumor activities

It has been reported that an inhibitory effect of oral administration of ginsenoside Rh₂ on tumor growth in nude mice bearing human ovarian cancer cells (HRA), resulting in a remarkable retardation of the tumor growth. In particular, tumor growth in mice treated with 15, 30 and 120 μ M of ginsenoside Rh₂ was significantly inhibited, compared to that in CDDP (*cis*-diaminedichloroplatinum) (II)-treated mice as well as in untreated mice (Tode et al., 1993). Further investigation showed that p.o. but not i.p. treatment with ginsenoside Rh₂ resulted in induction of apoptosis in the tumor in addition to augmentation of the natural killer activity in spleen cells from tumor-bearing nude mice, suggesting that an evaluation of the treatment of recurrent or refractory ovarian tumors is warranted (Nakata et al., 1998).

Induction of apoptosis

Apoptosis is responsible for pathological mechanism related to human diseases such as cancer, autoimmune disease, viral infection and neurodegenerative disorder (Thompson, 1995). Ginsenoside Rh₂ has been shown to arrest cell cycle at the G1 phase and to prolong the S phase (Fujikawayammamoto et al., 1987; Lee et al., 1996). It was reported that ginsenoside Rh₂ induced apoptosis through protein kinase C in human neuroblastoma SK-N-BE and rat glioma C6Bu-1 cells (Kim et al., 1999a–d). In addition, ginsenoside Rh₂ has been shown to induce apoptosis independently of Bcl-2, Bcl-x_L, or Bax in C6Bu-1 cells (Kim et al., 1999a–d). In a parallel study, it was found that ginsenoside Rh₂-induced cell death was mediated by the generated reactive oxygen species and activation of caspase pathway in a Bcl-x_L-independent manner. These reports demonstrate that the induction of apoptosis by ginseng can be one of its anti-carcinogenic mechanisms. In structure and apoptosis relationship, the presence of sugars in protopanaxadiol and protopanaxatriol aglycone structures has been found to reduce the potency to induce apoptosis in the human leukemia (THP-1)

cell line and alternately alter membrane integrity (Popovich et al., 2002).

Inhibitory activities of 20(S)-ginsenoside Rg₃ on multi-drug resistance (MDR)

One of the major side effects to the effective treatment of human malignancies is the acquisition of broad anti-cancer drug resistance by tumor cells. This phenomenon has been termed "multi-drug resistance (MDR)". Therefore, development of new modulator for inhibition of drug resistance is required. MDR inhibitory activity was determined by measuring cytotoxicity to MDR cells using multi-drug resistant human fibrocarcinoma KB V20C, which is resistant to 20 nM of vincristine and expresses high level of *mdr1* gene. 20(S)-Ginsenoside Rg₃ (Rg₃), saponin of red ginseng, was found to have the most potent inhibitory activity on MDR and its ID₅₀ (dose for 50% inhibition) was 8.2×10^{-5} M. In cytotoxicity assay, ginsenoside Rg₃ did not affect the growth of normal cells (Park et al., 1996). More recently, it was shown that the mechanism of Pgp (P-glycoprotein) inhibition and *in vivo* efficacy by Rg₃ has been elucidated as follows by us (Kim et al., 2003).

Inhibition of rhodamine 123 efflux by 20(S)-ginsenoside Rg₃

Rhodamine 123 acts as a good substrate for MDR-associated Pgp and agents that block Pgp have been found to increase the retention of rhodamine 123 in MDR cells. Therefore, drug-sensitive and -resistant cells were exposed to rhodamine 123 at the various concentrations of verapamil or Rg₃ for 30 min and retention of rhodamine 123 was determined. Treatment of KB cells with Rg₃ did not affect retention of rhodamine 123, but treatment of KBV20C cells significantly increased the accumulation of rhodamine 123 in a dose-dependent manner (Figure 6). Rg₃ did not increase efflux of rhodamine 123 effectively at lower doses, however Rg₃ was more potent than verapamil in modulating MDR function at the higher concentrations than 200 μ M of Rg₃, as reflected by higher rhodamine 123 retention in the resistant cells. Since incubation of 320 μ M of Rg₃ for 30 min with KBV20C cells significantly increased efflux of rhodamine 123 without losing

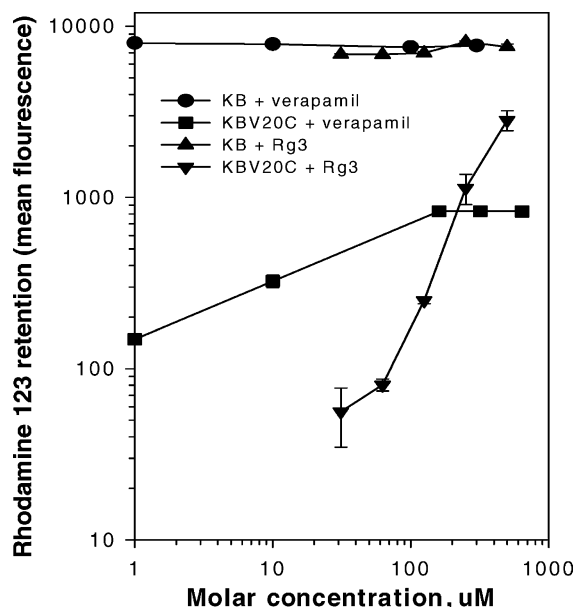


Figure 6. Increase of rhodamine 123 retention by Rg₃. Rhodamine 123 retention in resistant KBV20C but not in parent KB cells. KB or KBV20C cells were incubated at 37 °C for 24 h, and were resuspended in fresh media containing rhodamine 123, and further incubated at 37 °C for 30 min. After washing, the cells were incubated again in fresh media containing Rg₃, or verapamil at the indicated concentrations at 37 °C for 30 min. Cells were then removed and re-suspended in PBS, and analyzed by flow cytometry. The median fluorescence was used as a quantitative measure of intracellular fluorochrome accumulation and hence an indicator for Pgp inhibition. The vertical bar represents mean \pm SD of the two experiments in triplicate. If vertical bars are not apparent, the size of the SD was close to zero. The average fluorescence values in KB and KBV20C without Rg₃ or verapamil were 6777 and 44.5, respectively.

cells integrity in PBS buffer, subsequent time course experiment used 320 μ M of Rg₃. These results clearly demonstrated that Rg₃ reversed MDR in KBV20C cells specifically and effectively in a dose dependent manner.

Inhibition of [³H]-vinblastine efflux by Rg₃

To ascertain further that Rg₃ can enhance the accumulation of chemotherapeutic agents, KB or KBV20C cells were incubated with [³H]-vinblastine for the indicated time in the presence of either verapamil or Rg₃, and the accumulated [³H]-vinblastine in KB or KBV20C cells was subsequently counted. In KB cells, Rg₃ or verapamil did not exhibit any difference from the group without any treatment in [³H]-vinblastine accumulation

(data not shown). In KBV20C cells, 80 μM of Rg_3 had a marginal effect in reversing MDR, however at 320 μM [^3H]-vinblastine accumulation was enhanced by approximately four-fold after 30 min incubation at 37 $^\circ\text{C}$. In comparison, verapamil at 100 μM augmented drug accumulation by two-fold demonstrating that 320 μM of Rg_3 inhibited drug efflux more potently than 100 μM of verapamil (Figure 7). This result confirmed that Rg_3 is an effective MDR modulator. Also potentiation of the accumulation of drug in MDR cells, but not in drug-sensitive cells, indicate that the MDR reversing effects of Rg_3 were not due to the toxicity of Rg_3 itself.

Inhibition of [^3H]-azidopine binding to Pgp by Rg_3

Since Rg_3 did not affect *MDR1* gene expression, MDR reversal by Rg_3 might be due to inhibition of drug binding to Pgp by Rg_3 . To demonstrate

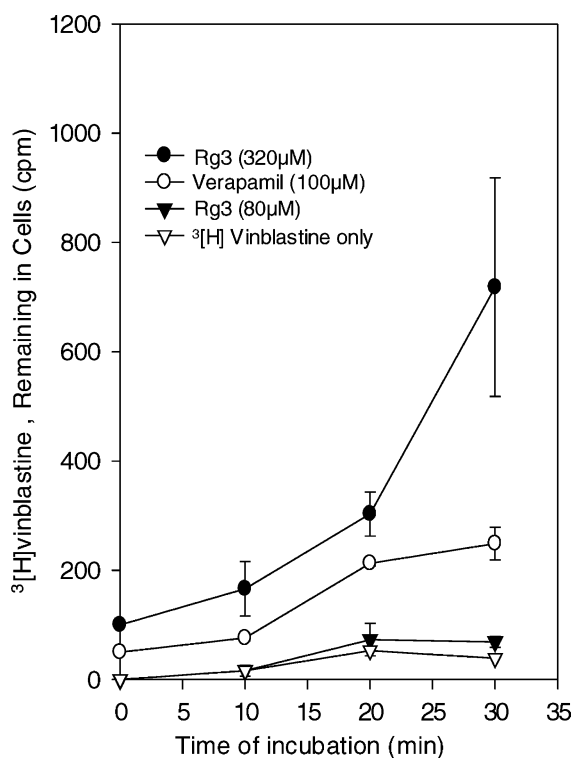


Figure 7. Increase of [^3H]-vinblastine accumulation by Rg_3 . [^3H]-vinblastine accumulation in KBV20C cells was determined after incubation with 26 μM [^3H]-vinblastine and the appropriate modulator for the indicated time. After the indicated time, the cells were washed with ice-cold PBS twice, solubilized in 0.2% Triton X-100 in 10 mM phosphate buffer (pH 7.4), harvested, and then counted. The vertical bar represents mean \pm SD of triplicate determinations.

competitive inhibition of drug binding by Rg_3 , photoaffinity labeling of Pgp with [^3H]-azidopine was used. Treatment of 100 μM of Rg_3 completely inhibited binding of [^3H]-azidopine to Pgp demonstrating that Rg_3 competes with [^3H]-azidopine for binding to Pgp (Figure 8).

In vivo efficacy

For *in vivo* evaluation of Rg_3 , a standard P388 murine leukemia model was used. In a preliminary experiment, DOX (doxorubicin) alone or a combination of a wide range of doses (2.5, 10, 40, 80 mg/kg) of the Rg_3 with 4 mg/kg of DOX was administered to mice (6/group) implanted with P388/DOX tumors. The result demonstrated that administration of a combination of 2.5 mg/kg of Rg_3 and 4 mg/kg of DOX did not result in significant increase in life span of mice or mice weight, and administration of 80 mg/kg of Rg_3 , by itself, did not produce any significant difference in life span compared with the vehicle control. In addition, combination treatment with 4 mg/kg DOX and 80 mg/kg Rg_3 produced no significant increase in life span of mice than combination treatment with 4 mg/kg DOX and 40 mg/kg Rg_3 (data not shown). Therefore mice were treated with a combination of 4 mg/kg DOX plus 10 or 40 mg/kg Rg_3 . When mice were treated with a combination of 4 mg/kg DOX plus 10 mg/kg of the Rg_3 , a significant increase in life-span ($p < 0.01$) was observed. Increasing the dose of Rg_3 upto

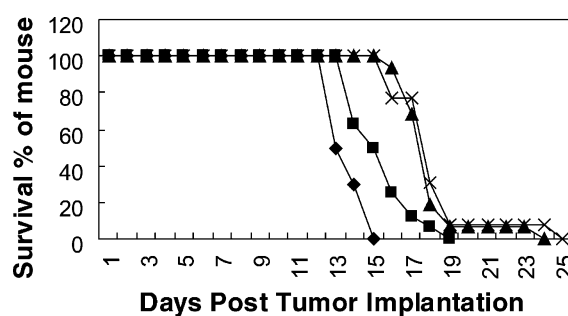


Figure 8. Inhibition of [^3H]-azidopine labeling of P-glycoprotein by Rg_3 . To determine inhibition of [^3H]-azidopine binding to Pgp, resistant KBV20C plasma membrane proteins were incubated with 0.2 μM [^3H]-azidopine in the absence or presence of Rg_3 or verapamil at the indicated concentrations and cross-linked by UV irradiation. Photolabeled membrane proteins were analyzed by SDS-PAGE and visualized by autoradiography.

40 mg/kg did not extend life span any further (Figure 9). These results suggest that the increase of DOX cytotoxicity by Rg₃ *in vivo* is modulated through the interaction of the Rg₃ with Pgp. Further clinical trial in reversal of Pgp-associated MDR is highly feasible.

In addition, it is also reported that quasipanaxatriol, 20(S)-protopanaxatriol, ginsenoside Rh₂ and compound K, metabolic final product of protopanaxadiol saponin, greatly enhanced the cytotoxicity of the anti-cancer drugs in P388/ADM cells (adriamycin-resistant P388 leukemia cells) (Hasegawa et al., 1995). The reversal of daunomycin resistance in P388/ADM by quasipanaxatriol with double bond introduced at C-20 of protopanaxatriol was found to show the effective accumulation of the drugs mediated by the daunomycin-efflux blockage. Furthermore, a recent similar result showed that protopanaxatriol ginsenoside has a chemosensitizing effect on Pgp-mediated MDR cells by increasing the intracellular accumulation of drugs through direct interaction with Pgp at the azidopine site (Choi et al., 2003). These results suggest that further clinical trial of ginseng components in reversal of Pgp-associated MDR is highly feasible.

Antihypertensive effects of ginsenosides

High blood pressure is associated with decreased life expectancy and increased risk of stroke, coro-

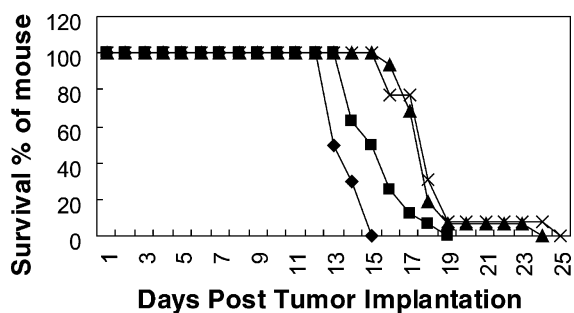


Figure 9. Increase of the survival of mice implanted intraperitoneally into P388/DOX murine leukemia tumor cells by Rg₃ in combination with DOX. Multidrug resistant P388/DOX cells (2×10^6 cells/0.1 ml) were inoculated intraperitoneally into BDF1 female mice. Compounds were administered intraperitoneally at every 3 days after tumor implantation. The results represent the survival percentage versus days after tumor implantation. Control, ◆; DOX 4 mg/kg, ■; DOX 4 mg/kg + Rg₃ 10 mg/kg, ▲; DOX 4 mg/kg + Rg₃ 40 mg/kg, ×.

nary heart disease and other end-organ disease such as renal failure. Ginseng contains active compounds normalizing blood pressure. The effect of a certain drug on the blood pressure can be analyzed by investigating the effect of the drug on the smooth muscle of blood vessel. It is well established that blood vessel smooth muscle tone is regulated by the available intracellular Ca²⁺ concentration, which in turn is profoundly influenced by interaction of the cellular membrane and sarcoplasmic reticulum in the smooth muscle. It is found that both protopanaxatriol and protopanaxadiol saponins inhibit Ca²⁺ binding to the cellular membrane, but protopanaxatriol is approximately 180% more potent than protopanaxadiol ginsenosides (Lee, 1980). It was reported that ginseng induced no significant change in blood pressure in those subjects with normal blood pressure, but had a normalizing effect on the subjects with abnormal blood pressure (Yammamoto, 1992). It has recently been reported that vasodilation and protective effect of ginsenoside Rg₁ against free radical injury might be related to enhanced synthesis and release of NO (Nitric Oxide), demonstrating the usefulness of ginseng in treatment of pulmonary and systemic hypertension (Gillis et al., 1993). The detailed mechanism and evidence of anti-hypertensive effect of ginseng are as follows.

Endothelium-dependent vasorelaxation

Endothelium plays an important role in regulating vascular tone by releasing several vasoactive autacoids including prostacyclin, endothelium-derived relaxing factor (EDRF) and endothelium-derived hyperpolarizing factor (EDHF) (Vane et al., 1990). EDRF has been identified as NO, which is produced from L-arginine by NOS (nitric oxide synthase) (Palmer et al., 1988). NO relaxes blood vessel mostly by stimulating soluble guanylyl cyclase, which leads to an increased production of cGMP in vascular smooth muscle (Rapaport and Murad, 1983). Ginsenosides have been found to lower blood pressure in a dose-dependent manner in rats at doses of 10–100 mg/kg, which is mediated by release of endothelium-derived NO, enhancing the accumulation of cGMP (Kim et al., 1994). Recently, it is also reported that ginseng can improve the vascular endothelial dysfunction in patients with hyper-

tension possibly through increasing synthesis of NO. Protopanaxatriol and its purified ginsenosides Rg₁ and Re caused endothelium-dependent relaxation, which is associated with the formation of cGMP. In contrast, protopanaxadiol or ginsenosides Rg₁ and Re did not affect vascular tone or production of cGMP in rat aorta. Moreover, ginsenosides Rg₁ and Re were less effective endothelium-dependent vasodilators than total ginsenosides and protopanaxatriol (Kang et al., 1995). Ginsenoside Rg₃ is one of protopanaxadiol group of red ginseng, steamed and dried ginseng. Present findings indicate that ginsenoside Rg₃ effectively stimulates the NO formation in endothelial cells, which accounts for the endothelium-dependent relaxation and production of cGMP in the rat aorta. The ginsenoside Rg₃-induced endothelium-dependent relaxation was markedly inhibited by a non-selective K⁺ channel blocker, but not by an ATP-sensitive K⁺ channel blocker (Kim et al., 1999a–d). These findings show that ginsenoside Rg₃ activates tetraethylammonium-sensitive K⁺ channels in endothelial cells, which presumably leads to an influx of Ca²⁺ and the subsequent activation of the endothelial NOS (eNOS) (Kim et al., 1999a–d). In order to investigate the effect of red ginseng on the blood pressure, the change of blood pressure and heart rate after intravenous injection of red ginseng was studied in the conscious normotensive and one-kidney, one-clip Goldblatt hypertensive rats. Experimental evidence indicates that the NO-releasing effect of red ginseng is, like other NO donors, partly contributed to hypotensive effects (Jeon et al., 2000). Clinical study has been performed to estimate the effect of Korean red ginseng on vascular endothelial cell dysfunction in patients with hypertension. To assess the function of the vascular endothelial cell, changes of forearm blood flow to infusion of acetylcholine, sodium nitroprusside and bradykinin in incremental doses were measured by venous occlusion plethysmography. In ginseng-treated hypertensive group, forearm blood flows at the highest dose of acetylcholine and bradykinin were significantly higher than those of the non-treated hypertensive group. It was found that Korean red ginseng could improve the vascular endothelial dysfunction in patients with hypertension possibly through increasing the NO synthesis (Sung et al., 2000). These results support that ginseng shows phar-

macological activities on circulatory diseases including hypertension. In another study, ginsenosides Rb₁ and Re has been found to decrease cardiac contraction in adult rat ventricular myocytes, which may be mediated in part through increased NO production (Scott et al., 2001). Based upon the previously published research data, several reviews underscore the potential benefit effects of ginseng on cardiovascular diseases and emphasizes the necessity for more rigorous systemic investigation (Zhou et al., 2004). When electrophysiological effects of ginsenosides were examined on action potential and membrane currents recorded from isolated guinea pig ventricular myocytes, inhibition of L-type Ca²⁺ current and enhancement of the delayed rectifier K⁺ current appear to be associated with ginsenoside Re (Bai et al., 2003). Effect of notoginsenoside R1, active component of *Panax notoginseng*, was found to increase the fibrinolytic potential in cultured human pulmonary artery smooth muscle cells by increasing the production of tissue-type plasminogen activator, suggesting the effect of notoginsenoside R1 in the treatment of cardiovascular diseases (Zhang et al., 1997).

Antiatherosclerosis and antiplatelet effects

Endothelial cell damage is considered to be the initial step in the genesis of thrombosis and arteriosclerosis, the common precursors of cardiovascular disorders. Platelet hyperfunction such as enhanced platelet aggregation associated with overproduction of thromboxane A₂ (TXA₂), a potent platelet aggregative and vasoconstrictive substance, has been frequently encountered in patients with cardiovascular thrombotic diseases. Prostaglandin I₂ (PGI₂), a potent anti-platelet aggregative and vasodilatory substance produced in vascular walls, is also reported to less synthesize in patients with atherosclerotic changes than normal subjects. Panaxynol was found to markedly inhibit the aggregation of platelets induced by collagen, arachidonic acid and platelet activating factor (PAF), while ginsenosides had no significant effect on the aggregation. It is suggested that panaxynol is the most potent anti-platelet agent in ginseng and its mechanism of action is chiefly due to the inhibition of thromboxane formation (Teng et al., 1989). In addition, it was

reported that panaxynol inhibited the aggregation, release and thromboxane formation in rabbit platelets, while ginsenosides R_o, R_{g₁} and R_{g₂} suppressed only the release (Kubo et al., 1990). Continually, it was also demonstrated that ginsenoside R_{g₁} inhibited platelet activation induced by TXA₂ through inhibition of TXA₂-induced Ca²⁺ mobilization, and ginsenoside R_{g₃} induced TXA₂-induced platelet aggregation. Ginsenoside R_c was shown to stimulate *in vitro* PGI₂ formation by cultured rat vascular smooth muscle cells through enhanced gene expression of cyclooxygenase (Hirai, 1999). These results suggest that ginsenosides R_{g₁} and R_{g₃}, which have anti-platelet and anti-atherosclerotic effects, may have clinical potential for the prevention and the treatment of certain thrombotic and atherosclerotic disorders. Moreover, American ginseng extract was proved to be associated with the inhibition of thrombin-induced endothelin release due to NO release (Yuan et al., 1999). This result suggests that American ginseng may play a therapeutic role in facilitating the hemodynamic balance of vascular endothelial cells.

Conclusions

For a long time, ginseng has been one of the highly valued herbs in the East. Recently, many clinical trials using ginseng have been undertaken in the West as well as in the East area. Here, we have summarized diverse pharmacological and physiological effect of ginsenosids on the various diseases such as diabetes, cancer and cardiovascular diseases. However, although there have been lots of evidence suggesting that ginseng can be useful for the treatment of various diseases, we still have a long way to go. First of all, a lot of experiments reported have been performed in animal models instead of human beings. Therefore, a large-scale, controlled clinical study is needed to validate these results and apply to human beings. Secondly, there is still little evidence how ginsenosides and so on can be effective in the molecular levels. Understanding of molecular regulation of ginsenosides will be necessary to apply ginseng clinically, and also to find out new therapeutic effect of ginseng.

References

- Astrup A & Finer N (2000) Redefining type 2 diabetes 'diabesity' or 'obesity dependent diabetes mellitus'. *Obes. Rev.* 1: 57–59.
- Atopkina LN, Malinovskaya GV, Elyakov GB, Uvarova NI, Woerdenbag HJ, Koulman A, Pras N & Potier P (1999) Cytotoxicity of natural ginseng glycosides and semisynthetic analogues. *Planta Med.* 65: 30–34.
- Attele AS, Zhou YP, Xie JT, Wu JA, Zhang L, Dey L, Pugh W, Rue PA, Polonsky KS & Yuan CS (2002) Antidiabetic effects of *Panax ginseng* berry extract and the identification of an effective component. *Diabetes* 51: 1851–1858.
- Bai CX, Sunami A, Namiki T, Sawanobori T & Furukawa T (2003) Electrophysiological effects of ginseng and ginsenoside Re in guinea pig ventricular myocytes. *Eur. J. Pharmacol.* 22: 35–44.
- Bensky D & Gamble A (1993) Chinese Herbal Medicine *Materia Medica*. Eastland Press, Seattle, WA.
- Chung SH, Choi CG & Park SH (2001) Comparisons between white ginseng radix and rootlet for antidiabetic activity and mechanism in KKAY mice. *Arch. Pharm. Res.* 24: 214–218.
- Choi CH, Kang G & Min YD (2003) Reversal of P-glycoprotein-mediated multidrug resistance by protopanaxatriol ginsenosides from Korean red ginseng. *Planta Med.* 69: 235–240.
- Duda RB, Kang SS, Archer SY, Meng S & Hordin RA (2001) American ginseng transcriptionally activates p21 mRNA in breast cancer cell lines. *J. Korean Med. Sci.* 16: S54–S60.
- Duda RB, Taback B, Kessel B, Dooley DD, Yang H, Marchiori J, Slomovic BM & Alvarez JG (1996) pS2 expression induced by American ginseng in MCF-7 breast cancer cells. *Ann. Surg. Oncol.* 3: 515–520.
- Duda RB, Zhong Y, Navas V, Li MZ, Toy BR & Alavarez JG (1999) American ginseng and breast cancer therapeutic agents synergistically inhibit MCF-7 breast cancer cell growth. *J. Surg. Oncol.* 72: 230–239.
- Friedberg EC (1986) *Cancer Biology*. (pp. 138–148) W. H. Freeman and Company, New York.
- Fujikawa-Yammamoto K, Ota T, Odashima S, Abe H & Arichi S (1987) Different response in the cell cycle of tumor cells to ginsenoside Rh₂. *Cancer J.* 1: 349–352.
- Fujimoto J, Sakaguchi H, Aoki I, Toyoki H, Khatun S & Tamaya T (2001) Inhibitory effect of ginsenoside-Rb₂ on invasiveness of uterine endometrial cancer cells to the basement membrane. *Eur. J. Gynaecol. Oncol.* 22: 339–341.
- Fukushima SH, Wanibuchi H & Li W (2001) Inhibition by ginseng of colon carcinogenesis in rats. *J. Korean Med. Sci.* S75–S80.
- Gillis CN, Kim HY, Chen X & Park H (1993) Pulmonary vascular effects of ginsenosides. *Proceedings of 6th International Ginseng Symposium*, pp. 36–39.
- Han BH, Park MH, Han YN, Woo LK, Sankawa U, Yahara S & Tanaka O (1982) Degradation of ginseng saponins under mild acidic conditions. *Planta Med.* 44: 146–149.
- Hasegawa H & Benno Y (2000a) Anticarcinogenesis in mice by ginseng hydrolyzing colonic bacteria. *Microb. Ecol. Health Dis.* 12: 85–91.
- Hasegawa H, Lee KS, Nagaoka T, Tezuka Y, Uchiyama M, Kadota S & Saiki I (2000b) Pharmacokinetics of ginsenoside deglycosylated by intestinal bacteria and its transformation to biologically active fatty acid ester. *Biol. Pharm. Bull.* 23: 298–304.

- Hasegawa H & Saiki I (2000c) Oleoyl triterpene glycoside biosynthesized from ginseng suppresses growth and metastasis of murine melanoma B16-F10 tumor via immunostimulation. *J. Trad. Med.* 17: 186–193.
- Hasegawa H, Sung JH, Matsumiya S & Uchiyama M (1996) Main ginseng saponin metabolites formed by intestinal bacteria. *Planta Med.* 62: 453–457.
- Hasegawa H, Sung JH, Matsumiya S, Uchiyama M, Inouye Y, Kasai R & Yamasaki K (1995) Reversal of daunomycin and vinblastine resistance in multidrug-resistant P388 leukemia in vitro through enhanced cytotoxicity by triterpenoids. *Planta Med.* 61: 409–413.
- Hasegawa H, Suzuki R, Nagaoka T, Tezuka Y, Kadota S & Saiki I (2002) Prevention of growth and metastasis of murine melanoma through enhanced natural killer cytotoxicity by fatty acid conjugate of protopanaxatriol. *Biol. Pharm. Bull.* 25: 861–866.
- Hirai A (1999). Studies on the mechanisms of anti-platelet and anti-atherosclerotic effects of Korean red ginseng: Focusing on arachidonic acid cascade. *Proceedings of '99 Korea-Japan Ginseng Symposium*, pp. 16–31.
- Huang KC (1999) *The Pharmacology of Chinese Herbs* CRC Press, Boca Raton, FL.
- Jeon BH, Kim CS, Park KS, Lee JW, Park JB, Kim KJ, Kim SH, Chang SJ & Nam KY (2000) Effect of Korea red ginseng on the blood pressure in conscious hypertensive rats. *Gen. Pharmacol.* 35: 135–141.
- Kang SY, Schini-Kerth VB & Kim ND (1995) Ginsenosides of the protopanaxatriol group cause endothelium-dependent relaxation in the rat aorta. *Life Sci.* 56: 1577–1586.
- Karikura M, Miyase T, Tanizawa H, Taniyama T & Takino Y (1991a) Studies on absorption, distribution, excretion and metabolism of ginseng saponins. VI. The decomposition products of ginsenoside Rb₂ in the stomach of rats. *Chem. Pharm. Bull. (Tokyo)* 39: 400–404.
- Karikura M, Miyase T, Tanizawa H, Taniyama T & Takino Y (1991b) Studies on absorption, distribution, excretion and metabolism of ginseng saponins. VII. Comparison of the decomposition modes of ginsenoside-Rb₁ and -Rb₂ in the digestive tract of rats. *Chem. Pharm. Bull. (Tokyo)* 39: 2357–2361.
- Kasai R, Besso H, Tanaka O, Saruwatari YI & Mizutani T (1983) Saponins of red ginseng. *Chem. Pharm. Bull.* 31: 2120–2125.
- Kim YS, Jin SH, Lee YH, Kim SI & Park JD (1999a) Ginsenoside Rh₂ induces apoptosis independently of Bcl-2, Bcl-xL or Bax in C6Bu-1 cells. *Proceedings of '99 Korea-Japan Ginseng Symposium*, pp. 150–162.
- Kim ND, Kang SY, Kim MJ, Park JH & Schini-Kerth VB (1999b) The ginsenoside Rg₃ evokes endothelium-independent relaxation in rat aortic rings: Role of K⁺ channels. *Eur. J. Pharmacol.* 367: 51–57.
- Kim ND, Kang SY, Park JH & Schini-Kerth VB (1999c) Ginsenoside Rg₃ mediates endothelium-dependent relaxation in response to ginsenosides in rat aorta: Role of K⁺ channels. *Eur. J. Pharmacol.* 367: 41–49.
- Kim ND, Kang SY & Schini VB (1994) Ginsenosides evoke endothelium-dependent vascular relaxation in rat aorta. *Gen. Pharmacol.* 25: 1071–1077.
- Kim YS, Kim DS & Kim SI (1998) Ginsenoside Rh₂ and Rh₃ induce differentiation of HL-60 cells into granulocytes: Modulation of protein kinase C isoforms during differentiation by ginsenoside Rh₂. *Int. J. Biochem. Cell Biol.* 30: 27–338.
- Kim SW, Kwon HY, Chi DW, Shim JH, Park JD, Lee YH, Pyo S & Rhee DK (2003) Reversal of P-glycoprotein-mediated multidrug resistance by ginsenoside Rg₃. *Biochem. Pharmacol.* 65: 75–82.
- Kim HS, Lee EH, Ko SR, Choi KJ, Park JH & Im DS (2004) Effects of ginsenosides Rg₃ and Rh₂ on the proliferation of prostate cancer cells. *Arch. Pharm. Res.* 27: 429–435.
- Kim HE, Oh JH, Lee SK & Oh YJ (1999d) Ginsenoside Rh₂ induces apoptotic cell death in rat C6 glioma via a reactive oxygen- and caspase-dependent but Bcl-X(L)-independent pathway. *Life Sci.* 65: PL33–40.
- Kimura M (1980) Hypoglycemic component in ginseng radix and its insulin release. *Proceedings of the 3rd International Ginseng Symposium*. Korean Ginseng Research Institut, Seoul, Korea.
- Kimura I, Nakashima N, Sugihara Y, Fu-jun C & Kimura M, (1999) The antihyperglycaemic blend effect of traditional chinese medicine byakko-ka-ninjin-to on alloxan and diabetic KK-CA(y) mice. *Phytother. Res.* 13: 484–488.
- Kimura M & Suzuki J (1981) The pattern of action of blended Chinese traditional medicines to glucose tolerance curves in genetically diabetic KK-CAy mice. *J. Pharmacobiodyn.* 4: 907–915.
- Kimura M, Waki I, Chujo T, Kikuchi T, Hiyama C, Yamazaki K & Tanaka O (1981a) Effects of hypoglycemic components in ginseng radix on blood insulin level in alloxan diabetic mice and on insulin release from perfused rat pancreas. *J. Pharmacobiodyn.* 4: 410–417.
- Kimura M, Waki I, Tanaka O, Nagai Y & Shibata S (1981b) Pharmacological sequential trials for the fractionation of components with hypoglycemic activity in alloxan diabetic mice from ginseng radix. *J. Pharmacobiodyn.* 4: 402–409.
- Kitagawa I, Yoshikawa M, Yoshihara M, Hayashi T & Taniyama T (1983) Chemical studies on crude drug procession. I. On the constituents of Ginseng Radix Rubra. *Yakugaku Zasshi* 103: 612–622.
- Kobashi K (1998) Glycosides are natural prodrugs: Evidence using germfree and gnotobiotic rats associated with a human intestinal bacterium. *J. Trad. Med.* 15: 1–13.
- Kudo K, Tachikawa E, Kashimoto T & Takahashi E (1998) Properties of ginseng saponin inhibition of catecholamine secretion in bovine adrenal chromaffin cells. *Eur. J. Pharmacol.* 341: 139–144.
- Kumar A (1993) Chemopreventive action of ginseng on DMBA-induced papillomagenesis in the skin of mice. *Proc. 6th Intl. Ginseng Symp.* (pp. 66–68).
- Kuo SC, Teng CM, Lee JC, Ko FN, Chen SC & Huang TF (1990) Antiplatelet components in *Panax ginseng*. *Planta Med.* 56: 164–167.
- Lee KS (1980) Effect of ginseng saponin on the vascular smooth muscle. *Proceedings of 3rd International Ginseng Symposium*, pp. 71–76.
- Lee YH, Kim SI, Lee SK, Chung HY & Kim KW (1993). Differentiation mechanism of ginsenosides in cultured murine F9 teratocarcinoma stem cells. *Proceedings of 6th International Ginseng Symposium*, pp. 127–131.
- Lee KY, Park JA, Chung E, Lee YH, Kim SI & Lee SK (1996) Ginsenoside-Rh₂ blocks the cell cycle of SK-HEP-1 cells at the G1/S boundary by selectively inducing the protein expression of p27 kipl. *Cancer Lett.* 110: 193–200.
- Lee JY, Shin JW, Chun KS, Park KK, Chung WY, Bang YJ, Sung JH & Surh YJ (2005) Antitumor promotional effects of a novel intestinal bacterial metabolite (IH-901) derived from

- the protopanaxadiol-type ginsenosides in mouse skin. *Carcinogenesis* 26: 359–367.
- Mochizuki M, Yoo YC, Matsuzawa K, Sato K, Saiki I, Tonooka S, Samikawa K & Azuma I (1995) Inhibitory effect of tumor metastasis in mice by saponins, ginsenoside-Rb₂, 20 (R)- and 20 (S)-ginsenoside-Rg₃, of red ginseng. *Biol. Pharm. Bull.* 18: 1197–1202.
- Nakata H, Kikuchi Y, Tode T, Hirata J, Kita T, Ishii K, Kudoh K, Nagata I & Shinomiya N (1998) Inhibitory effects of ginsenoside Rh₂ on tumor growth in nude mice bearing human ovarian cancer cells. *Jpn. J. Cancer Res.* 89: 733–740.
- Odani T, Tanizawa H & Takino Y (1983) Studies on the absorption, distribution, excretion and metabolism of ginseng saponins. III. The absorption, distribution and excretion of ginsenoside Rb₁ in the rat. *Chem. Pharm. Bull.* (Tokyo) 31: 1059–1066.
- Odashima S, Ohta T, Kohno H, Matsuda T, Kitagawa I, Abe H & Arichi S (1985) Control of phenotypic expression of cultured B16 melanoma cells by plant glycosides. *Cancer Res.* 45: 2781–2784.
- Odashima S, Ota T, Fujikawa-Yamamoto K & Abe H (1989) of phenotypic reverse transformation by plant glycosides in cultured cancer. *Gan To Kagaku Ryoho* 16: 1483–1489.
- Palmer RM, Ashton DS & Moncada S (1988) Vascular endothelial cells synthesize nitric oxide from L-arginine. *Nature* 333: 664–666.
- Park JD (1996) Recent studies on the chemical constituents of Korean ginseng (*Panax ginseng* CA Meyer). *Korean J. Ginseng Sci.* 20: 389–415.
- Park JD, Baek NI, Lee YH, Kim DS, Park JH, Ryu JH & Kim SI (1998) Four new dammarane-glycosides, ginsenosides Rg₅, Rh₄, Rs₃ and Rf₂ from Korean red ginseng, the root of *Panax ginseng* C. A. Meyer. *Proceedings of the 7th International Symposium on ginseng*, pp. 115–126.
- Park MT, Cha HJ, Jeong JW, Kim SI, Chung HY, Kim ND, Kim OH & Kim KW (1999) Glucocorticoid receptor-induced down-regulation of MMP-9 by ginseng components, PD and PT contributes to inhibition of the invasive capacity of HT1080 human fibrosarcoma cells. *Mol. Cells* 9: 476–483.
- Park JD, Kim DS, Kwon HY, Son SK, Lee YH, Baek NI, Kim SI & Rhee DK (1996) Effects of ginseng saponin on modulation of multidrug resistance. *Arch. Pharm. Res.* 19: 213–218.
- Popovich DG & Kitts DD (2002) Structure–function relationship exists for ginsenosides in reducing cell proliferation and inducing apoptosis in the human leukemia (THP-1) cell line. *Arch. Biochem. Biophys.* 1:(406), 1–8.
- Popovich DG & Kitts DD (2004) Ginsenosides 20(S)-protopanaxadiol and Rh₂ reduce cell proliferation and increase sub-G1 cells in two cultured intestinal cell lines, Int-407 and Caco-2. *Can. J. Physiol. Pharmacol.* 82: 183–190.
- Rapoport RM & Murad F (1983) Agonist-induced endothelium-dependent relaxation in rat thoracic aorta may be mediated through cGMP. *Circ. Res.* 52: 352–357.
- Sato K, Mochizuki M, Saiki I, Yoo YC, Samukawa K & Azuma I (1994) Inhibition of tumor angiogenesis and metastasis by a saponin of *Panax ginseng*, ginsenoside-Rb₂. *Biol. Pharm. Bull.* 17: 635–639.
- Scott GI, Colligan PB, Ren BH & Ren J (2001) Ginsenosides Rb₁ and Re decrease cardiac contraction in adult rat ventricular myocytes: Role of nitric oxide. *Br. J. Pharmacol* 134: 1159–1165.
- Shibata S (2001) Chemistry and cancer preventing activities of ginseng saponins and some related triterpenoid compounds. *J. Korean Med.* 16: S28–37.
- Shinkai K, Akedo H, Mukai M, Imamura F, Isoai A, Kobayashi M & Kitagawa I (1996) Inhibition of in vitro tumor cell invasion by ginsenoside Rg₃. *Jpn. J. Cancer Res.* 87: 357–362.
- Shibata S, Fujita M, Itokawa H, Tanaka O & Ishii T (1963a) Studies on the constituents of Japanese and Chinese crude drugs. XI. Panaxadiol. A sapogenin of ginseng roots. (1). *Chem. Pharm. Bull.* 11: 759–761.
- Shibata S, Tanaka O, Nagai M & Ishii T (1963b) Studies on the constituents of Japanese and Chinese crude drugs. XII. Panaxadiol. A sapogenin of ginseng roots (2). *Chem. Pharm. Bull.* 11: 762–767.
- Shibata S, Tanaka O, Shoji J & Saito H (1995) Chemistry and pharmacology of *Panax*. In: *Economic and Medicinal Plant Research*. Vol. 1. (pp. 217–284). Academic Press, New York.
- Sonnenborn U & Proppert Y (1990) Ginseng (*Panax ginseng* C.A Meyer). *Zeitschrift fur phytotherapie* 11: 35–49.
- Suda K, Murakami K, Murata J, Hasegawa H & Saiki I (2000) Induction of apoptosis in Lewis lung carcinoma cells by an intestinal bacterial metabolite resulted from orally administered ginseng protopanaxadiol saponins. *J. Trad. Med.* 17: 236–244.
- Sung J, Han KH, Zo JH, Park HJ, Kim CH & Oh BH (2000) Effects of red ginseng upon vascular endothelial function in patients with essential hypertension. *Am. J. Chin. Med.* 28: 205–216.
- Tatsuka M, Maeda M & Ota T (2001) Anticarcinogenic effect and enhancement of metastatic potential of BALB/c 3T3 cells by ginsenoside Rh₂. *Jpn. J. Cancer Res.* 92: 1184–1189.
- Teng CM, Kuo SC, Ko FN, Lee JC, Lee LG, Chen SC & Huang TF (1989) Antiplatelet actions of panaxynol and ginsenosides isolated from ginseng. *Biochim. Biophys. Acta.* 990: 315–320.
- Thompson CB (1995) Apoptosis in the pathogenesis and treatment of disease. *Science* 267: 1456–1462.
- Tode T, Kikuchi Y, Hirata J, Kita T, Imaizumi E & Nagata I (1993) Inhibitory effects of oral administration of ginsenoside Rh₂ on tumor growth in nude mice bearing serious cyst adenocarcinoma of the human ovary. *Nippon Sanka Fujinka Gakkai Zasshi* 45: 1275–1282.
- Vane JR, Anggard EE & Botting RM (1990) Regulatory functions of the vascular endothelium. *N. Engl. J. Med.* 323: 27–36.
- Vuksan V, Sievenpiper JL, Koo VY, Francis T, Beljan-Zdravkovic U, Xu Z & Vidgen E (2000) American ginseng (*Panax quinquefolius* L.) reduces postprandial glycemia in nondiabetic subjects and subjects with type 2 diabetes mellitus. *Arch. Intern. Med.* 160: 1009–1013.
- Wakabayashi C, Murakami K, Hasegawa H, Murata J & Saiki I (1998) An intestinal bacterial metabolite of ginseng protopanaxadiol saponins has the ability to induce apoptosis in tumor cells. *Biochem. Biophys. Res. Commun.* 246: 725–730.
- Wu XG, Zhu DH & Li X (2001) Anticarcinogenic effect of red ginseng on the development of liver cancer induced by diethylnitrosamine in rats. *J. Korean Med.* 16: S61–S65.
- Xia LJ & Han R (1996) Differentiation of B16 melanoma cells induced by ginsenoside Rh₂. *Yao Xue Xue Bao* 31: 742–745.

- Yammamoto M (1992) Effects of administration of red ginseng on hypertension, normotension and hypotension *Ginseng Rev.* 9: 15–20.
- Yokozawa T, Kobayashi T, Oura H & Kawashima Y (1985) Studies on the mechanism of the hypoglycemic activity of ginsenoside-Rb₂ in streptozotocin-diabetic rats. *Chem Pharm. Bull. (Tokyo)* 33: 869–72.
- Yuan CS, Attele AS, Wu JA, Lowell TK, Gu Z & Lin Y (1999) *Panax quinquefolium* L. inhibits thrombin- induced endothelin release in vitro. *Am. J. Chin. Med.* 27: 331–338.
- Yun TK (2003) Experimental and epidemiological evidence on non-organ specific cancer preventive effect of Korean ginseng and identification of active compounds *Mutat. Res.* 523–524: 63–74.
- Yun TK, Lee YS, Lee YH, Kim SI & Yun HY (2001) Anticarcinogenic effect of *Panax ginseng* C. A. Meyer and identification of active compounds. *J. Korean Med.* 16:(Suppl), S6–S18.
- Yun TK, Yun TS & Han IW (1983) Anticarcinogenic effect of long-term oral administration of red ginseng on new born mice exposed to various chemical carcinogens. *Cancer Detect. Prev.* 6: 515–525.
- Zhang WJ, Wojta J & Binder BR (1997) Effect of notoginsenoside R1 on the synthesis of components of the fibrinolytic system in cultured smooth muscle cells of human pulmonary artery. *Cell Mol. Biol.* 43: 581–587.
- Zhou W, Chai H, Lin PH, Lumsden AB, Yao Q & Chen CJ (2004) Molecular mechanisms and clinical applications of ginseng root for cardiovascular disease. *Med. Sci. Monit.* 10: RA187–RA192.